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<p>(54) Title: RG NUCLEIC ACIDS FOR CONFERRING DISEASE RESISTANCE TO PLANTS (57) Abstract The present invention provides RG nucleic acids and proteins which confer disease resistance to plants. The nucleic acids can be used to produce transgenic plants resistant to pests. Antibodies to proteins of the invention are also provided.</p>		

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RG NUCLEIC ACIDS FOR CONFERRING DISEASE RESISTANCE TO PLANTS

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The present application is a continuation-in-part application ("CIP") of U.S. Patent Application Serial No. ("USSN") 08/781,734, filed January 10, 1997. The
10 aforementioned application is explicitly incorporated herein by reference in its entirety and for all purposes.

This invention was made with Government support under Grant Nos. 92-37300-7547 and 95-37300-1571, awarded by the United States Department of Agriculture.
15 The Government has certain rights in this invention.

FIELD OF THE INVENTION

The present invention relates generally to plant molecular biology. In particular, it relates to nucleic acids and methods for conferring pest resistance in plants.
20 particularly lettuce.

BACKGROUND OF THE INVENTION

Recently, several resistance genes have been cloned by several groups from several plants. Many of these genes are sequence related. The derived amino acid
25 sequences of the most common class, *RPS2*, *RPM1* (bacterial resistances in *Arabidopsis* (Mindrinos *et al.* *Cell* 78:1089-1099 (1994)); Bent *et al.* *Science* 265:1856-1860 (1994); Grant *et al.*, *Science* 269:843-846 (1995)), *L6* (fungal resistance in flax; Lawrence, *et al.*, *The Plant Cell* 7:1195-1206 (1995)), and *N*, (virus resistance in tobacco; Whitham, *et al.*, *Cell* 78:1101-1115 (1994); and U.S. Patent No. 5,571,706), all contain leucine-rich
30 repeats (LRR) and nucleotide binding sites (NBS).

The NBS is a common motif in several mammalian gene families encoding signal transduction components (e.g., *Ras*) and is associated with ATP/GTP-binding sites.

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LRR domains can mediate protein-protein interactions and are found in a variety of proteins involved in signal transduction, cell adhesion and various other functions. LRRs are leucine rich regions often comprising 20-30 amino acid repeats where leucine and other aliphatic residues occur periodically. LRRs can function extracellularly or intracellularly.

Since the onset of civilization, plant diseases have had catastrophic effects on crops and the well-being of the human population. Plant diseases continue to effect enormous human and economic costs. An increasing human population and decreasing amounts of arable land make all approaches to preventing and treating plant pathogen destruction critical. The ability to control and enhance a plant's protective responses against pathogens would be of enormous benefit. Tissue-specific and temporal control of mechanisms responsible for plant cell death would also be of great practical and economic value. The present invention fulfills these and other needs.

What is needed in the art are plant disease resistance genes and means to create transgenic disease resistance plants, particularly in lettuce. Further, what is needed in the art is a means to DNA fingerprint cultivars and germplasm with respect to their disease resistance haplotypes for use in plant breeding programs. The present invention provides these and other advantages.

SUMMARY OF THE INVENTION

The present invention provides isolated nucleic acid constructs. These constructs comprise an RG (resistance gene) polynucleotide which encodes an RG polypeptide having at least 60% sequence identity to an RG polypeptide selected from the group consisting of: an RG1 polypeptide, an RG2 polypeptide, an RG3 polypeptide, and an RG4 polypeptide. RG1, RG2, RG3, RG4, and the like, represent individual "RG families." Each "RG family," as defined herein, is a group of polypeptide sequences that have at least 60% amino acid sequence identity. Individual members of an RG family, *i.e.*, individual species of the genus, typically map to the same genomic locus. The invention provides for constructs comprising nucleotides encoding the RG families of the

invention, which can include sequences encoding a leucine rich region (LRR), and/or a nucleotide binding site (NBS), or both.

The invention provides for an isolated nucleic acid construct comprising an RG polynucleotide which encodes an RG polypeptide having at least 60% sequence identity to an RG polypeptide from an RG family selected from the group consisting of: an RG1 polypeptide, an RG2 polypeptide, an RG3 polypeptide, an RG4 polypeptide, an RG5 polypeptide, and an RG7 polypeptide. In alternative embodiments, the nucleic acid construct comprises an RG polynucleotide which encodes an RG polypeptide comprising an leucine rich region (LRR), or, an RG polypeptide comprising a nucleotide binding site (NBS). The nucleic acid construct can comprise a polynucleotide which is a full length gene. In another embodiment, the nucleic acid construct encodes a fusion protein.

In one embodiment, the nucleic acid construct comprises a sequence encoding an RG1 polypeptide. The RG1 polypeptide can be encoded by a polynucleotide sequence selected from the group consisting of SEQ ID NO:1 (RG1A), SEQ ID NO:2 and SEQ ID NO:137 (RG1B), SEQ ID NO: 3 (RG1C), SEQ ID NO:4 (RG1D), SEQ ID NO:5 (RG1E), SEQ ID NO:6 (RG1F), SEQ ID NO:7 (RG1G), SEQ ID NO:8 (RG1H), SEQ ID NO:9 (RG1I), and SEQ ID NO:10 (RG1J).

In another embodiment, the nucleic acid construct comprises a sequence encoding an RG2 polypeptide. The RG2 polypeptide can be encoded by a polynucleotide sequence selected from the group consisting of: SEQ ID NO:21 and SEQ ID NO:27 (RG2A); SEQ ID NO:23 and SEQ ID NO:28 (RG2B); SEQ ID NO:29 (RG2C); SEQ ID NO:30 (RG2D); SEQ ID NO:31 (RG2E); SEQ ID NO:32 (RG2F); SEQ ID NO:33 (RG2G); SEQ ID NO:34 (RG2H); SEQ ID NO:35 (RG2I); SEQ ID NO:36 (RG2J); SEQ ID NO:37 (RG2K); SEQ ID NO:38 (RG2L); SEQ ID NO:39 (RG2M); SEQ ID NO:87 (RG2A); SEQ ID NO:89 (RG2B); SEQ ID NO:91 (RG2C); SEQ ID NO:93 (RG2D) and SEQ ID NO:94 (RG2D); SEQ ID NO:96 (RG2E); SEQ ID NO:98 (RG2F); SEQ ID NO:100 (RG2G); SEQ ID NO:102 (RG2H); SEQ ID NO:104 (RG2I); SEQ ID NO:106 (RG2J) and SEQ ID NO:107 (RG2J); SEQ ID NO:109 (RG2K) and (SEQ ID NO:110 (RG2K); SEQ ID NO:112 (RG2L); SEQ ID NO:114 (RG2M); SEQ ID NO:116 (RG2N); SEQ ID NO:118 (RG2O); SEQ ID NO:120 (RG2P); SEQ ID NO:122 (RG2Q); SEQ ID NO:124 (RG2S); SEQ ID NO:126 (RG2T); SEQ ID NO:128 (RG2U); SEQ ID NO:130 (RG2V); and, SEQ ID NO:132 (RG2W).

In other embodiments, the nucleic acid construct comprises a RG3 sequence (SEQ ID NO:68) encoding an RG3 polypeptide (SEQ ID NO:138) (RG3). In other embodiments, the nucleic acid construct comprises an RG4 sequence (SEQ ID NO:69) encoding an RG4 polypeptide (SEQ ID NO:139) (RG4).

5 In other embodiments, the nucleic acid construct comprises a RG5 sequence (SEQ ID NO:134) encoding an RG5 polypeptide (SEQ ID NO:135). The RG5 polypeptide can be encoded by a polynucleotide sequence as set forth in SEQ ID NO:134.

The invention also provides for a nucleic acid construct which comprises an RG7 sequence encoding an RG7 polypeptide. The RG7 polypeptide can be encoded by a polynucleotide sequence as set forth in SEQ ID NO:136.

10 In further embodiments, the nucleic acid construct can further comprise a promoter operably linked to the RG polynucleotide. In alternative embodiments, the promoter can be a plant promoter; a disease resistance promoter; a lettuce promoter; a constitutive promoter; an inducible promoter; or, a tissue-specific promoter. The nucleic acid construct can comprise a promoter sequence from an RG gene linked to a heterologous polynucleotide.

The invention also provides for a transgenic plant comprising a recombinant expression cassette comprising a promoter operably linked to an RG polynucleotide. The expression cassette can comprise a plant promoter or a viral promoter; the plant promoter can be a heterologous promoter. In one embodiment, the transgenic plant is lettuce. In alternative embodiments, the transgenic plant comprises an expression cassette which includes an RG polynucleotide selected from the group consisting of SEQ ID NO:1 (RG1A); SEQ ID NO:2 and SEQ ID NO:137 (RG1B); SEQ ID NO:3 (RG1C); SEQ ID NO:4 (RG1D); SEQ ID NO:5 (RG1E); SEQ ID NO:6 (RG1F); SEQ ID NO:7 (RG1G); SEQ ID NO:8 (RG1H); SEQ ID NO:9 (RG1I) and SEQ ID NO:10 (RG1J); SEQ ID NO:21 and SEQ ID NO:27 (RG2A); SEQ ID NO:23 and SEQ ID NO:28 (RG2B); SEQ ID NO:29 (RG2C); SEQ ID NO:30 (RG2D); SEQ ID NO:31 (RG2E); SEQ ID NO:32 (RG2F); SEQ ID NO:33 (RG2G); SEQ ID NO:34 (RG2H); SEQ ID NO:35 (RG2I); SEQ ID NO:36 (RG2J); SEQ ID NO:37 (RG2K); SEQ ID NO:38 (RG2L); SEQ ID NO:39 (RG2M); SEQ ID NO:87 (RG2A); SEQ ID NO:89 (RG2B); SEQ ID NO:91 (RG2C); SEQ ID NO:93 (RG2D) and SEQ ID NO:94 (RG2D); SEQ ID NO:96 (RG2E); SEQ ID NO:98 (RG2F); SEQ ID NO:100 (RG2G); SEQ ID NO:102 (RG2H); SEQ ID NO:104

(RG2I); SEQ ID NO:106 (RG2J) and SEQ ID NO:107 (RG2J); SEQ ID NO:109 (RG2K) and (SEQ ID NO:110 (RG2K); SEQ ID NO:112 (RG2L); SEQ ID NO:114 (RG2M); SEQ ID NO:116 (RG2N); SEQ ID NO:118 (RG2O); SEQ ID NO:120 (RG2P); SEQ ID NO:122 (RG2Q); SEQ ID NO:124 (RG2S); SEQ ID NO:126 (RG2T); SEQ ID NO:128 (RG2U); SEQ ID NO:130 (RG2V); and, SEQ ID NO:132 (RG2W); SEQ ID NO:68 (RG3); SEQ ID NO:69 (RG4); SEQ ID NO:134 (RG5); or SEQ ID NO:136 (RG7).

The invention provide for a transgenic plant comprising an expression cassette comprising an RG polynucleotide which can encode an RG1 polypeptide selected from the group consisting of SEQ ID NO:11 (RG1A), SEQ ID NO:12 (RG1B), SEQ ID NO:13 (RG1C), SEQ ID NO:14 (RG1D), SEQ ID NO:15 (RG1E), SEQ ID NO:16 (RG1F), SEQ ID NO:17 (RG1G), SEQ ID NO:18 (RG1H), SEQ ID NO:19 (RG1I), or SEQ ID NO:20 (RG1J); or, an RG2 polypeptide selected from the group consisting of SEQ ID NO:22 and SEQ ID NO:41 (RG2A); SEQ ID NO:24 and SEQ ID NO:42 (RG2B); SEQ ID NO:43 (RG2C); SEQ ID NO:44 (RG2D); SEQ ID NO:45 (RG2E); SEQ ID NO:46 (RG2F); SEQ ID NO:47 (RG2G); SEQ ID NO:48 (RG2H); SEQ ID NO:49 (RG2I); SEQ ID NO:50 (RG2J); SEQ ID NO:51 (RG2K); SEQ ID NO:52 (RG2L); SEQ ID NO:53 (RG2M); SEQ ID NO:88 (RG2A); SEQ ID NO:90 (RG2B); SEQ ID NO:92 (RG2C); SEQ ID NO:95 (RG2D); SEQ ID NO:97 (RG2E); SEQ ID NO:99 (RG2F); SEQ ID NO:101 (RG2G); SEQ ID NO:103 (RG2H); SEQ ID NO:105 (RG2I); SEQ ID NO:108 (RG2J); SEQ ID NO:111 (RG2K); SEQ ID NO:113 (RG2L); SEQ ID NO:115 (RG2M); SEQ ID NO:117 (RG2N); SEQ ID NO:119 (RG2O); SEQ ID NO:121 (RG2P); SEQ ID NO:123 (RG2Q); SEQ ID NO:125 (RG2S); SEQ ID NO:127 (RG2T); SEQ ID NO:129 (RG2U); SEQ ID NO:131 (RG2V); and, SEQ ID NO:133 (RG2W); an RG4 polypeptide as set forth by SEQ ID NO:72; an RG5 polypeptide with a sequence as set forth by SEQ ID NO:135; or, an RG7 polypeptide.

The invention also provides for a method of enhancing disease resistance in a plant, the method comprising introducing into the plant a recombinant expression cassette comprising a promoter functional in the plant and operably linked to an RG polynucleotide sequence. In this method, the plant can be a lettuce plant; and, the RG polynucleotide can encode an RG polypeptide selected from the group consisting of an RG1 polypeptide selected from the group consisting of SEQ ID NO:11 (RG1A), SEQ ID NO:12 (RG1B), SEQ ID NO:13 (RG1C), SEQ ID NO:14 (RG1D), SEQ ID NO:15 (RG1E), SEQ ID

NO:16 (RG1F), SEQ ID NO:17 (RG1G), SEQ ID NO:18 (RG1H), SEQ ID NO:19 (RG1I), or SEQ ID NO:20 (RG1J); or, an RG2 polypeptide selected from the group consisting of SEQ ID NO:22 and SEQ ID NO:41 (RG2A); SEQ ID NO:24 and SEQ ID NO:42 (RG2B); SEQ ID NO:43 (RG2C); SEQ ID NO:44 (RG2D); SEQ ID NO:45 (RG2E); SEQ ID NO:46 (RG2F); SEQ ID NO:47 (RG2G); SEQ ID NO:48 (RG2H); SEQ ID NO:49 (RG2I); SEQ ID NO:50 (RG2J); SEQ ID NO:51 (RG2K); SEQ ID NO:52 (RG2L); SEQ ID NO:53 (RG2M); SEQ ID NO:72; SEQ ID NO:74; SEQ ID NO:88 (RG2A); SEQ ID NO:90 (RG2B); SEQ ID NO:92 (RG2C); SEQ ID NO:95 (RG2D); SEQ ID NO:97 (RG2E); SEQ ID NO:99 (RG2F); SEQ ID NO:101 (RG2G); SEQ ID NO:103 (RG2H); SEQ ID NO:105 (RG2I); SEQ ID NO:108 (RG2J); SEQ ID NO:111 (RG2K); SEQ ID NO:113 (RG2L); SEQ ID NO:115 (RG2M); SEQ ID NO:117 (RG2N); SEQ ID NO:119 (RG2O); SEQ ID NO:121 (RG2P); SEQ ID NO:123 (RG2Q); SEQ ID NO:125 (RG2S); SEQ ID NO:127 (RG2T); SEQ ID NO:129 (RG2U); SEQ ID NO:131 (RG2V); and, SEQ ID NO:133 (RG2W). In this method, the promoter can be a plant disease resistance promoter, a tissue-specific promoter, a constitutive promoter, or an inducible promoter.

The invention also provides for a method of detecting RG resistance genes in a nucleic acid sample, the method comprising: contacting the nucleic acid sample with an RG polynucleotide to form a hybridization complex; and, wherein the formation of the hybridization complex is used to detect the RG resistance gene in the nucleic acid sample. In this method, the RG polynucleotide can be an RG1 polynucleotide, an RG2 polynucleotide, an RG3 polynucleotide, an RG4 polynucleotide, an RG5 polynucleotide or an RG7 polynucleotide. In this method, the RG resistance gene can be amplified prior to the step of contacting the nucleic acid sample with the RG polynucleotide, and, the RG resistance gene can be amplified by the polymerase chain reaction. In one embodiment, the RG polynucleotide is labeled.

The invention further provides for an RG polypeptide having at least 60% sequence identity to a polypeptide selected from the group consisting of: an RG1 polypeptide, an RG2 polypeptide, an RG3 polypeptide, an RG4 polypeptide, an RG5 polypeptide, and an RG7 polypeptide.

A further understanding of the nature and advantages of the present invention may be realized by reference to the remaining portions of the specification, the figures and claims.

All publications, patents and patent applications cited herein are hereby
5 expressly incorporated by reference for all purposes.

DETAILED DESCRIPTION OF THE INVENTION

This invention relates to families of RG genes, particularly from *Lactuca sativa*. Nucleic acid sequences of the present invention can be used to confer resistance in
10 plants to a variety of pests including viruses, fungi, nematodes, insects, and bacteria. Sequences from within the RG genes can be used to fingerprint cultivars or germplasm for the presence of desired resistance genes. Promoters of RG genes can be used to drive heterologous gene expression under conditions in which RG genes are expressed. Further, the present invention provides RG proteins and antibodies specifically reactive to RG
15 proteins. Antibodies to RG proteins can be used to detect the type and amount of RG protein expressed in a plant sample.

The present invention has use over a broad range of types of plants, including species from the genera *Cucurbita*, *Rosa*, *Vitis*, *Juglans*, *Fragaria*, *Lotus*, *Medicago*, *Onobrychis*, *Trifolium*, *Trigonella*, *Vigna*, *Citrus*, *Linum*, *Geranium*, *Manihot*,
20 *Daucus*, *Arabidopsis*, *Brassica*, *Raphanus*, *Sinapis*, *Atropa*, *Capsicum*, *Datura*, *Hyoscyamus*, *Lycopersicon*, *Nicotiana*, *Solanum*, *Petunia*, *Digitalis*, *Majorana*, *Ciahorium*, *Helianthus*, *Lactuca*, *Bromus*, *Asparagus*, *Antirrhinum*, *Heterocallis*, *Nemesis*, *Pelargonium*, *Panieum*, *Pennisetum*, *Ranunculus*, *Senecio*, *Salpiglossis*, *Cucumis*, *Browaalia*, *Glycine*, *Pisum*, *Phaseolus*, *Lolium*, *Oryza*, *Zea*, *Avena*, *Hordeum*, *Secale*,
25 *Triticum*, and, *Sorghum*. In particularly preferred embodiments, species from the family *Compositae* and in particular the genus *Lactuca* are employed such as *L. sativa* and such subspecies as *crispa*, *longifolia*, and *asparagina*.

The nucleic acids of the present invention can be used in marker-aided selection. Marker-aided selection does not require the complete sequence of the gene or
30 precise knowledge of which sequence confers which specificity. Instead, partial sequences can be used as hybridization probes or as the basis for oligonucleotide primers to amplify nucleic acid, e.g., by PCR. Partial sequences can be used in other methods, such as to

follow the segregation of chromosome segments containing resistance genes in plants. Because the RG marker is the gene itself, there can be negligible recombination between the marker and the resistance phenotype. Thus, RG polynucleotides of the present invention provide an optimal means to DNA fingerprint cultivars and wild germplasm with respect to their disease resistance haplotypes. This can be used to indicate which germplasm accessions and cultivars carry the same resistance genes. At present, selection of plants (e.g., lettuce) for resistance to some diseases is slow and difficult. But linked markers allow indirect selection for such resistance genes. Moreover, RG markers also allow resistance genes to be identified and combined in a manner that would not otherwise be possible. Numerous accessions have been identified that provide resistance to all isolates of downy mildew (*Bremia lactucae*). However, without molecular markers it is impossible to combine such resistances from different sources. The nucleic acid sequences of the invention provide for a fast and convenient means to identify and combine resistances from different sources. The RG markers of the invention can also be used to identify recombinants that have new combinations of resistance genes in *cis* on the same chromosome.

In addition, RG markers may allow the identification of the Mendelian factors determining traits, such as field resistance to downy mildew. Once such markers have been identified, they will greatly increase the ease with which field resistance can be transferred between lines and combined with other resistances.

In another application, primers to RG sequences can be also designed to amplify sequences that are conserved in multiple RG family members. This gives genetic information on multiple RG family members. Alternatively, one or more primers can be made to sequences unique to a single resistance gene genus or a single RG specie. This allows an analysis of individual family groups (an RG genus) or an individual family member (a specie). Primers made to individual RGs at the edge of each cluster can be used to select for recombinants within the cluster. This minimizes the amount of linkage drag during introgression. Classical and molecular genetics has shown that pest resistance genes tend to be clustered in the genome. Pest resistance loci comprise arrays of genes and exhibit a variety of complex haplotypes rather than being simple alternate allelic forms. Pest resistance is conferred by families, or genuses, of related RG sequences, individual members, or species, of which have evolved to have a different specificity.

Oligonucleotide primers can be designed that amplify members from multiple haplotypes, or genuses, or amplify only members of one genus, or only amplify an individual specie. This will provide codominant information and allow heterozygotes to be distinguished from homozygotes.

5 Further, comparison of RG sequences will allow a determination of which sequences are critical for resistance and will ultimately lead to engineering resistance genes with new specificities. Resistance gene sequences were not previously available for lettuce. Marker-aided selection will greatly increase the precision and speed of breeding for disease resistance. Transgenic approaches will allow pyramiding of resistance genes
10 into a single Mendelian unit, transfer between sexually-incompatible species, substitute for conventional backcrossing procedures, and allow expression of other genes in parallel with resistance genes.

 The RG polynucleotides also have utility in the construction of disease resistant transgenic plants. This avoids lengthy and sometimes difficult backcrossing
15 programs currently necessary for introgression of resistance. It is also possible to transfer resistance polynucleotides between sexually-incompatible species, thereby greatly increasing the germplasm pool that can be used as a source of resistance genes. Cloning of multiple RG sequences in a single cassette will allow pyramiding of genes for resistance against multiple isolates of a single pathogen such as downy mildew or against multiple
20 pathogens. Once introduced, such a cassette can be manipulated by classical breeding methods as a single Mendelian unit.

 Transgenic plants of the present invention can also be constructed using an RG promoter. The promoter sequences from RG sequences of the invention can be used with RG genes or heterologous genes. Thus, RG promoters can be used to express a
25 variety of genes in the same temporal and spatial patterns and at similar levels to resistance genes.

Nucleic acids of the Invention and Their Preparation

RG Polynucleotide Families

30 The present invention provides isolated nucleic acid constructs which comprise an RG polynucleotide. In alternative embodiments, the RG polynucleotide is at least 18 nucleotides in length, typically at least 20, 25, or 30 nucleotides in length, more

typically at least 100 nucleotides in length, generally at least 200 nucleotides in length, preferably at least 300 nucleotides in length, more preferably at least 400 nucleotides in length, and most preferably at least 500 nucleotides in length.

In particularly preferred embodiments, the RG polynucleotide encodes a RG protein which confers resistance to plant pests. This RG protein can be longer, equivalent, or shorter than the RG protein encoded by an RG gene. In various embodiments, an RG polynucleotide can hybridize under stringent conditions to members of an RG family (an RG genus); *e.g.*, it can hybridize to a member of the RG1 RG family, such as an RG1 polynucleotide selected from the group consisting of: SEQ ID NO:1 (RG1A); SEQ ID NO:2 and SEQ ID NO:137 (RG1B); SEQ ID NO: 3 (RG1C); SEQ ID NO:4 (RG1D); SEQ ID NO:5 (RG1E); SEQ ID NO:6 (RG1F); SEQ ID NO:7 (RG1G); SEQ ID NO:8 (RG1H); SEQ ID NO:9 (RG1I) and SEQ ID NO:10 (RG1J).

In other embodiments, the polynucleotide can also hybridize under stringent conditions to a member of the RG2 family; such as an RG2 polynucleotide selected from the group consisting of: SEQ ID NO:21 and SEQ ID NO:27 (RG2A); SEQ ID NO:23 and SEQ ID NO:28 (RG2B); SEQ ID NO:29 (RG2C); SEQ ID NO:30 (RG2D); SEQ ID NO:31 (RG2E); SEQ ID NO:32 (RG2F); SEQ ID NO:33 (RG2G); SEQ ID NO:34 (RG2H); SEQ ID NO:35 (RG2I); SEQ ID NO:36 (RG2J); SEQ ID NO:37 (RG2K); SEQ ID NO:38 (RG2L); SEQ ID NO:39 (RG2M); SEQ ID NO:87 (RG2A); SEQ ID NO:89 (RG2B); SEQ ID NO:91 (RG2C); SEQ ID NO:93 (RG2D) and SEQ ID NO:94 (RG2D); SEQ ID NO:96 (RG2E); SEQ ID NO:98 (RG2F); SEQ ID NO:100 (RG2G); SEQ ID NO:102 (RG2H); SEQ ID NO:104 (RG2I); SEQ ID NO:106 (RG2J) and SEQ ID NO:107 (RG2J); SEQ ID NO:109 (RG2K) and (SEQ ID NO:110 (RG2K); SEQ ID NO:112 (RG2L); SEQ ID NO:114 (RG2M); SEQ ID NO:116 (RG2N); SEQ ID NO:118 (RG2O); SEQ ID NO:120 (RG2P); SEQ ID NO:122 (RG2Q); SEQ ID NO:124 (RG2S); SEQ ID NO:126 (RG2T); SEQ ID NO:128 (RG2U); SEQ ID NO:130 (RG2V); and, SEQ ID NO:132 (RG2W).

In alternative embodiments, each RG2 gene can also include an AC15 sequence which hybridizes under stringent conditions to a polynucleotide selected from the group consisting of: SEQ ID NO:56 (AC15-2A); SEQ ID NO:57 (AC15-2B); SEQ ID NO:58 (AC15-2C); SEQ ID NO:59 (AC15-2D); SEQ ID NO:60 (AC15-2E); SEQ ID NO:61 (AC15-2G); SEQ ID NO:62 (AC15-2H); SEQ ID NO:63 (AC15-2I); SEQ ID

NO:64 (AC15-2J); SEQ ID NO:65 (AC15-2L); SEQ ID NO:66 (AC15-2N); SEQ ID NO:67 (AC15-2O).

In other embodiments, an RG polynucleotide can hybridize under stringent conditions to an RG3 (SEQ ID NO:68), an RG4 (SEQ ID NO:69), and RG5 (SEQ ID NO:135), and an RG7 (SEQ ID NO:137), RG family member.

The present invention further provides nucleic acid constructs which comprise an RG polynucleotide which encodes RG polypeptides from various RG families; such as an RG polypeptide having at least 60% sequence identity to an RG polypeptide selected from the group consisting of: an RG1 polypeptide, an RG2 polypeptide, an RG3 polypeptide, and RG4 polypeptide, and RG5 polypeptide, and an RG7 polypeptide.

Exemplary RG1 polypeptides have the sequences shown in SEQ ID NO:2 (RG1A), SEQ ID NO:4 (RG1B), SEQ ID NO:6 (RG1C), SEQ ID NO:8 (RG1D), SEQ ID NO:10 (RG1E), SEQ ID NO:12 (RG1F), SEQ ID NO:14 (RG1G), SEQ ID NO:16 (RG1H), SEQ ID NO:20 (RG1J). Exemplary RG2 polypeptides have the sequences shown in SEQ ID NO:22 and SEQ ID NO:41 (RG2A); SEQ ID NO:24 and SEQ ID NO:42 (RG2B); SEQ ID NO:43 (RG2C); SEQ ID NO:44 (RG2D); SEQ ID NO:45 (RG2E); SEQ ID NO:46 (RG2F); SEQ ID NO:47 (RG2G); SEQ ID NO:48 (RG2H); SEQ ID NO:49 (RG2I); SEQ ID NO:50 (RG2J); SEQ ID NO:51 (RG2K); SEQ ID NO:52 (RG2L); SEQ ID NO:53 (RG2M); SEQ ID NO:88 (RG2A); SEQ ID NO:90 (RG2B); SEQ ID NO:92 (RG2C); SEQ ID NO:95 (RG2D); SEQ ID NO:97 (RG2E); SEQ ID NO:99 (RG2F); SEQ ID NO:101 (RG2G); SEQ ID NO:103 (RG2H); SEQ ID NO:105 (RG2I); SEQ ID NO:108 (RG2J); SEQ ID NO:111 (RG2K); SEQ ID NO:113 (RG2L); SEQ ID NO:115 (RG2M); SEQ ID NO:117 (RG2N); SEQ ID NO:119 (RG2O); SEQ ID NO:121 (RG2P); SEQ ID NO:123 (RG2Q); SEQ ID NO:125 (RG2S); SEQ ID NO:127 (RG2T); SEQ ID NO:129 (RG2U); SEQ ID NO:131 (RG2V); and, SEQ ID NO:133 (RG2W).

An exemplary RG3 polypeptide has the sequence shown in SEQ ID NO:138. An exemplary RG4 polypeptide has the sequence shown in SEQ ID NO:139. RG polynucleotides will have at least 60% identity, more typically at least 65% identity, generally at least 70% identity, and preferably at least 75% identity, more preferably at least 80% identity, and most preferably at least 85%, 90%, or 95% identity at the deduced amino acid level. The regions where substantial identity is assessed can be inclusive or exclusive of the nucleotide binding site or the leucine rich region.

Vectors and Transcriptional Control Elements

The invention, providing methods and reagents for making novel species and genres of RG nucleic acids described herein, further provides methods and reagents for expressing these nucleic acids using novel expression cassettes, vectors, transgenic plants and animals, using constitutive and inducible transcriptional and translational *cis*- (e.g., promoters and enhancers) and *trans*-acting control elements.

The expression of natural, recombinant or synthetic plant disease resistance polypeptide-encoding or other (*i.e.*, antisense, ribozyme) nucleic acids can be achieved by operably linking the coding region a promoter (that can be plant-specific or not, constitutive or inducible), incorporating the construct into an expression cassette (such as an expression vector), and introducing the resultant construct into an *in vitro* reaction system or a suitable host cell or organism. Synthetic procedures may also be used. Typical expression systems contain, in addition to coding or antisense sequence, transcription and translation terminators, polyadenylation sequences, transcription and translation initiation sequences, and promoters useful for transcribing DNA into RNA. The expression systems optionally at least one independent terminator sequence, sequences permitting replication of the cassette *in vivo*, e.g., plants, eukaryotes, or prokaryotes, or a combination thereof, (e.g., shuttle vectors) and selection markers for the selected expression system, e.g., plant, prokaryotic or eukaryotic systems. To ensure proper polypeptide expression under varying conditions, a polyadenylation region at the 3'-end of the coding region can be included (see Li (1997) *Plant Physiol.* 115:321-325, for a review of the polyadenylation of RNA in plants). The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from T-DNA (e.g., using *Agrobacterium tumefaciens* T-DNA replacement vectors, see e.g., Thykjaer (1997) *Plant Mol Biol.* 35:523-530; using a plasmid containing a gene of interest flanked by *Agrobacterium* T-DNA border repeat sequences; Hansen (1997) "T-strand integration in maize protoplasts after codelivery of a T-DNA substrate and virulence genes," *Proc. Natl. Acad. Sci. USA* 94:11726-11730.

To identify the promoters, the 5' portions of the clones described here are analyzed for sequences characteristic of promoter sequences. For instance, promoter sequence elements include the TATA box consensus sequence (TATAAT), which is usually 20 to 30 base pairs upstream of the transcription start site. In plants, further

upstream from the TATA box, at positions -80 to -100, there is typically a promoter element with a series of adenines surrounding the trinucleotide G (or T) N G (see, *e.g.*, Messing, in *Genetic Engineering in Plants*, pp. 221-227, Kosage, Meredith and Hollaender, eds. 1983). If proper polypeptide expression is desired, a polyadenylation region at the 3'-end of the RG coding region should be included. The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from viral genes, such as T-DNA.

The nucleic acids of the invention can be expressed in expression cassettes, vectors or viruses which are transiently expressed in cells using, for example, episomal expression systems (*e.g.*, cauliflower mosaic virus (CaMV) viral RNA is generated in the nucleus by transcription of an episomal minichromosome containing supercoiled DNA, Covey (1990) *Proc. Natl. Acad. Sci. USA* 87:1633-1637). Alternatively, coding sequences can be inserted into the host cell genome becoming an integral part of the host chromosomal DNA.

Selection markers can be incorporated into expression cassettes and vectors to confer a selectable phenotype on transformed cells and sequences coding for episomal maintenance and replication such that integration into the host genome is not required. For example, the marker may encode biocide resistance, such as antibiotic resistance, particularly resistance to chloramphenicol, kanamycin, G418, bleomycin, hygromycin, or herbicide resistance, such as resistance to chlorosulfuron or Basta, to permit selection of those cells transformed with the desired DNA sequences, see for example, Blondelet-Rouault (1997) *Gene* 190:315-317; Aubrecht (1997) *J. Pharmacol. Exp. Ther.* 281:992-997. Because selectable marker genes conferring resistance to substrates like neomycin or hygromycin can only be utilized in tissue culture, chemoresistance genes are also used as selectable markers *in vitro* and *in vivo*. See also, Mengiste (1997) "High-efficiency transformation of *Arabidopsis thaliana* with a selectable marker gene regulated by the T-DNA 1' promoter," *Plant J.* 12:945-948, showing that the 1' promoter is an attractive alternative to the cauliflower mosaic virus (CaMV) 35S promoter for the generation of T-DNA insertion lines, the 1' promoter may be especially beneficial for the secondary transformation of transgenic strains containing the 35S promoter to exclude homology-mediated gene silencing.

The endogenous promoters from the RG genes of the present invention can be used to direct expression of the genes. These promoters can also be used to direct expression of heterologous structural genes. The promoters can be used, for example, in recombinant expression cassettes to drive expression of genes conferring resistance to any number of pathogens or pests, including fungi, bacteria, and the like.

Constitutive Promoters

In construction of recombinant expression cassettes, vectors, transgenics, of the invention, a promoter fragment can be employed to direct expression of the desired gene in all tissues of a plant or animal. Promoters that drive expression continuously under physiological conditions are referred to as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include those from viruses which infect plants, such as the cauliflower mosaic virus (CaMV) 35S transcription initiation region; the 1'- or 2'-promoter derived from T-DNA of *Agrobacterium tumefaciens*; the promoter of the tobacco mosaic virus; and, other transcription initiation regions from various plant genes known to those of skill. See also Holtorf (1995) "Comparison of different constitutive and inducible promoters for the overexpression of transgenes in *Arabidopsis thaliana*," *Plant Mol. Biol.* 29:637-646.

Inducible Promoters

Alternatively, a plant promoter may direct expression of the plant disease resistance nucleic acid of the invention under the influence of changing environmental conditions or developmental conditions. Examples of environmental conditions that may effect transcription by inducible promoters include pathogenic attack, anaerobic conditions, elevated temperature, drought, or the presence of light. Such promoters are referred to herein as "inducible" promoters. For example, the invention incorporates the drought-inducible promoter of maize (Busk (1997) *supra*); the cold, drought, and high salt inducible promoter from potato (Kirch (1997) *Plant Mol. Biol.* 33:897-909).

Embodiments of the invention also incorporate use of plant promoters which are inducible upon injury or infection to express the invention's plant disease resistance (RG) polypeptides. Various embodiments include use of, *e.g.*, the promoter for a tobacco (*Nicotiana tabacum*) sesquiterpene cyclase gene (EAS4 promoter), which is expressed in wounded leaves, roots, and stem tissues, and upon infection with microbial pathogens (Yin

(1997) *Plant Physiol.* 115(2):437-451); the ORF13 promoter from *Agrobacterium rhizogenes* 8196, which is wound inducible in a limited area adjacent to the wound site (Hansen (1997) *Mol. Gen. Genet.* 254:337-343); the Shpx6b gene promoter, which is a plant peroxidase gene promoter induced by microbial pathogens (demonstrated using a fungal pathogen, see Curtis (1997) *Mol. Plant Microbe Interact.* 10:326-338); the wound-inducible gene promoter wun1, derived from potato (Siebertz (1989) *Plant Cell* 1:961-968); the wound-inducible *Agrobacterium pmas* gene (mannopine synthesis gene) promoter (Guevara-Garcia (1993) *Plant J.* 4:495-505).

Alternatively, plant promoters which are inducible upon exposure to plant hormones, such as auxins, are used to express the nucleic acids of the invention. For example, the invention can use the auxin-response elements E1 promoter fragment (AuxREs) in the soybean (*Glycine max* L.) (Liu (1997) *Plant Physiol.* 115:397-407); the auxin-responsive Arabidopsis GST6 promoter (also responsive to salicylic acid and hydrogen peroxide) (Chen (1996) *Plant J.* 10: 955-966); the auxin-inducible parC promoter from tobacco (Sakai (1996) 37:906-913); a plant biotin response element (Streit (1997) *Mol. Plant Microbe Interact.* 10:933-937); and, the promoter responsive to the stress hormone abscisic acid (Sheen (1996) *Science* 274:1900-1902).

Plant promoters which are inducible upon exposure to chemicals reagents which can be applied to the plant, such as herbicides or antibiotics, are also used to express the nucleic acids of the invention. For example, the maize In2-2 promoter, activated by benzenesulfonamide herbicide safeners, can be used (De Veylder (1997) *Plant Cell Physiol.* 38:568-577); application of different herbicide safeners induces distinct gene expression patterns, including expression in the root, hydathodes, and the shoot apical meristem. Coding sequence can be under the control of, *e.g.*, a tetracycline-inducible promoter, *e.g.*, as described with transgenic tobacco plants containing the *Avena sativa* L. (oat) arginine decarboxylase gene (Masgrau (1997) *Plant J.* 11:465-473); or, a salicylic acid-responsive element (Stange (1997) *Plant J.* 11:1315-1324. Using chemically- (*e.g.*, hormone- or pesticide-) induced promoters, harvesting of fruits and plant parts would be greatly facilitated. A chemical which can be applied to the transgenic plant in the field and induce expression of a polypeptide of the invention throughout all or most of the plant would make an environmentally safe defoliant or herbicide. Thus, the invention also provides for transgenic plants containing an inducible gene encoding for the RG

polypeptides of the invention whose host range is limited to target plant species, such as weeds or crops before, during or after harvesting.

Abcission promoters are activated upon plant ripening, such as fruit ripening, and are especially useful incorporated in the expression systems (*e.g.*, expression cassettes, vectors) of the invention. In some embodiments, when a plant disease resistant polypeptide-encoding nucleic acid is under the control of such a promoter, rapid cell death, induced by expression of the invention's polypeptide, can accelerate and/or accentuate abscission, increasing the efficiency of the harvesting of fruits or other plant parts, such as cotton, and the like. Induction of rapid cell death at this time would accelerate separation of the fruit from the plant, greatly augmenting harvesting procedures. See, *e.g.*, Kalaitzis (1997) *Plant Physiol.* 113:1303-1308, discussing tomato leaf and flower abscission; Payton (1996) *Plant Mol. Biol.* 31:1227-1231, discussing ethylene receptor expression regulation during fruit ripening, flower senescence and abscission; Koehler (1996) *Plant Mol. Biol.* 31:595-606, discussing the gene promoter for a bean abscission cellulase; Kalaitzis (1995) *Plant Mol. Biol.* 28: 647-656, discussing cloning of a tomato polygalacturonase expressed in abscission; del Campillo (1996) *Plant Physiol.* 111:813-820, discussing pedicel breakstrength and cellulase gene expression during tomato flower abscission.

Tissue-Specific Promoters

Tissue specific promoters are transcriptional control elements that are only active in particular cells or tissues. Plant promoters which are active only in specific tissues or at specific times during plant development are used to express the nucleic acids of the invention. Examples of promoters under developmental control include promoters that initiate transcription only in certain tissues, such as leaves, roots, fruit, seeds, ovules, pollen, pistils, or flowers. Such promoters are referred to as "tissue specific". The operation of a promoter may also vary depending on its location in the genome. Thus, an inducible promoter may become fully or partially constitutive in certain locations.

For example, a seed-specific promoter directs expression in seed tissues. Such promoters may be, for example, ovule-specific, embryo-specific, endosperm-specific, integument-specific, seed coat-specific, or some combination thereof. A leaf-specific promoter has been identified in maize, Busk (1997) *Plant J.* 11:1285-1295. The ORF13 promoter from *Agrobacterium rhizogenes* exhibits high activity in roots (Hansen (1997) *supra*). A maize pollen-specific promoter has been identified in maize (Guerrero (1990)

Mol. Gen. Genet. 224:161-168). A tomato promoter active during fruit ripening, senescence and abscission of leaves and, to a lesser extent, of flowers can be used (Blume (1997) *Plant J.* 12:731-746). A pistil specific promoter has been identified in the potato (*Solanum tuberosum* L.) SK2 gene, encoding a pistil-specific basic endochitinase (Ficker (1997) *Plant Mol. Biol.* 35:425-431). The Blec4 gene from pea (*Pisum sativum* cv. Alaska) is active in epidermal tissue of vegetative and floral shoot apices of transgenic alfalfa, making it a useful tool to target the expression of foreign genes to the epidermal layer of actively growing shoots. The activity of the Blec4 promoter in the epidermis of the shoot apex makes it particularly suitable for genetically engineering defense against insects and diseases that attack the growing shoot apex (Mandaci (1997) *Plant Mol Biol.* 34:961-965).

The invention also provides for use of tissue-specific plant promoters include a promoter from the ovule-specific *BEL1* gene described in Reiser (1995) *Cell* 83:735-742, GenBank No. U39944. Suitable seed specific promoters are derived from the following genes: *MAC1* from maize, Sheridan (1996) *Genetics* 142:1009-1020; *Cat3* from maize, GenBank No. L05934, Abler (1993) *Plant Mol. Biol.* 22:10131-1038; the gene encoding oleosin 18kD from maize, GenBank No. J05212, Lee (1994) *Plant Mol. Biol.* 26:1981-1987; viviparous-1 from *Arabidopsis*, Genbank No. U93215; the gene encoding oleosin from *Arabidopsis*, Genbank No. Z17657; *Atmyc1* from *Arabidopsis*, Urao (1996) *Plant Mol. Biol.* 32:571-576; the 2s seed storage protein gene family from *Arabidopsis*, Conceicao (1994) *Plant* 5:493-505; the gene encoding oleosin 20kD from *Brassica napus*, GenBank No. M63985; *napA* from *Brassica napus*, GenBank No. J02798, Josefsson (1987) *JBL* 26:12196-1301; the napin gene family from *Brassica napus*, Sjodahl (1995) *Planta* 197:264-271; the gene encoding the 2S storage protein from *Brassica napus*, Dasgupta (1993) *Gene* 133:301-302; the genes encoding oleosin a, Genbank No. U09118, and, oleosin B, Genbank No. U09119, from soybean; and, the gene encoding low molecular weight sulphur rich protein from soybean, Choi (1995) *Mol Gen, Genet.* 246:266-268. The tissue specific E8 promoter from tomato is particularly useful for directing gene expression so that a desired gene product is located in fruits. Other suitable promoters include those from genes encoding embryonic storage proteins.

One of skill will recognize that a tissue-specific promoter may drive expression of operably linked sequences in tissues other than the target tissue. Thus, as

used herein a tissue-specific promoter is one that drives expression preferentially in the target tissue, but may also lead to some expression in other tissues as well.

The invention also provides for use of tissue-specific promoters derived from viruses which can include, *e.g.*, the tobamovirus subgenomic promoter (Kumagai (1995) *Proc. Natl. Acad. Sci. USA* 92:1679-1683; the rice tungro bacilliform virus (RTBV), which replicates only in phloem cells in infected rice plants, with its promoter which drives strong phloem-specific reporter gene expression; the cassava vein mosaic virus (CVMV) promoter, with highest activity in vascular elements, in leaf mesophyll cells, and in root tips (Verdaguer (1996) *Plant Mol. Biol.* 31:1129-1139).

In some embodiments, the nucleic acid construct will comprise a promoter functional in a specific plant cell, such as in a species of *Lactuca*, operably linked to an RG polynucleotide. Promoters useful in these embodiments include RG promoters. In additional embodiments, the nucleic acid construct will comprise a RG promoter operably linked to a heterologous polynucleotide. The heterologous polynucleotide is chosen to provide a plant with a desired phenotype. For example, the heterologous polynucleotide can be a structural gene which encodes a polypeptide which imparts a desired resistance phenotype. Alternatively, the heterologous polynucleotide may be a regulatory gene which might play a role in transcriptional and/or translational control to suppress, enhance, or otherwise modify the transcription and/or expression of an endogenous gene within the plant. The heterologous polynucleotide of the nucleic acid construct of the present invention can be expressed in either sense or anti-sense orientation as desired. It will be appreciated that control of gene expression in either sense or anti-sense orientation can have a direct impact on the observable plant characteristics.

Modifying and Inhibiting RG Gene Expression

The invention also provides for RG nucleic acid sequences which are complementary to the RG polypeptide-encoding sequences of the invention; *i.e.*, antisense RG nucleic acids. Antisense technology can be conveniently used to modify gene expression in plants. To accomplish this, a nucleic acid segment from the desired gene is cloned and operably linked to a promoter such that the anti-sense strand of RNA will be transcribed. The construct is then transformed into plants and the antisense strand of RNA is produced. In plant cells, it has been shown that antisense RNA inhibits gene expression by preventing the accumulation of mRNA which encodes the enzyme of interest, see, *e.g.*,

Sheehy (1988) *Proc. Nat. Acad. Sci. USA* 85:8805-8809; Hiatt et al., U.S. Patent No. 4,801,340.

Antisense sequences are capable of inhibiting the transport, splicing or transcription of RG-encoding genes. The inhibition can be effected through the targeting of genomic DNA or messenger RNA. The transcription or function of targeted nucleic acid can be inhibited, *e.g.*, by hybridization and/or cleavage. One particularly useful set of inhibitors provided by the present invention includes oligonucleotides which are able to either bind RG gene or message, in either case preventing or inhibiting the production or function of RG. The association can be through sequence specific hybridization. Such inhibitory nucleic acid sequences can, for example, be used to completely inhibit a plant disease resistance response. Another useful class of inhibitors includes oligonucleotides which cause inactivation or cleavage of RG message. The oligonucleotide can have enzyme activity which causes such cleavage, such as ribozymes. The oligonucleotide can be chemically modified or conjugated to an enzyme or composition capable of cleaving the complementary nucleic acid. One may screen a pool of many different such oligonucleotides for those with the desired activity.

Antisense Oligonucleotides

The invention provides for with antisense oligonucleotides capable of binding RG message which can inhibit RG activity by targeting mRNA. Strategies for designing antisense oligonucleotides are well described in the scientific and patent literature, and the skilled artisan can design such RG oligonucleotides using the novel reagents of the invention. In some situations, naturally occurring nucleic acids used as antisense oligonucleotides may need to be relatively long (18 to 40 nucleotides) and present at high concentrations. A wide variety of synthetic, non-naturally occurring nucleotide and nucleic acid analogues are known which can address this potential problem. For example, peptide nucleic acids (PNAs) containing non-ionic backbones, such as N-(2-aminoethyl) glycine units can be used. Antisense oligonucleotides having phosphorothioate linkages can also be used, as described in WO 97/03211; WO 96/39154; Mata (1997) *Toxicol Appl Pharmacol* 144:189-197; Antisense Therapeutics, ed. Agrawal (Humana Press, Totowa, N.J., 1996). Antisense oligonucleotides having synthetic DNA backbone analogues provided by the invention can also include phosphoro-dithioate, methylphosphonate,

phosphoramidate, alkyl phosphotriester, sulfamate, 3'-thioacetal, methylene(methylimino), 3'-N-carbamate, and morpholino carbamate nucleic acids, as described herein.

Combinatorial chemistry methodology can be used to create vast numbers of oligonucleotides that can be rapidly screened for specific oligonucleotides that have appropriate binding affinities and specificities toward any target, such as the sense and antisense RG sequences of the invention (for general background information, see, *e.g.*, Gold (1995) *J. of Biol. Chem.* 270:13581-13584).

Inhibitory Ribozymes

The invention provides for with ribozymes capable of binding RG message which can inhibit RG activity by targeting mRNA. Strategies for designing ribozymes and selecting the RG-specific antisense sequence for targeting are well described in the scientific and patent literature, and the skilled artisan can design such RG ribozymes using the novel reagents of the invention. Ribozymes act by binding to a target RNA through the target RNA binding portion of a ribozyme which is held in close proximity to an enzymatic portion of the RNA that cleaves the target RNA. Thus, the ribozyme recognizes and binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cleave and inactivate the target RNA. Cleavage of a target RNA in such a manner will destroy its ability to direct synthesis of an encoded protein if the cleavage occurs in the coding sequence, or, preventing transport of the message from the nucleus to the cytoplasm. After a ribozyme has bound and cleaved its RNA target, it is typically released from that RNA and so can bind and cleave new targets repeatedly.

Catalytic RNA molecules or ribozymes can also be used to inhibit expression of any plant gene. It is possible to design ribozymes that specifically pair with virtually any target RNA and cleave the phosphodiester backbone at a specific location, thereby functionally inactivating the target RNA. In carrying out this cleavage, the ribozyme is not itself altered, and is thus capable of recycling and cleaving other molecules, making it a true enzyme. The inclusion of ribozyme sequences within antisense RNAs confers RNA-cleaving activity upon them, thereby increasing the activity of the constructs. The design and use of target RNA-specific ribozymes is described, *e.g.*, in Haseioff (1988) *Nature* 334:585-591.

In some circumstances, the enzymatic nature of a ribozyme can be advantageous over other technologies, such as antisense technology (where a nucleic acid

molecule simply binds to a nucleic acid target to block its transcription, translation or association with another molecule) as the effective concentration of ribozyme necessary to effect a therapeutic treatment can be lower than that of an antisense oligonucleotide. This potential advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single
5 ribozyme molecule is able to cleave many molecules of target RNA. In addition, a ribozyme is typically a highly specific inhibitor, with the specificity of inhibition depending not only on the base pairing mechanism of binding, but also on the mechanism by which the molecule inhibits the expression of the RNA to which it binds. That is, the inhibition is caused by cleavage of the RNA target and so specificity is defined as the ratio
10 of the rate of cleavage of the targeted RNA over the rate of cleavage of non-targeted RNA. This cleavage mechanism is dependent upon factors additional to those involved in base pairing. Thus, the specificity of action of a ribozyme can be greater than that of antisense oligonucleotide binding the same RNA site.

The enzymatic ribozyme RNA molecule can be formed in a hammerhead
15 motif, but may also be formed in the motif of a hairpin, hepatitis delta virus, group I intron or RNaseP-like RNA (in association with an RNA guide sequence). Examples of such hammerhead motifs are described by Rossi (1992) *Aids Research and Human Retroviruses* 8:183; hairpin motifs by Hampel (1989) *Biochemistry* 28:4929, and Hampel (1990) *Nuc. Acids Res.* 18:299; the hepatitis delta virus motif by Perrotta (1992) *Biochemistry* 31:16;
20 the RNaseP motif by Guerrier-Takada (1983) *Cell* 35:849; and the group I intron by Cech U.S. Pat. No. 4,987,071. The recitation of these specific motifs is not intended to be limiting; those skilled in the art will recognize that an enzymatic RNA molecule of this invention has a specific substrate binding site complementary to one or more of the target gene RNA regions, and has nucleotide sequence within or surrounding that substrate
25 binding site which imparts an RNA cleaving activity to the molecule.

Sense Supression

Another method of suppression is sense suppression. Introduction of nucleic acid configured in the sense orientation has been shown to be an effective means by which to block the transcription of target genes. For an example of the use of this method
30 to modulate expression of endogenous genes see, Napoli et al., *The Plant Cell* 2:279-289 (1990), and U.S. Patent No. 5,034,323.

Cloning of RG Polypeptides

4

Synthesis and/or cloning of RG polynucleotides and isolated nucleic acid constructs of the present invention are provided by methods well known to those of ordinary skill in the art. Generally, the nomenclature and the laboratory procedures in recombinant DNA technology described below are those well known and commonly employed in the art. Standard techniques are used for cloning, DNA and RNA isolation, amplification and purification. Generally enzymatic reactions involving DNA ligase, DNA polymerase, restriction endonucleases and the like are performed according to the manufacturer's specifications. These techniques and various other techniques are generally performed according to Sambrook *et al.*, *Molecular Cloning - A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, (1989).

The isolation of RG genes may be accomplished by a number of techniques. For instance, oligonucleotide probes based on the sequences disclosed here can be used to identify the desired gene in a cDNA or genomic DNA library. To construct genomic libraries, large segments of genomic DNA are generated by random fragmentation, e.g. using restriction endonucleases, and are ligated with vector DNA to form concatemers that can be packaged into the appropriate vector. To prepare a cDNA library, mRNA is isolated from the desired organ, such as roots and a cDNA library which contains the RG gene transcript is prepared from the mRNA. Alternatively, cDNA may be prepared from mRNA extracted from other tissues in which RG genes or homologs are expressed.

The cDNA or genomic library can then be screened using a probe based upon the sequence of a cloned RG gene such as the genes disclosed herein. Probes may be used to hybridize with genomic DNA or cDNA sequences to isolate homologous genes in the same or different plant species.

Those of skill in the art will appreciate that various degrees of stringency of hybridization can be employed in the assay; and either the hybridization or the wash medium can be stringent. As the conditions for hybridization become more stringent, there must be a greater degree of complementarity between the probe and the target for duplex formation to occur. The degree of stringency can be controlled by temperature, ionic strength, pH and the presence of a partially denaturing solvent such as formamide. For example, the stringency of hybridization is conveniently varied by changing the polarity of the reactant solution through manipulation of the concentration of formamide within the range of 0% to 50%.

Alternatively, the RG nucleic acids of the invention can be amplified from nucleic acid samples using a variety of amplification techniques, such as polymerase chain reaction (PCR) technology, to amplify the sequences of the RG and related genes directly from genomic DNA, from cDNA, from genomic libraries or cDNA libraries. PCR and other *in vitro* amplification methods may also be useful, for example, to clone nucleic acid sequences that code for proteins to be expressed, to make nucleic acids to use as probes for detecting the presence of the desired mRNA in samples, for nucleic acid sequencing, or for other purposes.

Oligonucleotides can be used to identify and detect additional RG families and RG family species using a variety of hybridization techniques and conditions. Suitable amplification methods include, but are not limited to: polymerase chain reaction, PCR (PCR PROTOCOLS, A GUIDE TO METHODS AND APPLICATIONS, *ed.* Innis, Academic Press, N.Y. (1990) and PCR STRATEGIES (1995), *ed.* Innis, Academic Press, Inc., N.Y. (Innis)), ligase chain reaction (LCR) (Wu (1989) *Genomics* 4:560; Landegren (1988) *Science* 241:1077; Barringer (1990) *Gene* 89:117); transcription amplification (Kwoh (1989) *Proc. Natl. Acad. Sci. USA* 86:1173); and, self-sustained sequence replication (Guatelli (1990) *Proc. Natl. Acad. Sci. USA*, 87:1874); Q Beta replicase amplification and other RNA polymerase mediated techniques (*e.g.*, NASBA, Cingene, Mississauga, Ontario); see Berger (1987) *Methods Enzymol.* 152:307-316, Sambrook, and Ausubel, as well as Mullis (1987) U.S. Patent Nos. 4,683,195 and 4,683,202; Arnheim (1990) *C&EN* 36-47; Lomell *J. Clin. Chem.*, 35:1826 (1989); Van Brunt, *Biotechnology*, 8:291-294 (1990); Wu (1989) *Gene* 4:560; Sooknanan (1995) *Biotechnology* 13:563-564. Methods for cloning *in vitro* amplified nucleic acids are described in Wallace, U.S. Pat. No. 5,426,039.

The degree of complementarity (sequence identity) required for detectable binding will vary in accordance with the stringency of the hybridization medium and/or wash medium. The degree of complementarity will optimally be 100 percent; however, it should be understood that minor sequence variations in the probes and primers may be compensated for by reducing the stringency of the hybridization and/or wash medium as described earlier.

In some preferred embodiments, members of this class of pest resistance genes can be identified by their ability to be amplified by PCR primers based on the sequences disclosed here. Appropriate primers and probes for identifying RG sequences

from plant tissues are generated from comparisons of the sequences provided herein. See, e.g., Table 1. For a general overview of PCR see *PCR Protocols: A Guide to Methods and Applications*. (Innis, M, Gelfand, D., Sninsky, J. and White, T., eds.), Academic Press, San Diego (1990), incorporated herein by reference.

5 Briefly, the first step of each cycle of the PCR involves the separation of the nucleic acid duplex formed by the primer extension. Once the strands are separated, the next step in PCR involves hybridizing the separated strands with primers that flank the target sequence. The primers are then extended to form complementary copies of the target strands. For successful PCR amplification, the primers are designed so that the
10 position at which each primer hybridizes along a duplex sequence is such that an extension product synthesized from one primer, when separated from the template (complement), serves as a template for the extension of the other primer. The cycle of denaturation, hybridization, and extension is repeated as many times as necessary to obtain the desired amount of amplified nucleic acid.

15 In the preferred embodiment of the PCR process, strand separation is achieved by heating the reaction to a sufficiently high temperature for an sufficient time to cause the denaturation of the duplex but not to cause an irreversible denaturation of the polymerase (see U.S. Patent No. 4,965,188). Template-dependent extension of primers in PCR is catalyzed by a polymerizing agent in the presence of adequate amounts of four
20 deoxyribonucleotide triphosphates (typically dATP, dGTP, dCTP, and dTTP) in a reaction medium comprised of the appropriate salts, metal cations, and pH buffering system. Suitable polymerizing agents are enzymes known to catalyze template-dependent DNA synthesis.

Polynucleotides may also be synthesized by well-known techniques as
25 described in the technical literature. See, e.g., Carruthers *et al.*, *Cold Spring Harbor Symp. Quant. Biol.* 47:411-418 (1982), and Adams *et al.*, *J. Am. Chem. Soc.* 105:661 (1983). Double stranded DNA fragments may then be obtained either by synthesizing the complementary strand and annealing the strands together under appropriate conditions, or by adding the complementary strand using DNA polymerase with an appropriate primer
30 sequence.

RG Proteins

The present invention further provides isolated RG proteins encoded by the RG polynucleotides disclosed herein. One of skill will recognize that the nucleic acid encoding a functional RG protein need not have a sequence identical to the exemplified genes disclosed here. For example, because of codon degeneracy a large number of nucleic acid sequences can encode the same polypeptide. In addition, the polypeptides encoded by the RG genes, like other proteins, have different domains which perform different functions. Thus, the RG gene sequences need not be full length, so long as the desired functional domain of the protein is expressed.

The resistance proteins are at least 25 amino acid residues in length. Typically, the RG proteins are at least 50 amino acid residues, generally at least 100, preferably at least 150, more preferably at least 200 amino acids in length. In particularly preferred embodiments, the RG proteins are of sufficient length to provide resistance to pests when expressed in the desired plants. Generally then, the RG proteins will be the length encoded by an RG gene of the present invention. However, those of ordinary skill will appreciate that minor deletions, substitutions, or additions to an RG protein will typically yield a protein with pest resistance characteristics similar or identical to that of the full length sequence. Thus, full-length RG proteins modified by 1, 2, 3, 4, or 5 deletions, substitutions, or additions, generally provide an effective degree of pest resistance relative to the full-length protein.

The RG proteins which provide pest resistance will typically comprise at least one of an LRR or an NBS. Preferably, both are present. LRR and/or NBS regions present in the RG proteins of the present invention can be provided by RG genes of the present invention. In some embodiments, the LRR and/or NBS regions are obtained from other pest resistance genes. See, e.g., Yu *et al.*, *Proc. Natl. Acad. Sci. USA*, 93: 11751-11756 (1996); Bent *et al.*, *Science*, 265: 1856-1860 (1994).

Modified protein chains can also be readily designed utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the chains can vary from the naturally occurring sequence at the primary structure level by amino acid substitutions, additions, deletions, and the like. Modification can also include swapping domains from the proteins of the invention with related domains from other pest resistance genes.

Pests that can be targeted by RG genes and proteins of the present invention include such bacterial pests as *Erwinia carotovora* and *Pseudomonas marginalis*. Fungal pests which can be targeted by the present invention include *Bremia lactucae*, *Marssonina panattoniana*, *Rhizoctonia solani*, *Olpidium brassicae*, root aphid, *Sclerotinia sclerotiorum* and *S. minor*, and *Botrytis cinerea* which causes gray mold. RG genes also provide resistance to viral diseases such as lettuce and turnip mosaic viruses.

Fusion Proteins

RG polypeptides can also be expressed as recombinant proteins with one or more additional polypeptide domains linked thereto to facilitate protein detection, purification, or other applications. Such detection and purification facilitating domains include, but are not limited to, metal chelating peptides such as polyhistidine tracts and histidine-tryptophan modules that allow purification on immobilized metals, protein domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp, Seattle WA). The inclusion of a cleavable linker sequences such as Factor Xa or enterokinase (Invitrogen, San Diego CA) between the purification domain and plant disease resistant polypeptide may be useful to facilitate purification. One such expression vector provides for expression of a fusion protein comprising the sequence encoding a plant disease resistant polypeptide of the invention and nucleic acid sequence encoding six histidine residues followed by thioredoxin and an enterokinase cleavage site (*e.g.*, see Williams (1995) *Biochemistry* 34:1787-1797). The histidine residues facilitate detection and purification while the enterokinase cleavage site provides a means for purifying the desired protein(s) from the remainder of the fusion protein. Technology pertaining to vectors encoding fusion proteins and application of fusion proteins are well described, see *e.g.*, Kroll (1993) *DNA Cell. Biol.*, 12:441-53.

Antibodies Reactive to RG Polypeptides and Immunological Assays

The present invention also provides antibodies which specifically react with RG proteins of the present invention under immunologically reactive conditions. An antibody immunologically reactive with a particular antigen can be generated *in vivo* or by recombinant methods such as selection of libraries of recombinant antibodies in phage or similar vectors. "Immunologically reactive conditions" includes reference to conditions which allow an antibody, generated to a particular epitope of an antigen, to bind to that

epitope to a detectably greater degree than the antibody binds to substantially all other epitopes, generally at least two times above background binding, preferably at least five times above background. Immunologically reactive conditions are dependent upon the format of the antibody binding reaction and typically are those utilized in immunoassay protocols.

"Antibody" includes reference to an immunoglobulin molecule obtained by *in vitro* or *in vivo* generation of the humoral response, and includes both polyclonal and monoclonal antibodies. The term also includes genetically engineered forms such as chimeric antibodies (e.g., humanized murine antibodies), heteroconjugate antibodies (e.g., bispecific antibodies), and recombinant single chain Fv fragments (scFv). The term "antibody" also includes antigen binding forms of antibodies (e.g., Fab', F(ab')₂, Fab, Fv, rIgG, and, inverted IgG). See, Pierce Catalog and Handbook, 1994-1995 (Pierce Chemical Co., Rockford, IL). An antibody immunologically reactive with a particular antigen can be generated *in vivo* or by recombinant methods such as selection of libraries of recombinant antibodies in phage or similar vectors. See, e.g., Huse *et al.* (1989) *Science* 246:1275-1281; and Ward, *et al.* (1989) *Nature* 341:544-546; and Vaughan *et al.* (1996) *Nature Biotechnology*, 14:309-314.

Many methods of making antibodies are known to persons of skill. A number of immunogens are used to produce antibodies specifically reactive to an isolated RG protein of the present invention under immunologically reactive conditions. An isolated recombinant, synthetic, or native RG protein of the present invention is the preferred immunogens (antigen) for the production of monoclonal or polyclonal antibodies.

The RG protein is then injected into an animal capable of producing antibodies. Either monoclonal or polyclonal antibodies can be generated for subsequent use in immunoassays to measure the presence and quantity of the RG protein. Methods of producing monoclonal or polyclonal antibodies are known to those of skill in the art. See, e.g., Coligan (1991) *Current Protocols in Immunology* Wiley/Greene, NY; and Harlow and Lane (1989) *Antibodies: A Laboratory Manual* Cold Spring Harbor Press, NY; Goding (1986) *Monoclonal Antibodies: Principles and Practice* (2d ed.) Academic Press, New York, NY.

Frequently, the RG proteins and antibodies will be labeled by joining, either covalently or non-covalently, a substance which provides for a detectable signal. A wide

variety of labels and conjugation techniques are known and are reported extensively in both the scientific and patent literature. Suitable labels include radionucleotides, enzymes, substrates, cofactors, inhibitors, fluorescent moieties, chemiluminescent moieties, magnetic particles, and the like. Patents teaching the use of such labels include U.S. Patent Nos. 3,817,837; 3,850,752; 3,939,350; 3,996,345; 4,277,437; 4,275,149; and 4,366,241.

The antibodies of the present invention can be used to screen plants for the expression of RG proteins of the present invention. The antibodies of this invention are also used for affinity chromatography in isolating RG protein.

The present invention further provides RG polypeptides that specifically bind, under immunologically reactive conditions, to an antibody generated against a defined immunogen, such as an immunogen consisting of the RG polypeptides of the present invention. Immunogens will generally be at least 10 contiguous amino acids from an RG polypeptide of the present invention. Optionally, immunogens can be from regions exclusive of the NBS and/or LRR regions of the RG polypeptides. Nucleic acids which encode such cross-reactive RG polypeptides are also provided by the present invention. The RG polypeptides can be isolated from any number plants as discussed earlier. Preferred are species from the family *Compositae* and in particular the genus *Lactuca* such as *L. sativa* and such subspecies as *crispa*, *longifolia*, and *asparagina*.

"Specifically binds" includes reference to the preferential association of a ligand, in whole or part, with a particular target molecule (i.e., "binding partner" or "binding moiety") relative to compositions lacking that target molecule. It is, of course, recognized that a certain degree of non-specific interaction may occur between a ligand and a non-target molecule. Nevertheless, specific binding, may be distinguished as mediated through specific recognition of the target molecule. Typically specific binding results in a much stronger association between the ligand and the target molecule than between the ligand and non-target molecule. Specific binding by an antibody to a protein under such conditions requires an antibody that is selected for its specificity for a particular protein. The affinity constant of the antibody binding site for its cognate monovalent antigen is at least 10^7 , usually at least 10^8 , preferably at least 10^9 , more preferably at least 10^{10} , and most preferably at least 10^{11} liters/mole. A variety of immunoassay formats are appropriate for selecting antibodies specifically reactive with a particular protein. For

example, solid-phase ELISA immunoassays are routinely used to select monoclonal antibodies specifically reactive with a protein. See Harlow and Lane (1988) *Antibodies, A Laboratory Manual*, Cold Spring Harbor Publications, New York, for a description of immunoassay formats and conditions that can be used to determine specific reactivity. The antibody may be polyclonal but preferably is monoclonal. Generally, antibodies cross-reactive to such proteins as RPS2, RPM1 (bacterial resistances in *Arabidopsis*, L6 (fungal resistance in flax, PRF (resistance to *Pseudomonas syringae* in tomato), and *N*, (virus resistance in tobacco), are removed by immunoabsorption.

Immunoassays in the competitive binding format are typically used for cross-reactivity determinations. For example, an immunogenic RG polypeptide is immobilized to a solid support. Polypeptides added to the assay compete with the binding of the antisera to the immobilized antigen. The ability of the above polypeptides to compete with the binding of the antisera to the immobilized RG polypeptide is compared to the immunogenic RG polypeptide. The percent cross-reactivity for the above proteins is calculated, using standard calculations. Those antisera with less than 10% cross-reactivity with such proteins as RPS2, RPM1, L6, PRF, and *N*, are selected and pooled. The cross-reacting antibodies are then removed from the pooled antisera by immunoabsorption with these non-RG resistance proteins.

The immunoabsorbed and pooled antisera are then used in a competitive binding immunoassay to compare a second "target" polypeptide to the immunogenic polypeptide. In order to make this comparison, the two polypeptides are each assayed at a wide range of concentrations and the amount of each polypeptide required to inhibit 50% of the binding of the antisera to the immobilized protein is determined using standard techniques. If the amount of the target polypeptide required is less than twice the amount of the immunogenic polypeptide that is required, then the target polypeptide is said to specifically bind to an antibody generated to the immunogenic protein. As a final determination of specificity, the pooled antisera is fully immunosorbed with the immunogenic polypeptide until no binding to the polypeptide used in the immunosorption is detectable. The fully immunosorbed antisera is then tested for reactivity with the test polypeptide. If no reactivity is observed, then the test polypeptide is specifically bound by the antisera elicited by the immunogenic protein.

Production of transgenic plants of the invention

Isolated nucleic acid constructs prepared as described herein can be introduced into plants according techniques known in the art. In some embodiments, the introduced nucleic acid is used to provide RG gene expression and therefore pest resistance in desired plants. In some embodiments, RG promoters are used to drive expression of desired heterologous genes in plants. Finally, in some embodiments, the constructs can be used to suppress expression of a target endogenous gene, including RG genes.

To use isolated RG sequences in the above techniques, recombinant DNA vectors suitable for transformation of plant cells are prepared. Techniques for transforming a wide variety of higher plant species are well known and described in the technical and scientific literature. See, for example, Weising *et al. Ann. Rev. Genet.* 22:421-477 (1988).

A DNA sequence coding for the desired RG polypeptide, for example a cDNA or a genomic sequence encoding a full length protein, will be used to construct a recombinant expression cassette which can be introduced into the desired plant. An expression cassette will typically comprise the RG polynucleotide operably linked to transcriptional and translational initiation regulatory sequences which will direct the transcription of the sequence from the RG gene in the intended tissues of the transformed plant.

Such DNA constructs may be introduced into the genome of the desired plant host by a variety of conventional techniques. For example, the DNA construct may be introduced directly into the genomic DNA of the plant cell using techniques such as electroporation, PEG poration, particle bombardment and microinjection of plant cell protoplasts or embryogenic callus, or the DNA constructs can be introduced directly to plant tissue using ballistic methods, such as DNA particle bombardment. Alternatively, the DNA constructs may be combined with suitable T-DNA flanking regions and introduced into a conventional *Agrobacterium tumefaciens* host vector. The virulence functions of the *Agrobacterium tumefaciens* host will direct the insertion of the construct and adjacent marker into the plant cell DNA when the cell is infected by the bacteria.

Transformation techniques are known in the art and well described in the scientific and patent literature. The introduction of DNA constructs using polyethylene glycol precipitation is described in Paszkowski *et al. Embo J.* 3:2717-2722 (1984).

Electroporation techniques are described in Fromm *et al. Proc. Natl. Acad. Sci. USA* 82:5824 (1985). Ballistic transformation techniques are described in Klein *et al. Nature* 327:70-73 (1987).

5 *Agrobacterium tumefaciens*-mediated transformation techniques are well described in the scientific literature. See, for example Horsch *et al. Science* 233:496-498 (1984), and Fraley *et al. Proc. Natl. Acad. Sci. USA* 80:4803 (1983). Although *Agrobacterium* is useful primarily in dicots, certain monocots can be transformed by *Agrobacterium*. For instance, *Agrobacterium* transformation of rice is described by Hiei *et al. Plant J.* 6:271-282 (1994). A particularly preferred means of transforming lettuce is described in Michelmore *et al., Plant Cell Reports*, 6:439-442 (1987).

Transformed plant cells which are derived by any of the above transformation techniques can be cultured to regenerate a whole plant which possesses the transformed genotype and thus the desired RG-controlled phenotype. Such regeneration techniques rely on manipulation of certain phytohormones in a tissue culture growth medium, typically relying on a biocide and/or herbicide marker which has been introduced together with the RG nucleotide sequences. Plant regeneration from cultured protoplasts is described in Evans *et al., Protoplasts Isolation and Culture, Handbook of Plant Cell Culture*, pp. 124-176, Macmillan Publishing Company, New York, 1983; and Binding, *Regeneration of Plants, Plant Protoplasts*, pp. 21-73, CRC Press, Boca Raton, 1985.

15
20 Regeneration can also be obtained from plant callus, explants, organs, or parts thereof. Such regeneration techniques are described generally in Klee *et al. Ann. Rev. of Plant Phys.* 38:467-486 (1987).

The methods of the present invention are particularly useful for incorporating the RG polynucleotides into transformed plants in ways and under circumstances which are not found naturally. In particular, the RG polypeptides may be expressed at times or in quantities which are not characteristic of natural plants.

25

One of skill will recognize that after the expression cassette is stably incorporated in transgenic plants and confirmed to be operable, it can be introduced into other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed.

30

Detection of RG Resistance Genes

The present invention further provides methods for detecting RG resistance genes in a nucleic acid sample suspected of comprising an RG resistance gene. The means by which the RG resistance gene is detected is not a critical aspect of the invention. For example, RG resistance genes can be detected by the presence of amplicons using RG resistance gene specific primers. Additionally, RG resistance genes can be detected by assaying for specific hybridization of an RG polynucleotide to an RG resistance gene. In some embodiments, the RG resistance gene can be amplified prior to the step of contacting the nucleic acid sample with the RG polynucleotide.

In a typical detection method, the nucleic acid sample is contacted with an RG polynucleotide to form a hybridization complex. The hybridization complex may be detected directly (e.g., in Southern or northern blots), or indirectly (e.g., by subsequent primer extension during PCR amplification). The RG polynucleotide hybridizes under stringent conditions to an RG polynucleotide of the invention. Formation of the hybridization complex is directly or indirectly used to indicate the presence of the RG resistance gene in the nucleic acid sample.

Detection of the hybridization complex can be achieved using any number of well known methods. For example, the nucleic acid sample, or a portion thereof, may be assayed by hybridization formats including but not limited to, solution phase, solid phase, mixed phase, or *in situ* hybridization assays. Briefly, in solution (or liquid) phase hybridizations, both the target nucleic acid and the probe or primer are free to interact in the reaction mixture. In solid phase hybridization assays, probes or primers are typically linked to a solid support where they are available for hybridization with target nucleic in solution. In mixed phase, nucleic acid intermediates in solution hybridize to target nucleic acids in solution as well as to a nucleic acid linked to a solid support. In *in situ* hybridization, the target nucleic acid is liberated from its cellular surroundings in such as to be available for hybridization within the cell while preserving the cellular morphology for subsequent interpretation and analysis. The following articles provide an overview of the various hybridization assay formats: Singer *et al.*, *Biotechniques* 4(3):230-250 (1986); Haase *et al.*, *Methods in Virology*, Vol. VII, pp. 189-226 (1984); Wilkinson, "The theory and practice of *in situ* hybridization" In: *In situ Hybridization*, Ed. D.G. Wilkinson. IRL Press, Oxford University Press, Oxford; and *Nucleic Acid Hybridization: A Practical Approach*, Ed. Hames, B.D. and Higgins, S.J., IRL Press (1987).

The effect of the modification of RG gene expression can be measured by detection of increases or decreases in mRNA levels using, for instance, Northern blots. In addition, the phenotypic effects of gene expression can be detected by measuring nematode, fungal, bacterial, viral, or other pest resistance in plants. Suitable assays for determining pest resistance are well known. Micheltore and Crute, *Trans. Br. mycol. Soc.* 79(3): 542-546 (1982).

The means by which hybridization complexes are detected is not a critical aspect of the present invention and can be accomplished by any number of methods currently known or later developed. RG polynucleotides can be labeled by any one of several methods typically used to detect the presence of hybridized nucleic acids. One common method of detection is the use of autoradiography using probes labeled with ^3H , ^{125}I , ^{35}S , ^{14}C , or ^{32}P , or the like. The choice of radioactive isotope depends on research preferences due to ease of synthesis, stability, and half lives of the selected isotopes. Other labels include ligands which bind to antibodies labeled with fluorophores, chemiluminescent agents, and enzymes. Alternatively, probes can be conjugated directly with labels such as fluorophores, chemiluminescent agents or enzymes. The choice of label depends on sensitivity required, ease of conjugation with the probe, stability requirements, and available instrumentation. Labeling the RG polynucleotide is readily achieved such as by the use of labeled PCR primers.

The choice of label dictates the manner in which the label is bound to the probe. Radioactive probes are typically made using commercially available nucleotides containing the desired radioactive isotope. The radioactive nucleotides can be incorporated into probes, for example, by using DNA synthesizers, by nick translation with DNA polymerase I, by tailing radioactive DNA bases to the 3' end of probes with terminal deoxynucleotidyl transferase, by treating single-stranded M13 plasmids having specific inserts with the Klenow fragment of DNA polymerase in the presence of radioactive deoxynucleotides, dNTP, by transcribing from RNA templates using reverse transcriptase in the presence of radioactive deoxynucleotides, dNTP, or by transcribing RNA from vectors containing specific RNA viral promoters (e.g., SP6 promoter) using the corresponding RNA polymerase (e.g., SP6 RNA polymerase) in the presence of radioactive ribonucleotides rNTP.

The probes can be labeled using radioactive nucleotides in which the isotope resides as a part of the nucleotide molecule, or in which the radioactive component is attached to the nucleotide via a terminal hydroxyl group that has been esterified to a radioactive component such as inorganic acids, *e.g.*, ³²P phosphate or ¹⁴C organic acids, or esterified to provide a linking group to the label. Base analogs having nucleophilic linking groups, such as primary amino groups, can also be linked to a label.

Non-radioactive probes are often labeled by indirect means. For example, a ligand molecule is covalently bound to the probe. The ligand then binds to an anti-ligand molecule which is either inherently detectable or covalently bound to a detectable signal system, such as an enzyme, a fluorophore, or a chemiluminescent compound. Enzymes of interest as labels will primarily be hydrolases, such as phosphatases, esterases and glycosidases, or oxidoreductases, particularly peroxidases. Fluorescent compounds include fluorescein and its derivatives, rhodamine and its derivatives, dansyl, umbelliferone, etc. Chemiluminescers include luciferin, and 2,3-dihydrophthalazinediones, *e.g.*, luminol. Ligands and anti-ligands may be varied widely. Where a ligand has a natural anti-ligand, namely ligands such as biotin, thyroxine, and cortisol, it can be used in conjunction with its labeled, naturally occurring anti-ligands. Alternatively, any haptenic or antigenic compound can be used in combination with an antibody.

Probes can also be labeled by direct conjugation with a label. For example, cloned DNA probes have been coupled directly to horseradish peroxidase or alkaline phosphatase, (Renz. M., and Kurz, K. (1984) A Colorimetric Method for DNA Hybridization. *Nucl. Acids Res.* 12: 3435-3444) and synthetic oligonucleotides have been coupled directly with alkaline phosphatase (Jablonski, E., *et al.* (1986) Preparation of Oligodeoxynucleotide-Alkaline Phosphatase Conjugates and Their Use as Hybridization Probes. *Nuc. Acids. Res.* 14: 6115-6128; and Li P., *et al.* (1987) Enzyme-linked Synthetic Oligonucleotide probes: Non-Radioactive Detection of Enterotoxigenic *Escherichia Coli* in Faeca Specimens. *Nucl. Acids Res.* 15:5275-5287).

Definitions

Units, prefixes, and symbols can be denoted in their SI accepted form. Numeric ranges are inclusive of the numbers defining the range. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation, respectively. The

headings provided herein are not limitations of the various aspects or embodiments of the invention which can be had by reference to the specification as a whole. Accordingly, the terms defined immediately below are more fully defined by reference to the specification as a whole.

5 As used herein, the term "plant" includes reference to whole plants, plant organs (e.g., leaves, stems, roots, etc.), seeds and plant cells and progeny of same. The class of plants which can be used in the methods of the invention is generally as broad as the class of higher plants amenable to transformation techniques, including both monocotyledonous and dicotyledonous plants.

10 As used herein, "pest" includes, but is not limited to, viruses, fungi, nematodes, insects, and bacteria.

 As used herein, "heterologous" is a nucleic acid that originates from a foreign species, or, if from the same species, is substantially modified from its original form. For example, a promoter operably linked to a heterologous structural gene is from a species different from that from which the structural gene was derived, or, if from the same species, one or both are substantially modified from their original form.

 As used herein, "RG gene," alternatively referred to as "RLG gene," is a gene encoding resistance to plant pests, such as viruses, fungi, nematodes, insects, and bacteria, and which hybridizes under stringent conditions and/or has at least 60% sequence identity at the deduced amino acid level to the exemplified sequences provided herein. RG genes encode "RG polypeptides," alternatively referred to as "RLG polypeptides," which can comprise LRR motifs and/or NBS motifs. The RG polypeptides encoded by RG genes have at least 55% or 60% sequence identity, typically at least 65% sequence identity, preferably at least 70% sequence identity, often at least 75% sequence identity, more preferably at least 80% sequence identity, and most preferably at least 90% sequence identity at the deduced amino acid level relative to the exemplary RG sequences provided herein. The term "RG family" or "RG family genus" or "genus" includes reference to a group of RG polypeptide sequence species that have at least 60% amino acid sequence identity, and, the nucleic acids encoding these polypeptides. The individual species of a genus, i.e., the members of a family, typically are genetically mapped to the same locus.

 As used herein, "RG polynucleotide" includes reference to a contiguous sequence from an RG gene of at least 18, 20, 25, 30, 40, or 50 nucleotides in length, up to

at least about 100 or at least about 200 nucleotides in length. In some embodiments, the polynucleotide is preferably at least 100 nucleotides in length, more preferably at least 200 nucleotides in length, most preferably at least 500 nucleotides in length. Thus, RG polynucleotide may be a RG gene or a subsequence thereof.

5 As used herein, "isolated," when referring to a molecule or composition, such as, for example, an RG polypeptide or nucleic acid, means that the molecule or composition is separated from at least one other compound, such as a protein, other nucleic acids (*e.g.*, RNAs), or other contaminants with which it is associated *in vivo* or in its naturally occurring state. Thus, an RG polypeptide or nucleic acid is considered isolated
10 when it has been isolated from any other component with which it is naturally associated, *e.g.*, cell membrane, as in a cell extract. An isolated composition can, however, also be substantially pure. An isolated composition can be in a homogeneous state and can be in a dry or an aqueous solution. Purity and homogeneity can be determined, for example, using analytical chemistry techniques such as polyacrylamide gel electrophoresis (SDS-
15 PAGE) or high performance liquid chromatography (HPLC).

 The term "nucleic acid" or "nucleic acid molecule" or "nucleic acid sequence" refers to a deoxyribonucleotide or ribonucleotide oligonucleotide in either single- or double-stranded form. The term encompasses nucleic acids, *i.e.*, oligonucleotides, containing known analogues of natural nucleotides which have similar or
20 improved binding properties, for the purposes desired, as the reference nucleic acid. The term also includes nucleic acids which are metabolized in a manner similar to naturally occurring nucleotides or at rates that are improved thereover for the purposes desired. The term also encompasses nucleic-acid-like structures with synthetic backbones. DNA backbone analogues provided by the invention include phosphodiester, phosphorothioate, phosphorodithioate, methylphosphonate, phosphoramidate, alkyl phosphotriester,
25 sulfamate, 3'-thioacetal, methylene(methylimino), 3'-N-carbamate, morpholino carbamate, and peptide nucleic acids (PNAs); see *Oligonucleotides and Analogues, a Practical Approach*, edited by F. Eckstein, IRL Press at Oxford University Press (1991); *Antisense Strategies*, Annals of the New York Academy of Sciences, Volume 600, Eds. Baserga and Denhardt (NYAS 1992); Milligan (1993) *J. Med. Chem.* 36:1923-1937; *Antisense Research and Applications* (1993, CRC Press). PNAs contain non-ionic backbones, such
30 as N-(2-aminoethyl) glycine units. Phosphorothioate linkages are described in WO

97/03211; WO 96/39154; Mata (1997) *Toxicol Appl Pharmacol* 144:189-197. Other synthetic backbones encompassed by the term include methyl-phosphonate linkages or alternating methylphosphonate and phosphodiester linkages (Strauss-Soukup (1997) *Biochemistry* 36:8692-8698), and benzylphosphonate linkages (Samstag (1996) *Antisense Nucleic Acid Drug Dev* 6:153-156). The term nucleic acid is used interchangeably with gene, cDNA, mRNA, oligonucleotide primer, probe and amplification product. Unless otherwise indicated, a particular nucleic acid sequence includes the complementary sequence thereof.

The term "exogenous nucleic acid" refers to a nucleic acid that has been isolated, synthesized, cloned, ligated, excised in conjunction with another nucleic acid, in a manner that is not found in nature, and/or introduced into and/or expressed in a cell or cellular environment other than or at levels or forms different than the cell or cellular environment in which said nucleic acid or protein is found in nature. The term encompasses both nucleic acids originally obtained from a different organism or cell type than the cell type in which it is expressed, and also nucleic acids that are obtained from the same cell line as the cell line in which it is expressed. invention.

The term "recombinant," when used with reference to a cell, or to the nucleic acid, protein or vector refers to a material, or a material corresponding to the natural or native form of the material, that has been modified by the introduction of a new moiety or alteration of an existing moiety, or is identical thereto but produced or derived from synthetic materials. For example, recombinant cells express genes that are not found within the native (non-recombinant) form of the cell or express native genes that are otherwise expressed at a different level, typically, under-expressed or not expressed at all. The term "recombinant means" encompasses all means of expressing, *i.e.*, transcription or translation of, an isolated and/or cloned nucleic acid *in vitro* or *in vivo*. For example, the term "recombinant means" encompasses techniques where a recombinant nucleic acid, such as a cDNA encoding a protein, is inserted into an expression vector, the vector is introduced into a cell and the cell expresses the protein. "Recombinant means" also encompass the ligation of nucleic acids having coding or promoter sequences from different sources into one vector for expression of a fusion protein, constitutive expression of a protein, or inducible expression of a protein, such as the plant disease resistant, or RG. polypeptides of the invention.

The term "specifically hybridizes" refers to a nucleic acid that hybridizes, duplexes or binds to a particular target DNA or RNA sequence. The target sequences can be present in a preparation of total cellular DNA or RNA. Proper annealing conditions depend, for example, upon a nucleic acid's, such as a probe's length, base composition, and the number of mismatches and their position on the probe, and can be readily determined empirically providing the appropriate reagents are available. For discussions of nucleic acid probe design and annealing conditions, see, *e.g.*, Sambrook and Ausubel.

The terms "stringent hybridization," "stringent conditions," or "specific hybridization conditions" refers to conditions under which an oligonucleotide (when used, for example, as a probe or primer) will hybridize to its target subsequence, such as an RG nucleic acid in an expression vector of the invention but not to a non-RG sequence. Stringent conditions are sequence-dependent. Thus, in one set of stringent conditions an oligonucleotide probe will hybridize to only one specie of the genus of RG nucleic acids of the invention. In another set of stringent conditions (less stringent) an oligonucleotide probe will hybridize to all species of the invention's genus but not to non-RG nucleic acids. Longer sequences hybridize specifically at higher temperatures. Stringent conditions are selected to be about 5⁰C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength, pH, and nucleic acid concentration) at which 50% of the probes complementary to the target sequence hybridize to the target sequence at equilibrium (if the target sequences are present in excess, at T_m , 50% of the probes are occupied at equilibrium). Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, *i.e.*, about 0.01 to 1.0 M sodium ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30⁰C for short probes (*e.g.*, 10 to 50 nucleotides) and at least about 60⁰C for long probes (*e.g.*, greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Often, high stringency wash conditions preceded by low stringency wash conditions to remove background probe signal. An example of medium stringency wash conditions for a duplex of, *e.g.*, more than 100 nucleotides, is 1x SSC at 45⁰C for 15 minutes (see Sambrook for a description of SSC buffer). An example low stringency wash for a duplex of, *e.g.*, more than 100 nucleotides, is 4-6x SSC at 40⁰C for 15 minutes. a signal to noise ratio of 2x (or higher) than that observed for an unrelated

probe in the particular hybridization assay indicates detection of a "specific hybridization." Nucleic acids which do not hybridize to each other under stringent conditions can still be substantially identical if the polypeptides which they encode are substantially identical. This can occur, *e.g.*, when a nucleic acid is created that encodes for conservative
5 substitutions. Stringent hybridization and stringent hybridization wash conditions are different under different environmental parameters, such as for Southern and Northern hybridizations. An extensive guide to the hybridization of nucleic acids is found in, *e.g.*, Sambrook, Tijssen (1993) *supra*.

As used herein "operably linked" includes reference to a functional linkage
10 between a promoter and a second sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. Generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two protein coding regions, contiguous and in the same reading frame.

15 In the expression of transgenes one of skill will recognize that the inserted polynucleotide sequence need not be identical and may be "substantially identical" to a sequence of the gene from which it was derived. As explained herein, these variants are specifically covered by this term.

In the case where the inserted polynucleotide sequence is transcribed and
20 translated to produce a functional RG polypeptide, one of skill will recognize that because of codon degeneracy, a number of polynucleotide sequences will encode the same polypeptide. These variants are specifically covered by the term "RG polynucleotide sequence". In addition, the term specifically includes those full length sequences substantially identical (determined as described herein) with an RG gene sequence which
25 encode proteins that retain the function of the RG protein. Thus, in the case of RG genes disclosed here, the term includes variant polynucleotide sequences which have substantial identity with the sequences disclosed here and which encode proteins capable of conferring resistance to nematodes, bacteria, viruses, fungi, insects or other pests on a transgenic plant comprising the sequence.

30 Two polynucleotides or polypeptides are said to be "identical" if the sequence of nucleotides or amino acid residues, respectively, in the two sequences is the same when aligned for maximum correspondence, as described below. The term

"complementary to" is used herein to mean that the complementary sequence is identical to all or a specified contiguous portion of a reference polynucleotide sequence.

The terms "sequence identity," "sequence similarity" and "homology" refer to when two sequences, such as the nucleic acid and amino acid sequences or the polypeptides of the invention, when optimally aligned, as with, for example, the programs PILEUP, BLAST, GAP, FASTA or BESTFIT (see discussion, *supra*). "Percentage amino acid/nucleic acid sequence identity" refers to a comparison of the sequences of two polypeptides/nucleic acids which, when optimally aligned, have approximately the designated percentage of the same amino acids/nucleic acids, respectively. For example, "60% sequence identity" and "60% homology" refer to a comparison of the sequences of two RG nucleic acids or polypeptides which, when optimally aligned, have 60% identity. For example, in one embodiment, nucleic acids encoding RG polypeptides of the invention comprise a sequence with at least 50% nucleic acid sequence identity to SEQ ID NO:1. In other embodiments, the RG polypeptides of the invention are encoded by nucleic acids comprising a sequence with at least 50% sequence identity to SEQ ID NO:1, or, are encoded by nucleic acids comprising SEQ ID NO:1, or, have at least 60% amino acid sequence identity to the polypeptide of SEQ ID NO:2.

"Percentage of sequence identity" is determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.

The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 55% or 60% sequence identity, generally at least 65%, preferably at least 70%, often at least 75%, more preferably at least 80% and most preferably at least 90%, compared to a reference sequence using the programs described above (preferably BESTFIT) using standard parameters. One of skill will recognize that these values can be appropriately adjusted to determine corresponding

identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 55% or 60%, preferably at least 70%, more preferably at least 80%, and most preferably at least 95%. Polypeptides having "sequence similarity" share sequences as noted above except that residue positions which are not identical may differ by conservative amino acid changes. Conservative amino acid substitutions refer to the interchangeability of residues having similar side chains. For example, a group of amino acids having aliphatic side chains is glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains is serine and threonine; a group of amino acids having amide-containing side chains is asparagine and glutamine; a group of amino acids having aromatic side chains is phenylalanine, tyrosine, and tryptophan; a group of amino acids having basic side chains is lysine, arginine, and histidine; and a group of amino acids having sulfur-containing side chains is cysteine and methionine. Preferred conservative amino acids substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, and asparagine-glutamine.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under appropriate conditions. Appropriate conditions can be high or low stringency and will be different in different circumstances. Generally, stringent conditions are selected to be about 5°C to about 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typically, stringent wash conditions are those in which the salt concentration is about 0.02 molar at pH 7 and the temperature is at least about 50°C. However, nucleic acids which do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides which they encode are substantially identical. This may occur, *e.g.*, when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code.

Nucleic acids of the invention can be identified from a cDNA or genomic library prepared according to standard procedures and the nucleic acids disclosed here used as a probe. Thus, for example, stringent hybridization conditions will typically include at least one low stringency wash using 0.3 molar salt (*e.g.*, 2X SSC) at 65°C. The washes

are preferably followed by one or more subsequent washes using 0.03 molar salt (e.g., 0.2X SSC) at 50°C, usually 60°C, or more usually 65°C. Nucleic acid probes used to identify the nucleic acids are preferably at least 100 nucleotides in length.

As used herein, "nucleotide binding site" or "nucleotide binding domain" ("NBS") includes reference to highly conserved nucleotide-, *i.e.*, ATP/GTP-, binding domains, typically included in the "kinase domain" of kinase polypeptides, such as a kinase-1a, kinase 2, or a kinase 3a motif, as described herein. For example, the tobacco N and Arabidopsis RPS2 genes, among several recently cloned disease-resistance genes, share highly conserved NBS sequence. Kinase NBS subdomains further consist of three subdomain motifs: the P-loop, kinase-2, and kinase-3a subdomains (Yu (1996) *Proc. Acad. Sci. USA* 93:11751-11756). As discussed in detail herein, examples include the *Arabidopsis* RPP5 gene (Parker (1997) *supra*), the *A. thaliana* RPS2 gene (Mindrinos (1997) *supra*), and the flax L6 rust resistance gene (Lawrence (1995) *supra*) which all encode proteins containing an NBS; and Mindrinos (1994) *Cell* 78:1089-1099; and Shen (1993) *FEBS* 335:380-385. Using the teachings disclosed and incorporated herein and standard nucleic acid hybridization and/or amplification techniques, one of skill can identify members having NBS domains, including any of the genus of NBS-containing plant disease resistant polypeptides of the invention.

As used herein, "leucine rich region" ("LRR") includes reference to a region that has a leucine content of at least 20% leucine or isoleucine, or 30% of the aliphatic residues: leucine, isoleucine, methionine, valine, and phenylalanine, and arranged with approximate repeated periodicity. The length of the repeat may vary in length but is generally about 20 to 30 amino acids. An LRR-containing polypeptide typically will have the canonical 24 amino acid leucine-rich repeat (LRR) sequence, which is present in different proteins that mediate molecular recognition and/or interaction processes; as described in Bent (1994) *Science* 265:1856-1860; Parker (1997) *Plant Cell* 9:879-894; Hong (1997) *Plant Physiol.* 113:1203-1212; Schmitz (1997) *Nucleic Acids Res.* 25:756-763; Hipskind (1996) *Mol. Plant Microbe Interact.* 9:819-825; Tornero (1996) *Plant J.* 10:315-330; Dixon (1996) *Cell* 84:451-459; Jones (1994) *Science* 266:789-793; Lawrence (1995) *Plant Cell* 7:1195-1206; Song (1995) *Science* 270:1804-1806; as discussed in further detail *supra*. Using the teachings disclosed and incorporated herein and standard nucleic acid hybridization and/or amplification techniques, one of skill can

identify polypeptides having LRR domains, including any member of the genus of LRR-containing RG polypeptides of the invention.

The term "promoter" refers to a region or sequence determinants located upstream or downstream from the start of transcription and which are involved in recognition and binding of RNA polymerase and other proteins to initiate transcription. A "plant promoter" is a promoter capable of initiating and/or regulating transcription in plant cells; see also discussion on plant promoters, *supra*.

The term "constitutive promoter" refers to a promoter that initiates and helps control transcription in all tissues. Promoters that drive expression continuously under physiological conditions are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation; see also detailed discussion, *supra*.

The term "inducible promoter" refers to a promoter which directs transcription under the influence of changing environmental conditions or developmental conditions. Examples of environmental conditions that may effect transcription by inducible promoters include anaerobic conditions, elevated temperature, drought, or the presence of light. Such promoters are referred to herein as "inducible" promoters; see also detailed discussion, *supra*.

The term "abscission-induced promoter" or "abscission promoter" refers to a class of promoters which are activated upon plant ripening, such as fruit ripening, and are especially useful incorporated in the expression systems (*e.g.*, expression cassettes, vectors) of the invention. When the plant disease resistant polypeptide-encoding nucleic acid is under the control of an abscission promoter, rapid cell death, induced by expression of the invention's polypeptide, accelerates and/or accentuates abscission of the plant part, increasing the efficiency of the harvesting of fruits or other plant parts, such as cotton, and the like; see also detailed discussion, *supra*.

The term "tissue-specific promoter" refers to a class of transcriptional control elements that are only active in particular cells or tissues. Examples of plant promoters under developmental control include promoters that initiate transcription only (or primarily only) in certain tissues, such as roots, leaves, fruit, ovules, seeds, pollen, pistols, or flowers; see also detailed discussion, *supra*.

As used herein "recombinant" includes reference to a cell, or nucleic acid, or vector, that has been modified by the introduction of a heterologous nucleic acid or the alteration of a native nucleic acid to a form not native to that cell, or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found within the native (non-recombinant) form of the cell or express native genes that are otherwise abnormally expressed, under expressed or not expressed at all.

As used herein, a "recombinant expression cassette" or "expression cassette" is a nucleic acid construct, generated recombinantly or synthetically, with a series of specified nucleic acid elements which permit transcription of a particular nucleic acid in a target cell. The expression vector can be part of a plasmid, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of the expression vector includes a nucleic acid to be transcribed, and a promoter.

As used herein, "transgenic plant" includes reference to a plant modified by introduction of a heterologous polynucleotide. Generally, the heterologous polynucleotide is an RG structural or regulatory gene or subsequences thereof.

As used herein, "hybridization complex" includes reference to a duplex nucleic acid sequence formed by selective hybridization of two single-stranded nucleic acids with each other.

As used herein, "amplified" includes reference to an increase in the molarity of a specified sequence. Amplification methods include the polymerase chain reaction (PCR), the ligase chain reaction (LCR), the transcription-based amplification system (TAS), the self-sustained sequence replication system (SSR). A wide variety of cloning methods, host cells, and *in vitro* amplification methodologies are well-known to persons of skill.

As used herein, "nucleic acid sample" includes reference to a specimen suspected of comprising RG resistance genes. Such specimens are generally derived, directly or indirectly, from lettuce tissue.

The term "antibody" refers to a polypeptide substantially encoded by an immunoglobulin gene or immunoglobulin genes, or fragments or synthetic or recombinant analogues thereof which specifically bind and recognize analytes and antigens, such as a genus or subgenus of polypeptides of the invention, as described *supra*.

It is understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and scope of the appended claims.

5

EXAMPLES

The following examples are offered to illustrate, but not to limit the claimed invention.

Example 1

10 Example 1 describes the use of PCR to amplify RG genes from lettuce.

Multiple primers with low degeneracy, particularly at the 3' end, were designed based on the sequences of two known resistance genes from tobacco and flax.

DNA Templates

15 Lettuce genomic DNA was extracted from cultivar Diana and a mutant line derived from cultivar Diana using a standard CTAB protocol. To generate cDNA templates, RNA was isolated from cultivar Diana and the mutant following standard procedures; first strand cDNA was synthesized using Superscript reverse transcriptase from 1 Φ g total RNA as specified by the manufacturer (Life Technologies). BAC (bacterial artificial chromosome) clones from the *Dm3* region were isolated from a BAC library of
20 over 53,000 clones using marker AC15 that was known to be closely linked to *Dm3*. Bacterial plasmids containing clones of *L6* and *RPS2* were used as positive controls.

PCR with degenerate oligonucleotide primers

25 Oligonucleotide primers were designed based on conserved motifs in the nucleotide binding sites (NBS) of *L6*, *RPS2*, and *N*. Eight primers were made corresponding to the GVGKTT motif in the sense direction; each had 64-fold degeneracy. Six primers were made to the GLPLAL motif in the anti-sense direction; with either 16 or 256-fold degeneracy (Table 1).

30 Oligonucleotides included 14-mer adaptors of (CUA)₄ at the 5' end of the sense primers and (CAU)₄ at the 5' end of the antisense primers to allow rapid cloning of the PCR products into pAMP1 (Life Technologies).

PCR amplification was performed in 50 Φ l reaction volume with 1 Φ M of each of a pair of sense and antisense primers. The templates were denatured by heating to 94EC for 2 min. This was followed by 35 cycles of 30 sec at 94EC, 1 min at 50EC, 2 min at 72EC, with a single final extension of 5 min at 72EC. 25 ng of genomic DNA or cDNA was used. BAC clones as templates required less. The final dNTP concentration was 0.2 mM; $MgCl_2$ was 1.5 mM.

Forty-eight combinations of sense and antisense primers were tested on a panel of nine templates consisting of two genomic DNA samples, two cDNA preparations, three BAC clones and plasmids containing *L6* and *RPS2* as positive controls.

Amplification from *L6* and *RPS2* resulted in fragments of 516 and 513 repectively. Seven combinations of primers resulted in fragments of approximately this size with multiple templates (Table 2). Primers that gave RLG products were: PLOOPAA, PLOOPAG, PLOOPGA, PLOOPGG, PLOOPAC, GLPL3, GLPL4.

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Table 1

DEGENERATE PRIMER SEQUENCES for NBS PCR

Sense primers based on GVGKTT amino acid sequence from L6, N and rps2 PLOOP motif:

PLOOPAG 5' GGN GTN GGN AAA ACG AC 3'

PLOOPAA 5' GGN GTN GGN AAA ACA AC 3'

PLOOPAT 5' GGN GTN GGN AAA ACT AC 3'

PLOOPAC 5' GGN GTN GGN AAA ACC AC 3'

PLOOPGG 5' GGN GTN GGN AAG ACG AC 3'

PLOOPGA 5' GGN GTN GGN AAG ACA AC 3'

PLOOPGT 5' GGN GTN GGN AAG ACT AC 3'

PLOOPGC 5' GGN GTN GGN AAG ACC AC 3'

Antisense primers based on GLPLAL amino acid sequence:

GLPL1 5' AGN GCN AGN GGN AGG CC 3'

GLPL2 5' AGN GCN AGN GGN AGA CC 3'

GLPL3 5' AGN GCN AGN GGN AGT CC 3'

GLPL4 5' AGN GCN AGN GGN AGC CC 3'

GLPL5 5' AAN GCC AAN GGC AAA CC 3'

GLPL6 5' AAN GCC AAN GGC AAT CC 3'

TABLE 2. Characteristics of RLGs isolated from lettuce.

	Template	Primers	Number ^a	Size ^b (bp)	Copy number ^c	Dm linkage
5	RLG1	genomic DNA	PLOOPGA+GLPL6	6/6	522	DM4,
		cDNA	PLOOPGA+GLPL6	1/5		DM13
		genomic DNA	PLOOPAA+GLPL6	5/5		
		cDNA	PLOOPAA+GLPL6	1/1		
	RLG2	BACH8	PLOOPGG+GLPL3	3/3	510	DM1, Dm3
	RLG3	genomic DNA	PLOOPGA+GLPL4	3/6	461	Dm5 Dm8
10	RLG4	genomic DNA	PLOOPGA+GLPL4	1/6	524	

^a Number of RLG sequences out of total number of clones sequenced.

^b Size of fragment amplified from the nucleotide binding domain.

^c Estimated copy number from genomic Southern blot analysis and numbers of clones in the BAC library.

Example 2

Example 2 describes the genetic analysis used to obtain a preliminary indication of the linkage relationships of the amplified products and known clusters of resistance genes.

Bulked segregant analysis was performed to obtain a preliminary indication of the linkage relationships of the amplified products and known clusters of resistance genes. DNA from individuals were pooled for each susceptible and resistant bulk. Amplified products were then mapped by RFLP analysis from our intraspecific mapping population. Resistances from four clusters of resistance genes as well as over six hundred markers have now been mapped on this population. Linkage analysis was done using JIONMAP or MAPMAKER mapping programs. Due to a suppression of recombination in the *Dm3* region, sequences were mapped relative to *Dm3* using a panel of deletion mutants that provided greater genetic resolution than the mapping population (Anderson *et al.* 1996). All blots were washed twice at 63EC in 2x SSC/1% SDS for 20 min, followed by one wash at 63EC in 1x SSC/0.1% SDS for 10 or 30 min.

Most of the RLG sequences were analyzed by bulked segregant analysis (BSA) using pools of resistant and susceptible individuals for each of the four clusters of resistance genes. In genomic Southern analyses, all the RLGs revealed numerous fragments of varying intensity. The numbers of bands was highly dependent of the stringency of hybridization. BSA demonstrated that RLG1 was linked to the *Dm4*, 7 and *Dm13* clusters. Segregation analysis confirmed this linkage.

RLG2 was derived from BAC H8 that was known to be from the *Dm3* region. BSA with RLG2 demonstrated that the polymorphic bands that distinguished the parents of our mapping population mapped to the *Dm1*, *Dm3* cluster. Several bands absolutely cosegregated with *Dm1* or *Dm3*. To provide finer genetic resolution, RLG2 was also mapped using a panel of *Dm3* deletion mutants. A number of fragments were missing in largest deletion mutant demonstrating that several RLG2 family members are physically located very close to *Dm3*. No fragment was missing in all deletion mutants; however, this is not unexpected as there is extensive duplication within the region.

Example 3

Example 3 describes the screening of a bacterial artificial chromosome library.

Over 53,000 BAC clones containing lettuce genomic DNA were screened with two of the amplified products. High density filters each containing 1536 clones were hybridized to ³²P labelled probes. Filters were washed at 65°C with 40 mM Na₂PO₄/0.1% SDS for 5 min followed by 20 min in the same solution.

To isolate additional RLG sequences we screened our genomic BAC library. Clones were identified that hybridized to RLG1 and RLG2. Nearly all the clones that hybridized to RLG2 also hybridized to marker AC15 that had already been shown by deletion mutant analysis to be clustered around *Dm3*. This provided further evidence for clustering of RLG2 sequences.

Using primers conserved within each family, part of the NBS was amplified from each unique BAC clone and sequenced. This revealed that members within each family varied from 64% identical at the deduced amino acid level. The most divergent members only weakly cross-hybridized to each other. Currently, RLG sequences are

considered to be part of the same family of sequences if they are at least 55% identical at the deduced amino acid level and map to the same region of the chromosome.

Example 4:

5 Example 4 describes the cloning, identification, sequencing and characterization of RG polynucleotide sequences; including use of RG sequences from plasmid and PCR products.

 Doubled stranded plasmid DNA clones and PCR products were sequenced using an ABI377 automated sequencer and fluorescently labelled di-deoxy terminators.
10 Sequences were assembled using Sequencher (Genecodes), DNASTar (DNASTar) and Genetics Computer Group (GCG, Madison, WI) software. Database searches were performed using BLASTX and FASTA (GCG) algorithms.

 Sequences flanking the NBS region for RLG2 and for some of RLG1 were obtained by a series of IPCR and the products sequenced directly. IPCR worked less well
15 for RLG1. Therefore RLG1 was subcloned from a BAC clone into pBSK (Stratagene) and the double stranded plasmid sequenced by long range sequencing.

 Initially, a total of 30 clones were sequenced. Three of these seven primer combinations yielded sequences that comprised continuous open reading frames with sequence identity to the NBS of known resistance genes. Seven out of 10 clones amplified
20 from genomic DNA with the primer pair PLOOPGA/GLP6 were 522 bp long; they were identical to each other and named RLG1. All six clones amplified from genomic DNA or cDNA using the primers PLOOPAA/GLP6 were similar/the same as RLG1. All three clones sequenced from BAC clone H8 were 510 bp long, identical to each other but different from RLG1 and were therefore designated RLG2. The 11 clones sequenced from
25 four other primer combinations had no similarity to any NBS motifs and therefore were not studied further. Therefore, sequencing resulted in the identification of clones containing NBS motifs representing four RLG sequences.

 Comparison of the deduced amino acid sequences of RLG1 and RLG2 to those of known resistance genes revealed that RLG1 and RLG2 are as similar to each other
30 as they are to resistance genes from other species and that this is the same level of identity shown between the known resistance genes (Table 3). The percent identity (upper quadrant) and percent identity (lower quadrant) were determined using the MEGALIGN

routine of the DNASTAR package. Identity refers to the proportion of identical amino acids; identity refers to the proportion of identical and similar amino acids and takes into account substitutions of amino acids with similar chemical characteristics. RG1 and RG2 are as similar to each other and to cloned resistance genes as cloned resistance genes from a variety of species are to each other. L6, resistance to *Melampsora lini* in flax (Lawrence *et al.*, 1995). N, resistance to tobacco mosaic virus in tobacco (Whitham *et al.*, 1994). PRF, required for resistance to *Pseudomonas syringae* in tomato. RPS2, resistance to *Pseudomonas syringae* in *Arabidopsis thaliana* (Bent *et al.*, 1994; Mindrinos *et al.*, 1994). RPM1, resistance to *Pseudomonas syringae* pv. *maculicola* in *A. thaliana* (Grant *et al.*, 1995). The initial RG1 and RG2, sequences were amplified from lettuce using degenerate primers.

Table 3

IDENTITIES OF

RESISTANCE GENE HOMOLOGUES

		RG1	RG2	RG3	RG4	N gene	RPS2
Lettuce	RG1	***	22.7	15.0	29.2	25.4	23.8
Lettuce	RG2		***	32.2	21.6	22.7	33.0
Lettuce	RG3			***	17.2	15.0	32.8
Lettuce	RG4				***	44.3	22.7
Tobacco	N gene					***	21.6
<i>Arabidopsis</i>	RPS2						***

The regions homologous to the primers are included in this analysis as the genomic sequences for RLG1 and RLG2 were determined by IPCR. Interestingly, the genomic sequences for RLG1 exactly matched that of the primers used.

To obtain further evidence that we had amplified resistance genes, we amplified the regions flanking the NBSs of RLG1a and RLG2a by IPCR of BAC clones. These products were then directly sequenced without cloning to minimize the introduction of PCR artifacts. Sequence analysis of the 5' regions failed to detect any homology to known resistance genes. However, the sequence of the 3' region contained leucine-rich

repeats (LRRs). When this sequence was used to search GENBANK using BLASTX, it detected identity to the *Arabidopsis* resistance gene, *RPS2*. This region does not contain as regular LRRs as in some resistance genes; however, the repeat structure seems to be consistent with that of the flax resistance gene, *L6*. Therefore, the presence of an LRR region is further evidence that the sequences we amplified using degenerate oligonucleotide primers are probably resistance genes.

The sequences of the IPCR products also provided the genomic sequences of the regions complementary to the sequences of the degenerate oligonucleotide primers.

The genomic sequences for RLG1 were identical to one of the primers in the mixture.

The RLG sequences are resistance genes as supported by three criteria: the presence of multiple sequence motifs characteristic of resistance genes, genetic cosegregation with known resistance genes, and their existence as clustered multi-gene families. The presence of LRR regions in a similar position relative to the NBS as in cloned resistance genes provides stronger evidence than relying solely sequence similarity between NBS regions.

The clustering of RLG sequences at the same position as the known clusters of resistance genes make them strong candidates for encoding resistance genes. The hybridization patterns and genetic distribution of the RLG sequences are similar to that of cloned resistance genes in other species. Most of these hybridize to small multigene families and preliminary genetic evidence indicates that they are clustered in the genome. Therefore, the degenerate primers that we designed from other resistance genes seemed to have been specific enough to amplify resistance genes rather than P-loop containing proteins in general.

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SEQ ID NO: 1

RLG1A
[Strand]

```
1  ATCGTAACCGTTGCTACGAG  ANCGCTGKCCCTCTTCATC  TTTTGTCATATGTCATATTC  TCATNIAITMTGCCACATNT
81  AATTTTGTGGTTATTTTAA  TTAATTTTATTCACATGT  CATTTTATGAGTTTTCAT  TTTATTGAGTTTCACATAAT
161  ATTTAAATGTAATAACAATA  AATGCATATTTTATTTTCTT  TAAATAAACGCATATAATAT  ATAGATTAAAAATCATATAAT
241  ACATAGGTTAAACATCATATA  ATACATATGTTTCATCCCCAG  TTTATTTATATGTCATCACC  TTAATTTTATTTATTTAT
321  TTATTAGAGTAGATCTT  TGIGATATTAATAATTTAAT  TTGTTCAAATTTAAATTA  TTAATAATCCCAATTTGA
401  ATAAAAATTAAAAAATGGN  CCCACCATTAGTCCATCACT  TTTTCAGCTCATCAATATCG  TGAGTATTCTCCCTCGTTTC
481  CACCCCAATCAATATTTCCA  GCGAATGACAGACTCCTACG  GCGTTTCTGAATTTGCGTTC  CGACACTGTTCATTGAAGGA
561  GATAATAAATCAAAATGGAGC  TGCTCCAAATGTTCAATGCTG  ATGAAAGGTGAATTTGATGT  GAAGANAATGTCAGCGATCN
641  ATCTCCATCCGGAACCCACC  ACATTATCAGTGTACCACCA  AACCACTCAAACCGYGGAA  GTAGRRAKACWRKAAAGTCA
721  TGAAGAATAGATTATTTTIG  TCCTCATGGGCTGACTGAGG  AGCGGGTTTGTTCATCATT  TTTCTTTGANCAAGAATTA
801  TCGGTCCATCGAATTTTAC  ATCGACAAAGAAGTTTCACT  TCGCAATGTTTGTAAACA  ATTTTAAATCTTTTATCTT
881  TTCGTTGAAACTCCTCAATT  GCAACTTGCAACTTGCAACT  TTTGGGCCCACTAATTTGTG  GTGGCGTTAATTTAATCCA
961  CATATTCACTGTAAACAATA  ATTCAAATCGATCTCTGTTC  ATCCAATTCATCAACATCTC  TTGATAATTGAATCATCTCA
1041  CGCTTCATCCATTTCATCCA  CATCTATACTATATCTCTG  CTCATATCATATTAAACGAT  GGCTGAAATCGTCTTCTG
1121  CCTCTCTGACAGTGGTGT  GAAAAGCTGGCATYTGAAGC  CTTGAAGAAGATTGTTGCTG  CCAAAGAATTGAATCTGAG
1201  CTTAAGAAATGAAGGAGAC  ATTAGACCAAATCCAAGATC  TGCTTAACGATGCTTCCAG  AAGGAAGTAACTAATGAAGC
1281  CGTTAAAGATGGCTGAATG  ATCTCCAACATTTGGCTTAT  GACATAGACGACCTACTTGA  TGATTTGCAACTGAAGCTG
1361  TTCACACGCTGAGTTGACCGAG  GAGGGTGGAGCCTCCTCCAG  TATGGTAAGAAAATAATCC  CAAGTTGTTGCACAAGTTTC
1441  TCACAAAGTAATAGGATGCA  TGCCAAGTTAGATGATATG  CCACCAGGTTACAGAACTG  GTAGAGGCAAAAATAATCT
1521  TGGTTTAAAGTGTGTAACAT  ATGAAAGCCAAAATTTGAA  AGGTATGAGGCGCTCTTGGT  AGATGAAAGCGGTACTGTCG
1601  GACGTGAAGATGATAAGAAA  AAATTGCTGGAGAAGCTGTT  GGGGATAAAGATGAATCAG  GGAGTCAAACCTTCAGCATC
1681  GTGCCCATAGTTGGTATGGG  TGGAGTTGGTAAACAACATC  TAGCTAGACTTTTGTATGAT  GAAAAGAAAGTGAAGGATCA
1761  CTTGCAACTCAGGGCTGGG  TTTGTGTTCTGATGAGTTC  AGTGTCCCAATATAAGCAG  AGTTATTTATCAATCTGTGA
1841  CTGGGGAAAAGAAGGAGTTT  GAAGACTTAAATCTGCTTCA  AGAAGCTCTTAAAGAGAAAC  TTAGGAACCAGCTATTCTTA
1921  ATAGTTTTGGATGATGTGTG  GTCTGAAAGCTATGTTGAT  GGGAGAAATTAGTGGGCCCA  TTCTTGGCGGGTCTCCCTG
2001  AAGTAGAATAATCATGACAA  CTCGGAAGGAGCAATTTGCT  AGAAAGCTGGGCTTTTCTCA  TCAAGACCCCTCTGGAGGGTC
2081  TATCACAAGATGATGCTTIG  TCCTTGTGTTGCTCAACACGC  ATTTGGTGTACCAAACTTTG  ATTCAATCCAACCTAAGG
2161  CCACATGGAGACTGTTTGT  GAAGAAATGTGATGGCTTAC  CTCTAGCTTTAAGAACACTT  GGAAGGTTATTAAAGACAAA
2241  AACAGACAGGGAACATGGA  AGGAGCTGTTGGATAGTGAG  ATATGGAGGTTAGGAAGAG  CGATGAGATTGTTCCGGCTC
2321  TTAGACTAAGCTACATGAT  CTTTCTGCCWCTTTGAAGCT  RTTTRTTGCAATATGCTCCT  TGTTTCCCAAGCATATGAG
2401  TTTGACAAGGAGGAGTTGAT  TCTATTGTGGATGGCAGAAG  GGTTTTTGCACCAACCACT  AYAAACAAGTCAAGCAAGC
2481  KTTGGGCTCTGAAATTTTTR  AAGAGTTTGTGCAAGRTCR  TTTTTCACATGCTCCTTAA  TRRCAATCSTTGTGTTGA
2561  TGCATGACCTAATGAATGAT  TTGGCTACATTGTTGCTGG  AGAATTTTTTCAAGGTTAG  ACATAGAGATGAAGAAGGAA
2641  TTTAGGATGSAATCTTTGGA  RAAGCACCGMCATATGTCAT  TTGTATGTGAGRATTACATA  GGTACAAAARGTTTCGAGCC
2721  ATTTAGAGGAGCTAAAAATT  TGAGAACATTTTAGCATTG  TCTGTTGGGGTGGTGAAGA  TTGGAAGATGTTTACTTTAT
2801  CAAACAAGGTCTTGAATGAC  WTACTTCARGATTTACCATT  GTTAAGGGTCTTRAKTTTGA  TTRRTCTTAYAAATAASYRAG
2881  GTACCAARAATCTGSGTAG  TATGAASCATTTGCGGTATC  TTAATCTATCWRGAACCTTWA  ATCACMCATTTACCGGAWA
2961  TKTCCTGCAATCTTTATAATT  TACARACCTGATTTGTTCT  GGCTGTGAMTATTTAGTTAA  KTTGCCCAARACCTTCTCAA
3041  ASCTTAAAAATTTGCASCAT  TTTGACATGAGGRTACTCC  KAAKTTAARAACATGCCCT  TARGGATTGGTGARTTGAAA
3121  ARTCTACAAACTCTCTTNG  TAACATTGGCATAGCAATAA  CCGAGCTTAAGAACTTGCAM  AAYCTCCATGGGAAARTTTG
3201  TATTGGCGGGCTGGGAAAAA  TGGAAATGCMGTGKGATGC  ACGTTAAGCGAACTTGTCTC  AAAAAAGGTTWAATGARTTA
3281  NAAACTGGRTWTTGGGGTGA  TFAATTTAATGTTTCCGAA  ATGGGAACACTTGAAAAAGA  AGTCTCTCAATGAAGTGATC
3361  CTCATAATGGTACTCTANAA  AAAACCCANAATTTATGTCTA  TAGGGGGTATAGAGTTTCCA  AATTGGGTTGGTTNCACATA
3441  GGGTTTCTGAACTAGAGAT  GTGTTCATGGTGTATGAAAA  AGANTGTTTACGTAGTTTC  ATCAATCACCAGTGGGAAA
3521  TAGATGATATTTTACGGGCT  TACTGATGAGATGTGGAGAG  GTATGATAGGGTTTCTTGGG  GCGGTAGAAGAAATAAGCAT
3601  CCATCTCTGTGAATGAAATAA  GATATYTTGTGGGAATCAGAA  GCAGAGGCAAGTAAGGTTCT  TATGAATTTAAAGAAGTTGG
3681  ATTTAGGTGAATGTGAAAT  TTGGTGAGTTTAGGGGAGAA  AAAGGAGGATAATCATATAA  TTAATAGTGGGAGCAGCCTA
3761  ACATCTTTTAGGAGGTTGAA  TGTATGGAGATGTAAACGCT  TGGAGCATTGCAAGGTGTCCA  GATGATCGGAGAAATTTGTA
3841  TATGCAATGTGTGATTCAA  THACATCCGCTCTCTCCCA  ACAGGAGGAGGACAGAAGAT  CAAGTCAATACCATCACTG
3921  ATTGCAAGAAGCTTTCCGAA  GAGGAGTTGGGAGGACGAGA  GAGGACAAGAGTCTTATAA  ACTCAAAAATGCGAGTCTT
4001  GAATCTAGATATACGTAA  TTGGCCAAATCTGAAATCTA  TCAGTGAATTTAGTTGCTTC  ATTCACCTGAACAGATTATA
4081  TATATCAACTGTCGAGTR  TGGAGTCAATTTCTGACCAT  GAGTTGCCAAATCTCACCTC  CTTACAGATCGAAGGAGAG
4161  GACAGCGATTTTCTGACGAA  CGGTTACGATTCGACTGGCC  GTCTTTTT
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SEQ ID NO: 2

RLG1B

[Strand]

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1  AACCGTTCGT ACCGAGATCG CTGTCTCTCT CTTCCTGTAA TATAATGATA AGAAAAAATA TGATTAAAGG
71  TTTAAATCCA AAATCCATTA TTCCACCGGT GATATGATGC ACTAGCTGTA GTATGCAAAA ACAGTATTAT
141 AAATGCTAAC CAAAACAGCA GCTAAGAAAC AATATAAATA ATGGTTTGAA TCGTCTTTTC TCCGTACAQT
211 CATTTCTTCC AAATCCCTAT CATTCATACA TACAAGTGCT CCCATATTAG GTTTTCACTA TAAGCAATGG
281 CTGAAATCCT TGGTTCGTGG TTCTTTGCGG TGTTCCTTGA AAAGCTTGCT TCTGAAGCCT TGAAGAGGGT
351 TGCTTGCTCC AAAGTAATTG ACAAGGAGCT CGAGAAATTG AATAGCTCAT GAATCAATAT AAAAGCTCTG
421 CTCATATGATG CTTCTCAGAA GGAATAAAGT AAGGAAGCTG TTAAGAATG GTTGAATGCT CTTCAACATT
491 TGCTTACGA CATAGATGAT CTACTTGGCG ATTTGGCAAC CAAAGCTATC CATCGTAAGT TCTCTGAGGA
561 ATACGGGGCC ACCATCAACA AGGTACGAAA GTTAAATCCA TCTTGTTTCT CTAGTTTGTC AAGTACTAAG
631 ATGCGCAACA AGATACATAA TATTACCAGC AAGTTACAAG AACTATTAGA AGAGAGAAAT AATCTTGGAT
701 TATGTGAAT TGGTGAAGC CGAAACTTTC GAAATAGAAA ATCAGAGACC TCCTTGCTAG ATCCATCTAG
771 TATTGTTGGA CGCAGATG ATAAGGAAGC GTTGCTTCTC AAGCTATATG AACCATGTGA TAGAACTTT
841 AGCATCTTGC CNATAGTTGG TATGGGTGGG TTAGATAAGA CCACTTTAGG TAGACTTTTG TATGATNAAA
911 TGCAAGTGAA GGATCACTTC GAACTCAAGG CGTGGGTTTG TGTTCCTGAT GAGTTTGATA TCTTCGGTAT
981 AAGCAAAACC ATTTTCGAAT CGATAGAGGG GGGAAACCAA GAGTTTAAGG ATTTAAATCT GCTTCAGGTG
1051 GCTTAAAGG AGAAAATCTC AAAGAAACGA TTCTTGTTG TTCTTGATGA TGTATGGAGC GAGAGCTATA
1121 CTGATTGGGA AATCTAGAA CGTCCATTTC TAGCAGGAGC ACCAGGAAGT AAAGTAATCA TCACAACCCG
1191 CAAGTTGTCG TTGCTAAACC AATTGGGTCA TGATCAACCA TACCAATTGT CTGATTGTTC ACATGACAAT
1261 GCTCTATCCT TATTTTGTC ACACGCATTT GGTGTAATA GCCTTGATTG ACATCCGATA CTTAAACCCAC
1331 ATGGTGAAGG TATTGTTGAA AAATGTGATG GTTTGCCATT GGCTTTGATT GCACCTGGGA GGTATTTGAG
1401 GACAAAAGA GATGAGGAAG AATGGAAGGA ACTATTGAAT AGTGAGATAT GGAGGTTAGG AAAGAGAGAT
1471 GAGATATTC CGGTCTTAG ACTAAGCTAT AATGATCTTT CTGCTCTTTT GAAGCAGTTG TTTGCAATTT
1541 GCTCCTTGTT CCCCAAGAC TATGTGTTCA ACAAGGAGAA GTTGATTTTA TTATGGATGG CAGAAGGGTT
1611 TTTGCACAAT GAAAATACAA ACAAGTCAAT GGAACGCTTA GNTCTTGAAT ATTTTGAAGA CTTGTTGTCA
1681 AGGTCTTTT TTCAACATGC ACTCGATGAC AAATCGTTGT TTGTTGGTGA CGACCTCATG AATGACTTGG
1751 CCACATCTGT TGCTGGAGAT TATTTTTTAA GATTAGACAT TGAAATGAAA AAGGAAGCTT TGGAAAAATA
1821 CCGACATATG TCATTTGTTT GTGAGAGTTA CATGGTTTAC AAAAGGTTTC AACCATTTAAG AGGAGCTAAA
1891 AAATTGAGAA CTTTCTTAGC AATGCCGTTT GGGATGATAA AAAGTTGGAC AACATTTTAC TTATCAAATA
1961 AGGTCTTGA TGACTTACTT CACGAATTAC CATTGTTGAG AGTTCTAAGT TTGAGTTATC TTAGCATCAA
2031 GGAGGTACCT GAAATAATAG GCAATTTGAA ACTTTCGGG TATCTTAATT TATCACACAC GAGTATCACA
2101 CATTTTACCAG AAAATGCTG CAATCTTTAC AACTTACAAA CATTGATCCT TTGTGGCTGT TGTMTTATAA
2171 CCAAGTTTCC CACAACCTT TTAAGCTTA GAAATTTACG GCATTTGGAC ATTAGCGATA CTCCCGGTTT
2241 GAAGAAAGAT TCCTCGGGGA TTGGTGAATT GAAGAACCTA CACACYCTCT CCAAGCTCAT TATTTGAGGT
2311 GAAAAATAGAC TAAACGAGCT TAAGAACTTA CAAAATCTCC ATG
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RLG1b - Diana
[Strand]

```
1  TACTACTACT AGAATTCGGT GTTGTAAGA CGACTCTAGC TAGACTTTTG TATGAGGAAA TGCAAGGGAA
71  GGATCACTTC GAACTTAAGG CGTGGGTATG TGTTCCTGAT GAGTTTGATA TCTTCAATAT AAGCAAAATT
141 ATCTTACAAT CGATAGGTGG TGGAAACCAA GAATTTACGG ACTTAAACCT GCTTCGAGTA GCTTTAAAG
211 AGAAGATCTC AAAGAAAAGa TTTCTCTCTG TTCTTGATGA TGTTTGGAGT GAAAGCTATA CCGATTGGGA
281 AATTNTAGAA CGCCCATTTT TTGCAGGGGC ACCTGGAAGT AAGATTATTA TCACCACCCG GAAGCTGTCA
351 TTGTTAAACA AACTCGGTTA CAATCAACCT TACAACCTTT CGGTTTGTG ACATGAGAAT GCTTTGTCTT
421 TATTCTGTCA GCATGCATTG GGTGAAGATA ACTTCAATTC ACATCCAACA CTTAAACCAC ATGGCGGAGG
491 TATTGTTGAA AAATGTGATG GATTGCCATT GGCATTGTG ACATGATGAT GATG
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SEQ ID 137

SEQ ID NO: 3

R1G1C

[Strand]

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1   TCCCGTGCAA CGTNTATCAT TCAGAAGNGC CCAAAGACCA NAGATNTGTT TAANGNTGNT TMTCAGAAGG
71  AAGTAATTGA TGAAGCTGTN AAAAGATGGC TGATTGATNT CCAACAATTG GCTTAGGACA CTGANGACNA
141 ACTTGATGAT NTCGCCACAG AAGCTATTCA TCGTGAGTTG ATCCGTGAAA CTGGAGCTTC CNGCAGCATG
211 GTAGAAAGC TAATCCCAAG TTGTTGCACA AGTTTCTCAC AAAGTAATAG GATGCATGCC AGGTTAGATG
281 ATATTGCCGC TAAGTACAA GAACTGGTAG AGGCGAAAAA TAATCTTGGT TTAAGTGTGA TAACATACGA
351 AAAACCCAAA ATTGAAAGAG ATGAGGCGTN TTTGGTAGAT GCAAGTGGTA TCATTGGACG TGAAGATGAT
421 AAGAAAAAAT TGCTTCAGAA GCTGTTGGGG GATACTTATG AATCAAGTAG TCAAACTTC AACATCGTGC
491 CCATAGTTGG TATGGGTGGG GTAGGTAAAA CAACTCTAGC TAGACTTTTG TATGATGAAA AAAAAGTGAA
561 GGATCACTTC GAACTCAGGG TTTGGGTTTG TGTTCCTGAT GAGTTCAGTG TTCCAATAT AAGCAGAGTT
631 ATCTATCAAT CTGTGACTGG TGRAAACAAA GAATTTGCAG ATTTAAATCT GCTTCAAGAA GCCCTTAAAG
701 AGAAACTTCA GAACAACTA TTTCTAATAG TTTTAGATGA TGTATGGTCT GAAAGCTATG GTGATTGGGA
771 GAAATTAGTG GGCCCATTTT ATGCTGGGAC TTCTGGAAGT AGAATAATCA TGACTACTCG GAAGGAGCAA
841 TTACTCAAAC AGCTGGGTTT TTCTCATGAA GACCCCTGCG ATAGTATAGA CTCCCTGCAA CGTCTATCAC
911 AAGAAGATGC TTTGCTTTG TTTCTCAAC ACGCATTTGG TGTACCTAAC TTTGATTAC ATCCAACACT
981 AAGGCCATAT GGGGAACAGT TTGTGAAAAA ATGTGGGGGA TTGCCTTTGG CCTTGT
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SEQ ID NO:4

RLG1D

[Strand]

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1  CNTACCCATTC TACGAGATCG CTGTCCCTCC TCGATCTGCT TAACGATGCT TCCCAGAAGG AAGTNACTAA
71  TGAAGCCGTT AAAAGATGGC TGAATGATCT CCAACATTTG GCTTATGACA TANACGACCT ACTTGATGAT
141 CTTGCAACAS AAAGCTATT CTCSTGAGTT GACCGANGAA GGTGGAGCCT CCACCAGTAT GGTAAAGAAA
211 CTAATCCCAA GTTGTTCAC AAGTTTCTCA CAAAGTTATA GGATGCATGC CAAGTTAGAT GATATTGCCA
281 CCAGGTTACA AGAACTGGTA GAGGCAAAAA ATAATCTTGG TTTAAGTGTG ATAACATATG AAAAGCCCCA
351 AATTGAAAGG TATGAGGCAT CTTTGGTAGA CGAAAGTGGT ATTTTGGAC GTTNAGATGA TNAGAAAAAA
421 TTGATGGAGA AGCTGTTGGA GGATAAAGAT GAATCCGGAG TCNAACTTC AGCATCCTGC CCATAATTGG
491 TATGGGTGGA GTTGGCNAAA CAACTCTAGC TAGACTCTTG TTTGATGAAA AGACAGTGAA GGATCACTTC
561 GAACTCAGGG CTTGGGTTTG TGTTCCTGAT GAATTCAGTA TTCTCAACAT AAGCAAAGTT ATCTATCAAT
631 CTGTGACCGG GGAAAGAGAA GAGTTTGAAG ACTTAAATCT GCTTCAAGAA GCTCTTAGAG GGAAACTACA
701 AAACAAACTA TTTCTAATAG TTTTGGATGA TGTATGGTCG GAAAGCTATG GTGATTGGGA GAAATTAGTG
771 GGCCCAATTC ATGCTGGGAC TTCTGGAAGT AGAATAATCA TGACTIONCG GAAGGAGCAA TTACTIONAAC
841 AGTTGGGTTT TTCTCATCAA GACCCCTCTC GTTGATAGA CTCCCTGCAA CGTCTATCAC AAGATGATGC
911 TTTGTCTTTG TTTGCTCAAC ACGCAATTGG TGMCCA
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RLGLE
[Strand]

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1  TCTAGCTAGA CTTTGTGTATG ACGAGATGCA AGAGAAGGAT CACTTCGAAC TCAAGGCGTG GGTTTGTGTT
71  TCTGATGAGT TTGATATATT CAATATAAGC AAAATTATTT TCCAATCGAT AGGAGGTGGA AACCAAGAAT
141 TTAAGGACTT AAATCTCCTT CAAGTAGCTG TAAAAGAGAA GATTTCAAAG AAACGATTTC TACTTGTTC
211 TGATGATGTT TGGAGTGAAA GCTATGCCGA TTGGGAAATT CTGGAACGCC CATTTCCTGC AGGGGCAGCC
281 GGAAGTAAAA TTATCATGAC GACCCGGAAG CAGTCATTGC TAACCAAAC TCGTTACAAG CAACCTTACA
351 ACCTTTCCGT TTTGTACAT GACAGTGCTC TCTCTTTATT CTGTCAGCAT GCATTGGGTG AAGATAACTT
421 CGATTCCAT CCAACACTTA AACCACATGG CGAAGGCATT GTTGAAAAAT GTGCT
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SEQ ID NO:5

RLG1F
[Strand]

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1   ATTTTCNGCT  CBAACAAAN  AAAAGCAATG  GCTGAAATCT  TTCTTTGNGC  ATTCTAGACC  AGTATTCTTT
71  GAAAAGNIGG  CTTCTGAAGC  CTTGAAGAAG  ATCGCTCGCT  TCCATCGGAT  TGATTCTGAG  CTCAAGAAAC
141 TGAAGAGGTC  ATTAATCCAG  ATCAGATCTG  TGCTTAATGA  TGCTTCTGAG  AAGGAAATAA  GTGATGAAGC
211 TGTAAAGAA  TGGCTGAATG  GTCTCCAACA  TTTGTCTTAC  GACATAGACG  ACCTACTTGA  TGATTTGGCA
281 ACCGAAACTA  TGCATCGTGA  GTTGACCCAC  GGATCTGGAG  CCTCCACCAG  CTTGTAAGAA  AGATAATCCC
351 AACTTGTTC  ACAGATTTCT  CACTAAGTAG  TAAGATCGGT  AACCAAGTTAG  ATAATATTAC  CATCAAGTTA
421 CAAGAACG  TAGAGGAAAA  AGATAATCTT  GGCTTAAGTG  TGAAGGTTGA  AAGCCCAAAA  CATACCAACA
491 GAAGATTACA  GACCTCTTTG  GTAGATGCAT  CTAGCATTAT  TGGTCTGTGA  GGTGATAAGG  ATGCATTGCT
561 CCATAAGCTG  CTGGAGGATG  AACCAGTGA  TAGAAACTTT  AGCATCGTGC  CAATAGTTGG  TATGGGTGGT
631 GTGGGTAAGA  CGACTCTAGC  TAGACTTTTG  TATGACGAGA  TGCAAGAGAA  GGATCACTTC  GAAGTCAAGG
701 CGTGGGTTTG  TGTTCCTGAT  GAGTTTGATA  TCTTCAATAT  AAGCAAAGTT  ATCTTCCAAT  CGATAGGTGG
771 TGGARACCA  GAATTTAAGG  ACTTAAATCT  CCTTCAAGTA  GCTGTAAAAG  AGAAGATTTT  AAAGAAACGA
841 TTTCTTNTTG  TTCTGGATGA  TGTTTGGAGT  GAAAGCTATA  CAGAATGGGA  AATTCTAGCA  CGTCCATTTC
911 TTGCAAGGGC  ACCAGGAAGT  AAGATATCA  TGACGACCCG  GAAGTTGTCG  TTGCTAACCA  AACTCGGTTA
981 CAATCAACCT  TACACCTTT  CSGTTTGTG  ACATGATAAT  GCTTGTCTTT  TATTCTGTCA  GCAYGCATTG
1051 GGTGAAGATA  ACTTCGATTC  ACATCCAACA  CTTAAACCAC  ASGGTGAAAG  TATTGTTGAA  AAATGTGACG
1121 GTTTACCATT  GGCTTTRATT  GCACTTGGGA  GRTTGTGAR  GACAAAAACA  GATGAGGAAG  AATGGAARGA
1191 AGTGTGAAT  AGTGAAATAT  GGGGGTCAG  AAAGGGAGAT  GAGATTGTTC  CGGCTCTTAA  ACTAAGCTAC
1261 AATGATCTCT  CTGCTCTTT  GAAGAAGTTG  TTTGCATACT  GCTCCTTGT  CCCAAAAGAC  TATGTGTTCG
1331 ATAAGGAGGA  GTTGATTTTG  TGTGTGATGG  CAGAAGGGTT  TTTGCACCAA  TCAACCACAA  GCAAGTCBAT
1401 GGAACGCTTG  GGHGATGAAG  GTTTTGATGA  ATTGTTGTCA  AGATCATTTT  TTCAACATGC  CCCTGATGCC
1471 AAATCGATGT  TTGTGATGCA  TGACCTGATG  AATGACTTGG  CHACATCTGT  TGCTGGAGAT  TTTTMTTCAA
1541 GGATGGACAT  TGAGATGAAG  AARGAATTTA  GGAAGGAAGC  TTTGSAAAAG  YAYCGCCATA  TGTCAMTTGT
1611 TTGTGAKGAT  TACATGGTK  ACAAAGGTT  CRAGCCATTS  ACAAGGAGCT  AG
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SEQ ID NO: 6

RLGIG
[Strand]

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1  GTGAAGGATC ACTTCGAACT CAGGGCTTGG GTTGTGTGTT CTGATGAATT TAATATCCTC AATATAAGCA
71  AAGTAATTTA TCAATCTGTA ACCGGGGAAA AAAAGGAGTT TGAAGACTTA AATCTGCTTC AAGAAGCTCT
141 TAAAGAAAAA CTTTGGAATC AGTTATTTCT AATAGTTCTG GATGATGTGT GGTCTGAAAG CTATCGTGAT
211 TGGGAGAAAT TAGTGGGCCC ATTTTITTCG GGGTCTCCTG GAAGTATGAT TATCATGACA ACTCGGAAGG
281 AGCAATTGCC AAGAAAGCTG GGTTTTCCTC ATCAAGACCC TTTGCAGGT CTATCACATG ACGATGCTTT
351 GTCTTGTTT GCTCAACAG CATTGTGTGT ACCA
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SEQ ID NO: 7

RLGH
[Strand]

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1  TCTAGCTAGA CTTTGTATG AGGAAATGCA AGGGAAGGAT CACTTCGAAC TCAAGGCGTG GGTATGTGTT
71  TCTGATGAGT TTGATATCTT CAATATAAGC AAAATTATCT TACAATCGAT AGGTGGTGGA AACCAAGAAT
141 TTACGGACTT AAACCTGCTT CAAGTAGCTT TAAAAGAGAA GATCTCAAAG AAAAGATTTC TTCTTGTTC
211 TGATGATGTT TGGAGTGAAA GCTATACCGA TTGGGAAATT CTAGAAGGCC CATTTCCTGC AGGGGCACCT
281 GGAAGTAAGA TTATTATCAC CACCCGGAAG CTGTCATTGT TAAACAACT CGGTTACAAT CAACCTTACA
351 ACCTTTCGGT TTGTGCATAT GAGAATGCTT TGTCTTTATT CTGTCAGCAT GCATTGGGTG AAGATAACTT
421 CAATTCACAT CCAACACTTA AACCACATGG CGAAGGTATT GTTGAAAAAT GTGAT
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SEQ ID NO: 8

RLGI
[Strand]

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1  TCTAGCTAGA CTTGTGTATG ATGAGATGCA AGAGAAGGAT CACTTTGAAC TCAAGGCGTG GGTATGTGTT
71  TCTGATGAGT TTGATATATT CAATATAAGC AAAATTATTT TCCAATCGAT AGGAGGTGGA AACCAAGAAT
141 TTAAGGACTT AAACCTCCTT CAAGTAGCTG TAAAAGAGAA GATTTTAAAG AAACGATTTC TTCTTGTTC
211 TGACGACGTT TGGAGTGAAA GCTATGCCGA TTGGGAAATT NTGGAACGCC CATTTCTTGC AGGGGCAGCC
281 GGAAGTAAAA TTATCATGAC AACCCGAAAG CAGTCATTGC TAACCAAACT CGGTTACAAG CAACCTTACA
351 ACCCTTCCGT TTGTACACAT GACAGTGCTC TGTCTTTATT CTGTCAGCAT GCATTGGGTG AAGGTAACCT
421 CGATTACAT CCAACACTTA AACCACATGG CGAAGGCATT GTTGAAAAAT GTGCTGGATT GCCATTGGCA
491 TTGTCGACA
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SEQ ID NO. 9

RLGLJ

[Strand]

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1   TACTACTACT AGAATTCGGT GTTGGTAAGA CGACTCTAGC TAGACTTTTG TATGAGGAAA TGCAAGGGAA
71  GGATCACTTC GAACTTAAGG CGTGGGTATG TGTTTCTGAT GAGTTTGATA TCTTCAATAT AAGCAAAATT
141 ATCTTACCAT CGATAGGTGG TGGAAACCAA GAATTTACGG ACTTAAACCT GCTTCGAGTA GCTTTAAAAG
211 AGAAGATGTC AAAGAAAAGa TTCTTCTTG TTCTTGATGA TGTTTGGAGT GAAAGCTATA CCGATTGGGA
281 AATTINTAGAA CGCCCATTTT TTGCAGGGGC ACCTGGAAGT AAGATTATTA TCACCACCCG GAAGCTGTCA
351 TTGTTAAACA AACTCGGTTA CAATCAACCT TACAACCTTT CGGTTTGTG ACATGAGAAT GCTTTGTCTT
421 TATTCTGTCA GCATGCATTG GGTGAAGATA ACTTCAATTC ACATCCAACA CTAAACCAC ATGGCGGAGG
491 TATTGTGAA AAATGTGATG GATTGCCATT GGCATTGTG ACATGATGAT GATG
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SEQ ID NO:10

RLGIA a.a.

IVTVRTR?LSLLHLLSYVIFS?I?PH?ILWLF.INFYSTCHFMSFSILLSFT.YLNVITINAYLFFFK.THIYR
LKSynt.VKLI.YICSSPVYLYVSSLIYLLFIY.SR.SL.Y.KFNLFKI.NY..SHNLNKIKKNGPTISPSLFQLINIV
SILLRFHPNQYFQRMDSYGVSEFAFRHCSLKEIINQMELLQCSLLMKGELYVK?MSAI?LHPEPTTSLV
YHQTQNGGSR?T?KS.RIDYFCPHGLTEERV.FIIFL?KNYRSIEFLHRQRSFTSQCFVKQFLIFLSFR.NS
SIATCNLQ.LLGPQICGGR.FNPHIHCKQ.FKSISVHPIHQHLLIIEIIHASSISSTSILYSLLLSY.TMAEIVLS
AFLTUVFEKLA?EALKKIVRSKRIESELKKLKETLDQIQDILLNDASQKEVTNEAVKRWLNDLQHLAYDID
DLLDD?ATEAV?RELTEEGGASSSMVRKLIPSCCTSFSQSNRMHAKLDDIATRLQELVEAKNNLGLSVI
TYEKPKEIERYEASLVDESGTVGREDDKKKLEKLLGDKDESGSQNFSIVPIVGMGGVGKTTLARLLYDEK
KVKDHFELRAWVCVSEFSVPNISRVYQSVTGEKKEFEDNLILQEALKEKLRNQLFLIVLDDVWSESY
GDWEKLVGPFLAGSFGRIMTTRKEQLLRKLGFSHQDPLEGLSQDDALSIFAQHAFGVPNFDSHPTLR
PHGELFVKKCDGLPLALRTLGRLLRTKTDEEQWKELLDSEIWRGKSDIWPALRLSYNDLSA?LKLFA
YCSLFPKDYEFDKKEELLLWMAEGFLHQPT?NKSQRGLLEYF?ELLSRSFFQHAPN?KSLFVMHDLMDND
LATFVAGEFFSRLDIEMKKEFRM?SLEKHRHMSFVCE?YIGYK?FEPFRGAKNLRTFLALSVGVVEDWK
MFYLSNKVLND?LQDLPLLRLV?LI?L?I?VP??VGSM?HLRYLNLS?T?ITHLPE??CNLYNLQTLIV
SGC?YLV?LPKTF?LKNL?HFDMR?TP?LKNMPL?IGELK?LQTLF?NIGIAITELKNL?NLHGK?CIGG
LGKMENAVGCTLSELVSKKV?..?NW??G...I.CFPKWEHLKKKSSMK.CLIMVL?KKP?IMSIGGIEFPN
WVGLRVSETRDVFMVYEK?CFT.FHQSPSGK.MIFSG?TDEMWRGMIG?LGAVEEISIHSCNEIRYLWE
SEAEASKVLMNLKKLDLGECENLVSLGEKKEDNHNINSGSSLTSFRRLNWWRCNSLEHCRCPDSMENLY
MHMCDS?TSVSFPTGGGQKIKSLTITDCKKLSEELGGRETRVLYNSKMQMLESVDIRNWPNLKSISEL
SCFIHLNRLYISNCPS?ESFPDHELPNLTSLTDRRRGQRFSEYERLRFDWPSF

SEQ ID NO:11

RLGIB a.a.

NRSYENRCPLLPVI...EKI.LKV.IQNPLFHR.YDALAVVCKNSIINANQNSS.ETI.IMV.IVLSPTYTHFFQIPII
HTYKCSHIRFSLAMAEILGSAFFAVFFFEKLASEALKRVACSKVIDKELEKLNSS.INIKALLNDASQKEIS
KEAVKEWLNALQHLPYDIDLLGDLATKAIHRKFSEEGATINKVRKLIPSCFSSLSSTKMRNKIHNITS
KLQELLEERNLGLCEIGESRKLNRKSETS?LDPSSIVGRTDDKEALLKLYEPCDRNFSILPIVGMGGL
DKTTLGRLLYD?MQVKDHFELKAWVCVSDEFDIFGISKTFESIEGGNQEFKDLNLLQVALKEKISKKRFL
VLDDVWSESYTDWEILERPFLAGAPGSKVIITTRKLSLLNQLGHDQPYQLSDLSHDNALSIFCQHAFG
VNSFDSHPILKPHGEGIVEKCDGLPLALIALGRLLRTKRDEEEWKELLNSEIWRLGKRDEIIP?LRLSYND
LSASLKQLFAYCSLFPKDYVFNKEKLILLWMAEGFLHNENTNKSMEERL?LEYFDDLLSRFFQHALDDKS
LFVVHDLMDLATS VAGDYFLRLDIEMKKEALEKYRHMSFVCESYMYKRFEPFKGAKKLRTFLAMPV
GMIKSWTTFYLSNKVLDLLHELPLLRVLSLSYLSIKEVPEIIGNLKHLRYLNLSHTSITHLPENVCNLYN
LQTLILCGCCFITKFPNNFLKLRNLRHLDISDTPGLKKMSSGIGELKNLHTLSKLIIGGENRLNELKNLQNL
H

SEQ ID NO:12

RLG 1 c.a.a.

SRAT?IIQK?PKT?D?F????QKEVIDEAVKRWLID?QQLAYDT?D?LDD?ATEAIHRELIRETGAS?S
MVRKLIPSCCTSFSQSNRMHARLDDIAAK?QELVEAKNNLGLSVITYEKP KIERDEA?LVDASGIIIGRED
DKKKLLQKLLGDTYESSQNFNIVPIVGMGGVGKTTLARLLYDEKKVKDHFELRVWVCVSDEFSVPNIS
RVYQSVTGENKEFADLNLLQEALKEKLQNKLFLLVDDVWSESYGDWEKLVGPFHAGTSGSRIIMTTR
KEQLLKQLGFSHEDPLHSIDSLQRLSQEDALSLFSQHAFGVNFD SHPTLRPYGEQFVKKCGGLPLAL

SEQ ID NO:13

RLG ID

?T?LRDRCPSSICLTMLPRRK?LMKPLKDG.MISNIWLMT?TTYLMILQ?KAI??ELT?EGGASTSMVRK
LIPSCCTSFSSQSYRMHAKLDDIATRLQELVEAKNNLGLSVITYEKPPIERYEASLVDESGIFGR?DD?KK
LMEKLLEDKDESGVKLQHLP!IGMGGVG?TTLARLLFDEKTVKDHFEFRAWVCVSDEFSLNISKVIYQS
VTGEKKEFEDLNLLQEALRGKLNKLFVLDDVWSESYGDWEKLVGPFHAGTSGSRIIMTTRKEQLLK
QLGFSHQDPLRCIDSLQRLSQDDALSFAQHAFG?

SEQ ID NO: 14

RLGIE

LARLLYDEMQEKDHFELKAWVCVSDEFDIFNISKIIFQSIGGGNQEFKDLNLLQVAVKEKISKKRFLLVLD
DWSESYADWEILERPFLAGAAGSKIIMTTRKQSLTLKGYKQPYNLSVLSHDSALSFLCQHALGEDNF
DSHPTLKPHGEGIVEKCA

SEQ ID NO: 15

RLGIF

FSA?NK?KQWLKSFF?HSRPVFFEK?ASEALKKIARFHRIDSELKKLKRSIJQIRSVLNDASEKEISDEA
VKEWLNGLOHLSYDIDDLDDLATETMHRELTDTLEPPPACKKDNPTCCTDFSLSSKMRNKLDNITIKL
QELVEEKDNLGLSVKGESPKHTNRRLQTSYDASSIIGREGDKDALLHKLEDEPSDRNFSIVPIVGMGG
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LGEDNFD SHPTLKP?GESIVEKODGLPLALIALGRLL?TKTDEEEWKEVLNSEIWGSGKGD EIVPALKLS
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SEQ ID NO: 16

RLG1 G

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SEQ ID NO: 17

RLG 1 H

LARLLYEEMQGKDHFKAWVCVSDEFDIFNISKIILQSIGGGNQEFTDLNLLQVALKEKISKKRFLLVLD
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SHPTLKPHGEGIVEKCD

SEQ ID NO: 18

RLGI I

LARLVYDEMQEKDHFELKAWVCVSDEFDIFNISKIIFQSIGGGNQEFKDLNLLQVAVKEKILKKRFLLVLD
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SEQ ID NO: 19

RLG 15

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SEQ ID NO: 20

SEQ ID NO: 21
RLG 2A

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911 GGATAAATGT  GGTAAATTTA NAACAAGTGT TTNACNVTG AAGGGTWTGG AAAGGTTGAA AAAAGTTAA
981 ATGATAAAAT  GTTTACACAA ATGTTGTATC CGACTGAATA TNAITTTAA GGATNATTGT ATTAAATGT
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1121 ATTTTGTATA  TANATTAGGA ATGAAAATGA GCAACCCATA CATACTTATC TTTGGTAGIT TGGTTATTAT
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2241 ACAGACAAG  TGCAGACATA TGCCCTCGCA GAGTGTGTGT TTTTGTAAAG TGCGCAGACC AAAAAACGT
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2381 AAAAAAATA  GACCACCGGT TTTTTTTTTT TTTTNTTCT TTCTCTGTGA GCTGAAAATG CATTTTTAAT
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3361 AAATCCATGA  TTTACCTTTA AATATTAACA AAAATGACCA TAAATRAATA AAAAATTAGG ATACCAAACC
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3641 TTGAAACTAG  ATTTGGACTT TTGCCCTTGA TGATTCATAA GATATTGCAT GGAGTTGAGA TTGTGTAAGA
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3921 CTTCTTCTGA  CCAAACATAA TTATAACAAA ATTTGAATATC ATTTCTGTGA ATCAATTTTA ACTTTTGTTA
3991 TTATCAATCAT  GTCTAAAATT GCCACAAGTT TATTTTCATA GTCATATTGG ATTATGAAAG GACTATTTTT
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SEQ ID NO: 2/
RLG 2A cont.

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4271 GAAACACATA GGGTACCTCA TTTCCTGCAG GCAATATATG AGGGAATGG GTATCAAAAT GAGGGGATTG
4341 AATGCTACAA GACTTGGTGT CGAAGAGCAC GTGAACCGGA ACATAAGCAA CCAGCTTGAG GTTCCAGCCC
4411 AAGTCAAGGG TTGGTTTGAA GAAGTAGGAA AGATCAATGC AAAAGTGGAA AATTTCCCTA GCGATGTGTG
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4831 AGCGCGTGT AGGGGAAAAA ACAGACCCCA TTGCTATTCA ATCAGCTGTA GCAGATTACC TAGGTATAGA
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RLG2A cont.

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SEQ ID NO: 21

RLGA a.a.

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SEQ ID NO:22

RLG 23

SEQ ID NO: 23

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141 TTTAATATATC ATAATTIGAA AATCATCAAA TTGTATTCCA TGTATATTTA TGTATCAGA TAATTAATAA
211 TATGTGGGCC ACACAAATCC ACATCATCAG ACACCCACC TTATTGTCGG CTACCTCACC ACTTGATGA
281 TCCCGACATC TTCCCAACCC CACCGACGAC TTGGGGTCTC CTTAATATAT CAATTAATTT CTGTAAGTAT
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421 AATTCAAATCG CGTTTCATTC TTGCATTAAA AAAAAAGACT GTACTGTGT CAATATTTTA CTTATAACCT
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561 GGGTCACCGG GAATCAAAGC ACTTATGTAA AAGCAGGGGA AATACAAAAA ATTTACTCGA AACAAATTTT
631 ATTCAATTTA AGTGAGATAA TAAATGTTCTG ATTAGATTAT GAGAACTAGG AGATTAAAGT GATATATCCC
701 ATTTAAAGA AATTGCATTA TTAATTTTGG ATCTCTTGAT GATGACAAAA TTAACCTCGT ACAGGTTATA
771 TATCATATAC AAAATGAGTG GCTATGCTTT CGCTTTCCAA AAAGCAATTA TAGTTATACT ACACCTACAA
841 ATTTTAAAG GGGTTAAACA TATCAAAATA CTTGATAAGT AATTATATAA ATATGCATTT AACCTCTAA
911 AGAAAAATGCT ACTAAGCTTG GACCATCTCA GAATTACAAT CATACCCCTC CCCTCAAAAA AGATTGCGTAT
981 ATATCATGTC ATTTGGCATT CATTTCTTTT TCACAATTCA TAGTTCTATT CTCAAAAAAT TCGAGTTCTC
1051 GTATTTGTAA GGAAGATCAG AAGAGACTGT TCACACAGGT ACTCTCTTTT ATTTATTGAT TCACATTCAT
1121 ATATGTTTAT GTTTTCTTGC TTAATGGTIT CGTCAGTCTA ACTGCGCTTG CTGATTTAAA TTTCTTCACT
1191 TTCTTCCACG GATTTTTTAA ATATTAGTTT TGTGAATGAA CAATTTGGTGA AGGAAAGAAA CATGGGAGTC
1261 TTTTCTAAG TAAACCTAGA TACTTAGGTT ATAAGGGTAT ATGCTAAAAT GAACTATGCC CATTCACCTT
1331 TGCCCTTTCT TTTACTTTTT AGTTTTTAGA ATCCAAGTTT TCATATGTAT CTCGATGTGT GAGAAGAATA
1401 GGCATTAGAA AGGTAAAGGA CGTACATAAA ATTGAATTAAT TAGTGAATGT TCTTTGATAT CATTAATTTT
1471 ACTCTCATAA AAAGCATATA GATCAAAAC AAATTGCTAC TTGTTAGTGT AACAACTTCG ACTTAATAAT
1541 GTTAATAATC AAGATTCTCT TGATTTCAC TATTTTCTAA CCGAACAAAG TCACATAAAA CTCATATTCG
1611 TTTGAGTCTG AGTGGTTTAT ATTTGGGGTT TACATTTAA TTTTGTGTC ATGAATGTGA AAATAGACTG
1681 CTTATTGATT CTTTGTGTTT CATTGAGTTG ATTTTCATTA TTACTACCTT ACAAATTTGCT CAGTGATAGA
1751 TTTCCATTAA TTGTCTAATT CGGTTCGCTC TAAATATGTA GGAGCTACTA AAAGCAAAAA TATCGAGCAA
1821 TGTCGGACCC AACGGGGATT GCTGGTGCCA TTATTAACCC AATTGCTCAG ACGGCTTGG TTCCCGTTAC
1891 GGCCCATGTA GGCTACATGA TTCTCTGAG AAAATATGTG AGGGTCATGC AGATGAAAAAT GACAGAGTTG
1961 AATACCTCAA GAATCAGTGT AGAGGAACAC ATTAGCCCGA ACACAAGAAA TCATCTTCAG TTCCATCTCA
2031 AACTAAGGAA TGGTTGGACC AAGTAGAAGG GATCAGAGCA AATGTGGAAA ACTTTCGGAT TGATGTCATC
2101 ACTTGTGTGA GTCTCAGGAT CAGGCACAAG CTTCGACAGA AAGCNTTCAA GATAACTGAG CAGATTGAAA
2171 GTCTAATCGA ACAACTCTCC CTGATCAGTT GGACTGATGA TCCAGTTCYT CTAGGAAGAG TTGGTTCCAT
2241 GAATGCATCC ACCTCTGCAT CATTAAGTGA TGATTTCCCA TCAAGAGAGA AAACCTTTTAC ACAAGCACTA
2311 ATAGCTCTCG AACCCAACCA AAAATTCAC ATGGTAGCCT TGTGTGGGAT GGGTGGAGT GGGGAAGACTA
2381 GAATGATGCA AAGGCTGAAG AAGGCTGTTG AAGAAAAGAA ATTTGTTAAT TATATTGTTG GGGCAGTTAT
2451 AKGGGAAAAG ACGGACCCCT TTGCCATTCA AGAAGCTATA GCAGATTACC TCGGTATACA ACTCAATGAA
2521 AAAAGCTAAGC CAGCAAGAGC TGATAAGCTT CGTGAATGGT TCAAAAAGAA TTCAGATGGA GGTAAAGACTA
2591 AGTTCCCTCAT AGTACTTGAC GATGTTTGGC AATTAGTTGA TCTTGAAGAT ATTGGGTTAA GTCCCTTTCC
2661 AAATCAAGGT GTCGACTTCA AGGTCTTGTT GACATCACGA GACTCACAAG TTTCGACTAT GATGGGGTT
2731 GAAGCTAATT CAATTATTAA CGTGGGCCTT CTAACCTGAAG CAGAAGCTCA AAGTCTGTTT CAACAATTTG
2801 TAGAACTTTC TGAGCCCGAG CTCCAGAAGA TAGGAGAGGA TATCGTAAGG AAGTGTTCGG GTCTACCTAT
2871 TGCCCAAAAA ACCATGGCAT GTWCTCTTAG AAATAAAGA AAGGATGCAT GGAAGGATGC ACTTTCGGCG
2941 ATAGAGCACT ATGACATTCA CAATGTTGCG CCCAAAGTCT TTGAAACGAG CTACCACAAT CTCCAAGAAG
3011 AGGAGACTAA ATCCACTTTT TTAATGTGTG GTTTGTTTCC CGAAGACTTC GATATTCCTA CTGAGGAGTT
3081 GATGAGGTAT GGATGGGGCT TGAAGCTATT TGATAGAGTT TATACGATTA GAGAAGCAAG AACAGGGCTC
3151 AACACCTGCA TTGAGCGACT GGTGCAGACA AATTGTGTTA TTGAAAGTGA TGATGTTGGG TGTGTCAAGA
3221 TGCATGATCT GGTCCGTGCT TTGTTTITGG GTATGTTTTC TGAAGTCGAG CATGCTTCTA TTGTCAACCA
3291 TGGTAATATG CCTGGGTGGC CTGATGAAAA TGATATGATC GTGCACCTTT GCAAAAAGAA TTCAATTAACA
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3431 GAGATAGTC GCTAAGGTTT CCTCAAGACT TTTATGAAGG AATGGAAAAAG CTCCATGTTA TATCATACCA
3501 TAAAAAGAG TACCAATGTC TTCTTTTGGC ACCTCGATGC TCCACCAACA TTGGGGTGCT TCATCTCACT
3571 GAATGTTTAT TAAAGATGTT TGATTGCTCT TCTATCGGAA ATCTATCGAA TCTGGAAGTG CTGAGCTTTG
3641 CAAATTTCTCA CATTTAATGG TTACCTTCCA CAGTCAGAAA TTTAAAGAG CTAAGGTTAC TTGATCTGAG
3711 ATTTTGTGAT GGTCTCCGTA TAGAACAGGG TGTCTTGAAA AGTTTTGTCA AACTTGAAGA ATTTTATATT
3781 GGAGATAGTC CTGGGTTTAT AGATGATAAC TGCAATGAGA TGGCAGAGCG TTCTTACAA CTTTCTGCTG
3851 TAGAAATTCG GTTCTTTAAT AACAAAGGCTG AAGTGAAAAA TATGTCAATT GAGAATCTTT AACGATTCAA
3921 GATCTCAGTG GGATGCTCTT TTGATGAAAA TATCAATATG AGTAGCCACT CATACGAAAA CATGTTGCAA
3991 TTGGTGACCA ACAAAGGTGA TGTATTAGAC TCTAAACTTA ATGGGTTATT TTTGAAAAA GAGGTGCTTT
4061 TTTTAAAGTGT GCATGGCATG AATGATCTTG AAGATGTTGA GGTGAAGTCG ACACATCTCA CTCAGTCTTC

RLG 2B cont.

SEQ ID NO: 23

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4201 CTTGCAAAACA CTTTGTCAAG ACTTGAGCAT CTAGAAGTTT GTGAATGTGA GAATATGGAA GAACTCATAC
4271 ATACTGGAAT TGGGGGTGTG GGAGAAGAGA CAATTACTTT CCCTAAGCTG AAGTTTTTAT CTTTGAGTCA
4341 ACTACCGAAG TTATCAAGTT TGTGCCATAA TGTCACATA ATTGGGCTAC CACATCTCGT AGACTTGATA
4411 CTTAAGGGCA TTCCAGGTTT CACAGTCATT TATCCGCAGA ACAAGTTGCG AACATCTAGT TTGTTGAAGG
4481 AAGGGGTAGA TATATGTTCT TTATGTTAAT ACAATTTAAA TAATATTTTC AACCATAATT TCATAATATA
4551 TCTGTAATTT GATTGTATGA TGTGTTATTG TTTATATGTG GCTATTAAGG GATGATTATT TTCAGGGTTG
4621 TGATTCCTAA GTTGGAGACA CTTCAAATTG ATGACATGGA GAACTTAGAA GAAATATGGC CTTGTGAATC
4691 TAGTGGAGGT GAGAAAGTTA AGTTGAGAGC GATTAAAGTG AGTAGCTGTG ATAAGCTTGT GAATCTATTT
4761 CCGCGCAATC CCATGTCCTT GTTGCAATCAT CTGGAAGAGC TTACAGTCCA GAATTCGGGT TCCATTGAGT
4831 CGTTATTCAA CATTGACTTG GATTGTGTG GTGCAATTGG AGAAGAAGAC AACAGAGGCC TCTTAAGAAG
4901 CATCAACGTG GAGAATTTAG GGAAGCTAAG AGAGGTGTGG AGGATAAAG GTGCAGATAA CTCTGATCTC
4971 ATCAACGGTT TTCAAGCTGT TGAAGCATAA AAGATTGAAA AATGTAAGAG GTTTAGAAAT ATATTACAC
5041 CTATCACCOC CAATTTTTAT CTGGAGGCAC TTTTGGAGAT TCAGATAGAA GGTTCGGGAG GAAATCACGA
5111 ATCAGAAGAG CAGGTAACGC TTTCATTTTC ACTTCTTAA TTAATTAAGG ACTAAGCTCC TGTTTTGTGA
5181 ATAATAAGAA GGTGGATGA CTAACTTGG GCATCACAAT TGCAACAAA TGTTCACAA CATTGAAACGT
5251 TCAAAACCTT TCTTGAATTA AGGTTTCAAT ACAAGTCATT TAAAAATATG GCTTAAATTT TTTTATATT
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5461 TTTAATAAGT TAACAACCTA AATGTCAATT CCTAACAAA AAAACTTTTG TTCAGAAAAA CCGCAATTCA
5531 AGATAACTAA AATAAAAAA TTTGACATTC ACTAAGAGCA TTTTTTTTTC TAAATATGAT TGCAAAATGAA
5601 TAAAACTTAA ATTTATACAG AAAATCTTTT TATATATGTT ATACAAAATT TACAAATGTA AATTGGATAT
5671 GTTAATTAA CTTTATAAT TCTGGTATCA CAAAGGGATA TATAATAAAA TATTATTTTC TGTAGTCATT
5741 TGTAAATTGA CTAGTTTATA ACCCGTGGGA ACCATGAGTT CTAAAAATTG TTAACCTTTC ATAATAAAAA
5811 TTTTATAATTA TTATTATTTT TAAATAAATT ATTAATTAA AGATATATCA AAAAATTAAA GTTATTATAA
5881 CTTCAAAATT AACATATAAT TAGAAAAAT ATGATCATAA CTTCTGCAT CTCTTTGTAT AAATGCAGAG
5951 AAGCTATTAG TATATTTCTA ATCAAGTCCA AACCTAATGA AGCCTATATA ATTTTGTGAA AACCTAATTA
6021 GCATTAGGTT TTAAGAGTCA CCAAAATCAA AGAATAATCC AATGCTTTCA TTACCACTAT AATGGAATAA
6091 TTTCTTAGT TTAATGAAA TGAACAACAA CATTCAAAT AATTGTTGCT TATTAAACCA AAGACCCATT
6161 ACTTAGCCAA GAGTTTAAAC AAAAAAATT ACATTCAATG ATCATTATTC ATGACTAGAT ATATATGAAC
6231 GTTCTTCTAA GTTCTTCAAG TCTCCAGTGT CTCTTAATAA TGCAAGGAG AGCCATTAAA TTCTGATGTA
6301 CCAAGTTATT CAAGAAATTA CATCCAAGTC AACCAAGAG AAGTTTAGCC TAGCATGGCT AAACCTAAGA
6371 AACTAAAAA AGGATTAGAA GTACCAACAA TGTAGTAAGA ATCAGAGTAA AAGATGATGT TGTCTTGAT
6441 GTTCTTCTAA GTTCTTCAAG TCTCCAGTGT CTCTTAATAA TGCAAGGAG AGCCATTAAA TTCTGATGTA
6511 TTGATCCCTT CAAAAGCTGC ACCAACCTCC CTTAAATAAC ACTCAAAGCA AAAATGACAA AATGCCCTGA
6581 AGGACCCCTT GTGGGTGCTT TCGCGGGGTG GAGCTGCATA CGAAAGGTCT TTGGTCTTTG TGAGGGTGAT
6651 GTTGTGCGG ATAGCTTGTG GCATGCTTCC GCGCGGTTC CACACATGTG CACAGGTGAT GCATGGTGTG
6721 TCGCTTCTTG AGTTTGTAGC CTCCGATGCT TAGTCCACTT GCGCCCAATC GAGTCCAATC AGCTTATAAC
6791 CCATTTTCT TCAAGTTATC TTCAAAGTTA GCCCAATTG GCTTCTCCAA ATCATCCATA ACTTCACAGA
6861 ATGCGCCGTT CATCTTAATC CCGGATGCAC AATTATTCTC CGCTCTTCAT TTAAAGCAAG ATACCACCTT
6931 CTTTATGCTT CATCCATCAA TAGTACACTT CATGTATCAT CTCTACTAGT TATTTAGTCC ACAAATCCCT
7001 GTTGTCTCC AAATTTAATT ATCTCAATTA GTTCCCGGT COGCTACTTT CCTTAAATTT TGAATTAAG
7071 CTCAGAGAAA TATTAAGTAC CCGAAATGGT CATAAAATTA ACAAAGGA AAATGCATGA AGATTAACCTA
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7211 CACCCCTATG GCTTGTAGTC CACCCACCTT TCATTCCTTG TACCAATATG GGATGGAAC ATCATTAAAT
7281 AAGCCAAAAA GCTAACATAT AAGGGTTTAG TGACAAAGGT AAGTACTAAA GATGAAAAA ATCCATTTT
7351 CTTGTTTTTA CACAACACAC ACATAGGGGC AGACGTAGGA TTTCAAAGTA CAGATTGTTG GTGGCACATA
7421 AGTGTGCTG GTGACATTTT TTTTCTCTT TTACGTGGTG GCACAACAGT AGGAAAAACG AAAAAATCGA
7491 AATTTTCTT AATTTGTCTT AAAAAAACA GGGGTGTGTG GTGCCACTAT GGCAACAAA GTTGAACCTG
7561 CCTACCGCGG CACACACACA CACACACATA GAGAGAGAGA GAGAGAGAGA GAGAGAGAGA AAGAAAGAAA
7631 GAGAGAGAGA GTTGGGATG TGATACTTCT TTTAGGAAAA TGGAGTTATA TCTTTGATAT TGTATTTTTT
7701 TAAATGAATT TATNTATTTA ATCATTTTAG TTTATAAGTT NTATTTATIN GGNATGAAA AAAAAAGTCT
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7841 TATATATTA ATTCACAATA GTCTTTAGGA ATAGTATTAT ATATATAATT AATTTCTCAAT GGTCTTAGGA
7911 ATAGTAAGTT CTTATATTTT AAACCTTTTG CACAATTTCT TGCTTACTTT GACACTTTTC CTCTCTAAGT
7981 TTACTATAT ATATATATTA AAGCGCAAAG GTCAATAGGA TATAATATTT TCTATTATTC TACGTTTTG
8051 CACAAAAGTT TGAACACTTT GCCACTTTTT GTCCCTCTTT AACCTTTTCA ATGTTTTGCG ACAAAGTTTC
8121 CAAAACCTTG CCACTTTGAT CATTCCTCAA CTTTTCACCG CATTAGTTTG TGGAGTTGGC AGTTTTGGTC
8191 CCTCTACCT CGATATTCTC TACTGCTAGC CAAAAGGGT TCCAGAGTTT CACACTTTTG GTCCCTGACA

RLG 2B cont.

8261 GTAACCAAT GTGAGATGTC AAATTTTTC CACATTAGTT TGTGGAGTTG TCCCTTTTGG TCCCCCACA
8331 TTGCATATTC TACTATACGA TCTTATTTT CTCAAATAAC AACACGTATA TTTTATC:CT AATTGGAAAA
8401 AGAGTTTTTAA AA:AAATAAC GACTAGG::: G:GC:GAGTT TTTTTT:ACA AGTTTGTATC AAATCATATC
8471 AAAATTTAAG GTGGAACGGT GACCACATTA ACCAGAAATG TAATTTATTC TTTGATTTTG ATAATTTTTTA
8541 ATATTTTGTG GTGATCTATG TATTAAAAAG TAAACAACAA AGAACATAAT CCAAAACCCCT AAATTGCAAG
8611 TCTCGCCCAA TTTCTCTATC ACTAGTCCCTC ACTTACGATG GCGTTACGTC GCTCTCTCAC TGCTTACAAC
8681 CCTTTGTGTC TACTCATTAC AATAACGAAA AGTTGAATAT CCATATATTT ATTTGGATGT GGAATTGAAC
8751 GAATCTCGTC AAAATTTTGA TTTTGTGAT GGATTTGAGT AGAAGTTTGG GCAGAACGGG AATGATGGTC
8821 TGCAAGTGGT TATAAACTTG ATTTGAGTT ATTACTATAT ATGTAGCCTC TTTACAACGA CCAAGGTTTC
8891 TTCCAGGTAC CATTTGATCT TTTTAGAATC TAGTTTCTG AAACACCCCTG ATTTGGATCA AATATCACCA
8961 ACAACTCTTA AAAACTTGAT TAATCAATG TTTTCTTCAT CTGTATAACA AGTGGAAATGA TTTTCTACTT
9031 AGATTAACCT GAAAAAAG GTCCATGTGC GTCTGGTGA TCTGGTAAAT GAAGATGGAA GGGAGAGCTG
9101 ACTTTAAAGA CACAAACACG TCACCATATC TCTTATTTTA TTTTAAATTT GCTTTTGTG TATTTTCTTT
9171 TTTCTCTTTT CTTTCTTTCT TGATCTCCAG ATGGTATG TGTTGGATAA TTTACACCTA GAGATTGGGA
9241 ACGATGGGAA GGGGTCTGTG ATTTATGGCT GGCCGAGTTT TACTTATTA CTCAATTTCA ACCTAAATTC
9311 TGATTCCTGT TTGAAAAA GTTGCACTCT TATTTTGTG TTATCTGTGT GCATAGGATC CTTAGCATCT
9381 TTTAATAATT TATTTGAAGG TGAAAGATCC AACTATTTTT TAGCTGTGG CATTTTCCAT CATTTGCAAC
9451 TGTTCCTTGA AAAAAAATA CCTAAATAA AAATAACCAT TTTCAAATCC AAAATTATAA GAGAGAATTG
9521 TAAATGGACA TGGATCATA AATCATTAAC ACAGTTTCA GTTGAAGTTG CTAATTACAT TCTTGCTGT
9591 GCAGATTGAA ATTTCTATCAG AGAAAGAGAC ATTACAAGAA GCCACTGGCA GTATTTCAA TCTTGTATTC
9661 CCATCCCTGC TCATGCATC TTTTCATAAC CTCCTGTGTC TTACATTTGA TAATATGAA GAGATGGAGG
9731 TGGTATTTGA GATAGAGAT GAGAGTCCAA CATGTAGAGA ATTGGTAACA ACTCGCAATA ACCAACACA
9801 GCCTATTTATA CTTCCTTACC TCCAGGATTT GTATCTAAGG AATATGGACA ACACGAGTCA TGTGTGGAAG
9871 TGCAGCACT GGAATAAATT CTTCACTCTT CCAAAACAAC AATCAGAATC CCCATTCCAC AACCTCACA
9941 CCATAAATAT TCTTAAATGC AAAAGCATT AGTACTTGT TTCGCTCTC ATGGCAGAAC TTCTTTCCAA
10011 CCTAAAGGAT ATCCGGATA GTGAGTGTGA TGGTATTAAA GAAGTTGTTT CAAACAGAGA TGATGAGGAT
10081 GAAGAAATGA CTACATTTAC ATCTACCCAC ACAACCACCA CTTTGTTCCT TAGTCTTGAT TCTCTCACTC
10151 TAAGTTTCTT GGAGAATCTG AAGTGTATTG GTGGAAGTGG TGCCAAGGAT GAGGGGAGCA ATGAAATATC
10221 TTTCAAATAT ACCACTGCAA CTACTGCTGT TCTTGATCAA TTTGAAGTAT GCTTTGTACA TATTCATTA
10291 TTTATTTAAT TTCTTTTAT ATTTGCAATA TTCTATAAAT AATACATTTT ATACCCACTA TACTAAGATA
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10431 GGAAGTTCAA TAAATGATA ATGGCATCTT TTGATGGGTA ATATAGGCAA TTTAAGTTT ATTTCTGTTA
10501 AAGCAGTATT TAGCAAGTAC TGGCCAGTAG GAGAGGAGAA TATCACCTTT TGTGAAATC TGGTCATTGT
10571 ACCCAGAATT TAGTTAAATG TAACATTTTA GATATTAGGG GTTATCAGGT GACAGATATT GTAGAATAGA
10641 ACAATATGTA ATATTACCCA AAACATTTTT TTCTAAGGTT GCTCTGTAA ATATGTGCTT TCTTGATTTT
10711 ATTGAATTTG CATTCCTATA TTTTAGGTGG TAAAGTGATT GTCTCTTCAA TAAATCCCGA AATTTTTTAA
10781 TTAATAAATA AAAAAACAA AGTAAATTTT TGATATGGAG AGCACTGGTA TCATTTAGTA TATAAAAAAC
10851 AGATTTTGA TTAAGTTTCT TATATAAAG CTGTGTATAT AGTTTAAATTA GTTTTACATC ATTTTCCAT
10921 GTGGTGTGTC AGTTGTCTGA AGCAGGTGGT GTTCTTGTGA GCTTATGCCA ATACGCTAGA GAGATAAAAA
10991 TAGGCAACTG CCATGCATTG TCAAGTGTGA TTCCATGTTA TGCAGCAGTA CAAATGCAGA AAGCTT

SEQ ID NO: 23

RLG2B a.u.

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SMNASTSASLSDDFPSREKTFTQALIALEPNQKFHMVALCGMGGVGKTRMMQRLKKA?EEKL FNYIV
GAVI?EKTDPFAIQEAIADYLG IQLNEKTKPARADKLREWFKKNSDGGKTKFLIVLDDVWQLVDLEDIGL
SPFPNQGVDFKVLTSRDSQVCTMMGVEANSIINVGLL TEAEASL FQQFVETSEPELQKIGEDIVRKC
CGLPIAIKTMAC?LRNKRKDAWKDALSRIEHYDIHNVAPKVFETSYHNLQEEETKSTFLMCGLPEDFDI
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GNHESEEQVTL SISLS

SEQ ID NO: 24

[illegible]

RLG2A
RLG2B
RLG2C
RLG2D
RLG2E
RLG2F
RLG2G
RLG2H
RLG2I
RLG2J
RLG2K
RLG2L
RLG2M

CA TTC TAT TTA CCA GAG CTG TGA CCA GAT T A C T C T C T G T A T A G A C T G A A A A A A C A C T A A A C A C G A A G A C T T A A T A A G C T T C C G T A A T G G T T C T A A G G C C
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 C T T T C C A T T C C A A G A G C T T A G C A G A T A A C C T G G T A T A C A C T A T T A A C C T A T T A A C C A G A A G C T G T A T A G C T T C T G A A T G G T T T C A A A A G C
 C A T T C C A A G A T T G T C A A G A T T A C C T A C G A T C A A G A A A A C T G A A A A C A A G A C T G A A A A C A G A C T T C G T A A A T G G T T T C A A A A G C
 C A T T C C T A T T C A G A A G T T G T A G C A T T A C C T A T A G C T A G C A T A A A A C G C A T A A A C A A G A C T G A A G A C T T C G T C A A G G T T T C A A G G C C
 C A T T C C T A T T C A G A C T T A G C A G A T T A C C T T T G T A T A G G T A T A A A A A C C A C C A G C A T A T A G C T C C T G A A G A C T T C G T A A T T G G T T T C A A G G C C
 A G G T T C C A T T C A G A C T T A G C A G A T T A C C T A G T G A G C T C A T A T A A A A T T A A G A C A G A A G C T G T A T A G C T C C T G A A G A C T T A A G C C T G A A G C C
 C T T T C C A T T C C A A G A C T T A G C A G A T T A C C T C T T A T A G A C T T A A A A A C T A T A A G A C A G A A C T G T A T A G C T T C T A A A T C G G T T T C A A G C C
 T A T T C C T A T T C A G A C A G C T T A G C A G A T T A C C T C T T A T A G C T C A A G A A A C T A A G A C A G A A C T T A A G A C T T C T A A A T T G G T T T C A A G C C
 T A T T C C T A T T C A G A C A G C T T A G C A G A T T C C C T C T C T A G A C T C A A G A A A C T A A A G A C A A G A C T T A A G C T T C G T A A T T G G T T T C A A G C C
 C A T T T C C A T T C A G A C A G C T T A G C A G A T T A C T C T C T T A G A C T C T A A A A A C A G A C A T A A G C A A G A A C T T A A G C T T C G T A A C G G T T T C A A G C C
 C A T T T C C A T T C A G A C A G C T T A G C A G A T T A C C T G C T T A T A G A C T A A A A A C A G C A C A C A G A C T G T A T A G C T T C G T A A T G G T T T C A A G C C
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 C A T T T C C A T T C A A G A C T T A G C A G A T T A C C T C T A T A G C T C A A G A A A C T A A G A C A A G A C T T A A G C T T C G T A A T T G G T T T C A A A A G C

RLG2A
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RLG2C
RLG2D
RLG2E
RLG2F
RLG2G
RLG2H
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RLG2J
RLG2K
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RLG2D
RLG2E
RLG2F
RLG2G
RLG2H
RLG2I
RLG2J
RLG2K
RLG2L
RLG2M

RLG2A	810	820	830	840	850	860	870	880	890	900
RLG2B	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2C	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2D	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2E	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2F	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2G	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2H	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2I	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2J	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2K	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2L	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2M	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2N	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2O	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2P	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2Q	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2R	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2S	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2T	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2U	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2V	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2W	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2X	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2Y	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2Z	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2A	910	920	930	940	950	960	970	980	990	1000
RLG2B	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2C	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2D	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2E	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2F	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2G	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2H	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2I	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2J	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2K	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2L	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2M	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2N	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2O	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2P	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2Q	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2R	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2S	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2T	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2U	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2V	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2W	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2X	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2Y	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC
RLG2Z	GC	TC	AA	CA	CC	TC	CA	AG	CC	TC

[illegible]

RLG2A	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2B	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2C	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2D	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2E	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2F	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2G	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2H	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2I	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2J	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2K	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2L	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2H	1210	1220	1230	1240	1250	1260	1270	1280	1290	1300
RLG2A	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2B	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2C	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2D	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2E	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2F	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2G	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2H	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2I	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2J	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2K	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2L	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400
RLG2H	1310	1320	1330	1340	1350	1360	1370	1380	1390	1400

1410 1420 1430 1440 1450 1460 1470 1480 1490 1500
 CTTAAAAAATTTGGTCAAACTGGAGGCTCTATAT-GACAGTG-----GTT-----CGAGGTGCAAGG-----CGA----- 1437
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1475
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1417
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1411
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1432
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1437
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1466
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1420
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1442
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 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1398
 CTTGAAAAAGTTTGGTCAAACTGGAGGCTCTATATATTTGAGATGCAATCTGGGTTATAGATGATAACTGCAATGATGCGAGAGCGTTCTTACAACTT 1441

XXXXXXXXXXXXXXXXXXXX

1510 1520
 TCTGCAATGAAATTCGGGTTCTTTA -TTA- SEQ ID NO: 27
 -SEQ ID NO: 28
 -SEQ ID NO: 29
 -TG- SEQ ID NO: 30
 -TA- SEQ ID NO: 31
 -TA- SEQ ID NO: 32
 -SEQ ID NO: 33
 -SEQ ID NO: 34
 -SEQ ID NO: 35
 TGGAAATGGCAGAGCTTTTTCACA -SEQ ID NO: 36
 TCGATATCAAAAGGC -SEQ ID NO: 37
 -A- SEQ ID NO: 39

1439
 1500
 1417
 1412
 1432
 1438
 1466
 1420
 1442
 1474
 1437
 1398
 1441

SEQ ID NO:

40

RG2A protein in GRTTHMRLKGVKQKRNPNF IENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 98-41
 RG2B protein in OKTHMRLKGVKQKRNPNF IENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 100-42
 RG2C protein in NTRX--AKAEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 98-43
 RG2D protein in EVAK--RK--FGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 90-44
 RG2E protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 99-45
 RG2F protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 100-46
 RG2G protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 97-47
 RG2H protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 89-48
 RG2I protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 94-49
 RG2J protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 89-50
 RG2K protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 100-51
 RG2L protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 82-52
 RG2M protein in GRND--AKVEVAKKKEFGYIENVIGKNTPTAIQANADYVIGELNKTKEPARTKJRMFVNSGG--KKIIVTILXMAQFVDI NDIGI SPLPQQ 92-53

RG2A protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 196
 RG2B protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 195
 RG2C protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 193
 RG2D protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 185
 RG2E protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 193
 RG2F protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 195
 RG2G protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 189
 RG2H protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 194
 RG2I protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 189
 RG2J protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 195
 RG2K protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 177
 RG2L protein in VDFKVLITSRDSHIVCTVGVANSILNVLGLTEAEASLSFQQFVETS--E---PELQKIGEDIVRKCGGLPIAIKTHACTILNRKRDAMKDALSRLEHYD 187

[illegible]

[illegible]

SEQUENCE:

10 20 30 40 50 60 70 80 90 100
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG → 54
 20 30 40 50 60 70 80 90 100
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 97-56
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 97-57
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 96-58
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 96-59
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 95-60
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 97-61
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 96-62
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 74-63
 C-CANAAACAAATTGAAACCCGGATCATCCAAATACATCATCCACTTCCAAATGACCCCAATTCATTAACAA--GGTCTTAAGCTTGAAG 97-64
 ---GA---TAAATTAG-----TAACTCAATTTCAG--CCACTCTG 35-65
 TGTGAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 97-66
 AG-AGCAGACCGTGAACCTTGGATGGTAAATTTAGTAACTTAACG-CCTTC--AATTAACTTACCTTTTCTTATTAACCTAATTTCAGCTAAATTCG 85-67

110 120 130 140 150 160 170 180 190 200
 AATCTGTGTTGAAAATTAAGTT-GCATCTTTTATTTT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG
 110 120 130 140 150 160 170 180 190 200
 AATCTGTGTTGAAAATTAAGTT-GCATCTTTTATTTT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG 187
 AATCTGTGTTGAAAATTAAGTT-GCATCTTTTATTTT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG 187
 AATCTGTGTTGAAAATTAAGTT-GCATCTTTTATTTT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG 186
 AATCTGTGTTGAAAATTAAGTT-GCATCTTTTATTTT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG 188
 TCTCTGTGTTGAAAAT-TATCCCATCTTT-ACCTTAAATATTTAT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG 137
 AATCTGTGTTGAAAATTAAGTT-GCATCTTTTATTTT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG 176
 AATCTGTGTTGAAAATTAAGTT-GCATCTTTTATTTT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG 164
 CCAATCTGTGATTTACCAACCCGCAATCCAAACAGCCGCTACATA--ATTG-TGTGGTTTGTGTTTATTTTCCGCTGAGGTG 194
 TCTCTGTGTTGAAAAT-AGTTCCATCTTT-TATTAATAATGTTTTCATAGAGCATATAGAGCTTTTCAATATTTATTT-----GAAGGTG 122
 AATCTGTGTTGAAAATTAAGTT-GCATCTTTTATTTT-TG---TATTATCTGTGTGATAGCATCTCT-TAGCATCTTTTAAATTTATTT-----GAAGGTG 187
 ---TCT-----TATTA--TGCACATCTTAAATTAATAAGTATTTAATTTGTTGATAGCATCTCTTAAATTTATTTATTT-----GAAGGTG 168

AC15-2A
 AC15-2B
 AC15-2C
 AC15-2D
 AC15-2E
 AC15-2G
 AC15-2H
 AC15-2I
 AC15-2J
 AC15-2L
 AC15-2N
 AC15-2O

AC15-2A
 AC15-2B
 AC15-2C
 AC15-2D
 AC15-2E
 AC15-2G
 AC15-2H
 AC15-2I
 AC15-2J
 AC15-2L
 AC15-2N
 AC15-2O

210 220 230 240 250 260 270 280 290 300
 AAGAAGCATCTATTTTAACTCTGGCATTTTGCATCATTTGCACTGTTCTTGAAAAA---TACTCTAAATTCAAAATACCAATTTTCAATTC 286
 AAGAAGTCCACTATTTTAACTCTGGGCHTTTCCATCTTTGCACTGTTCTTGAAAA---ATACTTAAATTCAAAATACCAATTTTCTATTC 284
 AAGAAGTCCACTATTTTAACTCTGGGCHTTTCCATCTTTGCACTGTTCTTGAAAAA---TACTCTAAATTCAAAATACCAATTTTCAATTC 285
 AAGAAGTCCACTATTTTAACTCTGGGCHTTTCCATCTTTGCACTGTTCTTGAAAAA---TACTCTAAATTCAAAATACCAATTTTCAATTC 286
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[illegible]

AC15-2A
AC15-2B
AC15-2C
AC15-2D
AC15-2E
AC15-2G
AC15-2H
AC15-2I
AC15-2J
AC15-2L
AC15-2N
AC15-2O

AC15-2A	AC15-2B	AC15-2C	AC15-2D	AC15-2E	AC15-2F	AC15-2G	AC15-2H	AC15-2I	AC15-2J	AC15-2K	AC15-2L	AC15-2M	AC15-2N	AC15-2O	AC15-2P	AC15-2Q	AC15-2R	AC15-2S	AC15-2T	AC15-2U	AC15-2V	AC15-2W	AC15-2X	AC15-2Y	AC15-2Z	AC15-2AA	AC15-2AB	AC15-2AC	AC15-2AD	AC15-2AE	AC15-2AF	AC15-2AG	AC15-2AH	AC15-2AI	AC15-2AJ	AC15-2AK	AC15-2AL	AC15-2AM	AC15-2AN	AC15-2AO	AC15-2AP	AC15-2AQ	AC15-2AR	AC15-2AS	AC15-2AT	AC15-2AU	AC15-2AV	AC15-2AW	AC15-2AX	AC15-2AY	AC15-2AZ	AC15-2BA	AC15-2BB	AC15-2BC	AC15-2BD	AC15-2BE	AC15-2BF	AC15-2BG	AC15-2BH	AC15-2BI	AC15-2BJ	AC15-2BK	AC15-2BL	AC15-2BM	AC15-2BN	AC15-2BO	AC15-2BP	AC15-2BQ	AC15-2BR	AC15-2BS	AC15-2BT	AC15-2BU	AC15-2BV	AC15-2BW	AC15-2BX	AC15-2BY	AC15-2BZ	AC15-2CA	AC15-2CB	AC15-2CC	AC15-2CD	AC15-2CE	AC15-2CF	AC15-2CG	AC15-2CH	AC15-2CI	AC15-2CJ	AC15-2CK	AC15-2CL	AC15-2CM	AC15-2CN	AC15-2CO	AC15-2CP	AC15-2CQ	AC15-2CR	AC15-2CS	AC15-2CT	AC15-2CU	AC15-2CV	AC15-2CW	AC15-2CX	AC15-2CY	AC15-2CZ	AC15-2DA	AC15-2DB	AC15-2DC	AC15-2DD	AC15-2DE	AC15-2DF	AC15-2DG	AC15-2DH	AC15-2DI	AC15-2DJ	AC15-2DK	AC15-2DL	AC15-2DM	AC15-2DN	AC15-2DO	AC15-2DP	AC15-2DQ	AC15-2DR	AC15-2DS	AC15-2DT	AC15-2DU	AC15-2DV	AC15-2DW	AC15-2DX	AC15-2DY	AC15-2DZ	AC15-2EA	AC15-2EB	AC15-2EC	AC15-2ED	AC15-2EE	AC15-2EF	AC15-2EG	AC15-2EH	AC15-2EI	AC15-2EJ	AC15-2EK	AC15-2EL	AC15-2EM	AC15-2EN	AC15-2EO	AC15-2EP	AC15-2EQ	AC15-2ER	AC15-2ES	AC15-2ET	AC15-2EU	AC15-2EV	AC15-2EW	AC15-2EX	AC15-2EY	AC15-2EZ	AC15-2FA	AC15-2FB	AC15-2FC	AC15-2FD	AC15-2FE	AC15-2FF	AC15-2FG	AC15-2FH	AC15-2FI	AC15-2FJ	AC15-2FK	AC15-2FL	AC15-2FM	AC15-2FN	AC15-2FO	AC15-2FP	AC15-2FQ	AC15-2FR	AC15-2FS	AC15-2FT	AC15-2FU	AC15-2FV	AC15-2FW	AC15-2FX	AC15-2FY	AC15-2FZ	AC15-2GA	AC15-2GB	AC15-2GC	AC15-2GD	AC15-2GE	AC15-2GF	AC15-2GG	AC15-2GH	AC15-2GI	AC15-2GJ	AC15-2GK	AC15-2GL	AC15-2GM	AC15-2GN	AC15-2GO	AC15-2GP	AC15-2GQ	AC15-2GR	AC15-2GS	AC15-2GT	AC15-2GU	AC15-2GV	AC15-2GW	AC15-2GX	AC15-2GY	AC15-2GZ	AC15-2HA	AC15-2HB	AC15-2HC	AC15-2HD	AC15-2HE	AC15-2HF	AC15-2HG	AC15-2HH	AC15-2HI	AC15-2HJ	AC15-2HK	AC15-2HL	AC15-2HM	AC15-2HN	AC15-2HO	AC15-2HP	AC15-2HQ	AC15-2HR	AC15-2HS	AC15-2HT	AC15-2HU	AC15-2HV	AC15-2HW	AC15-2HX	AC15-2HY	AC15-2HZ	AC15-2IA	AC15-2IB	AC15-2IC	AC15-2ID	AC15-2IE	AC15-2IF	AC15-2IG	AC15-2IH	AC15-2II	AC15-2IJ	AC15-2IK	AC15-2IL	AC15-2IM	AC15-2IN	AC15-2IO	AC15-2IP	AC15-2IQ	AC15-2IR	AC15-2IS	AC15-2IT	AC15-2IU	AC15-2IV	AC15-2IW	AC15-2IX	AC15-2IY	AC15-2IZ	AC15-2JA	AC15-2JB	AC15-2JC	AC15-2JD	AC15-2JE	AC15-2JF	AC15-2JG	AC15-2JH	AC15-2JI	AC15-2JJ	AC15-2JK	AC15-2JL	AC15-2JM	AC15-2JN	AC15-2JO	AC15-2JP	AC15-2JQ	AC15-2JR	AC15-2JS	AC15-2JT	AC15-2JU	AC15-2JV	AC15-2JW	AC15-2JX	AC15-2JY	AC15-2JZ	AC15-2KA	AC15-2KB	AC15-2KC	AC15-2KD	AC15-2KE	AC15-2KF	AC15-2KG	AC15-2KH	AC15-2KI	AC15-2KJ	AC15-2KK	AC15-2KL	AC15-2KM	AC15-2KN	AC15-2KO	AC15-2KP	AC15-2KQ	AC15-2KR	AC15-2KS	AC15-2KT	AC15-2KU	AC15-2KV	AC15-2KW	AC15-2KX	AC15-2KY	AC15-2KZ	AC15-2LA	AC15-2LB	AC15-2LC	AC15-2LD	AC15-2LE	AC15-2LF	AC15-2LG	AC15-2LH	AC15-2LI	AC15-2LJ	AC15-2LK	AC15-2LL	AC15-2LM	AC15-2LN	AC15-2LO	AC15-2LP	AC15-2LQ	AC15-2LR	AC15-2LS	AC15-2LT	AC15-2LU	AC15-2LV	AC15-2LW	AC15-2LX	AC15-2LY	AC15-2LZ	AC15-2MA	AC15-2MB	AC15-2MC	AC15-2MD	AC15-2ME	AC15-2MF	AC15-2MG	AC15-2MH	AC15-2MI	AC15-2MJ	AC15-2MK	AC15-2ML	AC15-2MN	AC15-2MO	AC15-2MP	AC15-2MQ	AC15-2MR	AC15-2MS	AC15-2MT	AC15-2MU	AC15-2MV	AC15-2MW	AC15-2MX	AC15-2MY	AC15-2MZ	AC15-2NA	AC15-2NB	AC15-2NC	AC15-2ND	AC15-2NE
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AC15-2A	GTAACTACTCAATCAACAAACAAAC--CTAT--TATACCTTCCCAACCTCGAGATTTGATCTATAGGAATATGCAACAACATGATCTATGTGCA	630	640	650	660	670	680	690	700
AC15-2B	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCTTACTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	631	641	651	661	671	681	691	701
AC15-2C	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCTTACTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	632	642	652	662	672	682	692	702
AC15-2D	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCAACCTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	633	643	653	663	673	683	693	703
AC15-2E	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCAACCTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	634	644	654	664	674	684	694	704
AC15-2F	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCTTACTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	635	645	655	665	675	685	695	705
AC15-2H	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCAACCTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	636	646	656	666	676	686	696	706
AC15-2I	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCAACCTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	637	647	657	667	677	687	697	707
AC15-2J	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCAACCTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	638	648	658	668	678	688	698	708
AC15-2L	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCAACCTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	639	649	659	669	679	689	699	709
AC15-2N	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCAACCTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	640	650	660	670	680	690	700	710
AC15-2O	GTAACTACTCGAATAATACCAACACAGC--CTAT--TATACCTTCCCAACCTCGAGATTTGTATCTATAGGAATATGCAACAACATGATCTATGTGCA	641	651	661	671	681	691	701	711
AC15-2A	AGTCGACGACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATATGCAAAAGCAT	710	720	730	740	750	760	770	780
AC15-2B	AGTCGACGACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	711	721	731	741	751	761	771	781
AC15-2C	AGTCGACGACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	712	722	732	742	752	762	772	782
AC15-2D	AGTCGACGACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	713	723	733	743	753	763	773	783
AC15-2E	AGTG---CAACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	714	724	734	744	754	764	774	784
AC15-2G	AGTGCGACACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	715	725	735	745	755	765	775	785
AC15-2H	AGTGCGACACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	716	726	736	746	756	766	776	786
AC15-2I	AGTGCGACACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	717	727	737	747	757	767	777	787
AC15-2J	AGTGCGACACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	718	728	738	748	758	768	778	788
AC15-2L	AGTG---CAACCGGTTAGGTTCTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	719	729	739	749	759	769	779	789
AC15-2N	AGTGCGGCAACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	720	730	740	750	760	770	780	790
AC15-2O	AGTGCGGCAACTGGAATAAATCTTCACTCTCTCCAAACACAACTGACATGATCCCTTCCCAACCTCGACCTCAACCTCAATTAATATTTATGTCAAAAGCAT	721	731	741	751	761	771	781	791

SEQ ID NO:

AC15-2A	810	820	779
AC15-2B	TAAGTACTTGTGTTTCACTCTCAAGG - 56		777
AC15-2C	TAAGTACTTGTGTTTCACTCTCAAGG - 57		777
AC15-2D	TAAGTACTTGTGTTTCACTCTCAAGG - 58		788
AC15-2E	TAAGTACTTGTGTTTCACTCTCAAGG - 59		721
AC15-2F	TAAGTACTTGTGTTTCACTCTCAAGG - 60		781
AC15-2G	TAAGTACTTGTGTTTCACTCTCAAGG - 61		738
AC15-2H	TAAGTACTTGTGTTTCACTCTCAAGG - 62		722
AC15-2I	TAAGTACTTGTGTTTCACTCTCAAGG - 63		784
AC15-2J	TAAGTACTTGTGTTTCACTCTCAAGG - 64		699
AC15-2K	TAAGTACTTGTGTTTCACTCTCAAGG - 65		778
AC15-2L	TAAGTACTTGTGTTTCACTCTCAAGG - 66		763
AC15-2M	TAAGTACTTGTGTTTCACTCTCAAGG - 67		

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SEQ ID NO: 68

RLG3 (real RLG3)
[Strand]

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1  AATGGCAAAA GAAGTCGGAG CAAGAGCTAA GTTAGAGCAT CTATTGACG TCATTATCAT GGTAGATGTC
71  ACTCAAGCAC CCAACAAGAA CACAATTCAA AGTAGTATTT CAGAACACTT GGGATTAAAA CTGCAAGAAG
141 AGAGCTTGTT GSTAAGAGCA GCTAGGGTAA GTGCGAGGTT AAAAATGCTT ACAAGGGTGC TGGTGATATT
211 AGACGATATA TGGTCAAGGC TTGACATGGA GGAACCTGGG ATTCCCTTTG GATCAGATAG ACAACACCAC
281 GGCTGCAAAA TCTTGTTGAC TTCAAGAAGT ATTAGTGCTT GTAACCAGAT GAGAGCTGAT AGAATCTTTA
351 AAATACGAGA AATGCCACTG AATGAAGCAT GGCTTCCTTT CGAAAGAACA GCTAAAAAAG CTCCGAATCT
421 GCATCAAGTA GCAAGAGATA TCGTGGAGGA GTGTGGTGGG C
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RLG4
SEQ ID NO: 69

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1  GAATTCGGTG  TTGGTAAGAC  AACTCTTGCC  TCTTCTGTTT  ATGATGAAAT  CTCTAGCAAG  TTTGATGGTT
71  GCTGCTTTCT  AAAAATATCT  GGGAGGAATC  AAGTAATAAA  GACGGTATAG  AAAGATTGCA  AGAAAAATC
141  ATTTGTGATG  TTTTGAAACA  AGAGCAAGTG  GCGGTAGGGA  GAGTTGAAGA  AGGAAAGCGC  ATGATAAAGG
211  ATAGGTTACA  ACATAGAAAG  GTATTGATTG  TCGTTGATGA  TGTGACAAC  GTTGAGCAGC  TAGCTAGAAC
281  AGTTGGCTGG  ATCACATGAT  TGGTTTGGTG  AAGGTAGCCG  CATAATAATC  ACAACTAGAG  ATGAACATGT
351  ATTAATTGCA  CACAAAGTAG  ATGTGATACA  CAATATAAGC  TTGTTAAACA  ACGATGAAGC  TATGCATCTC
421  TTCTGCAAGC  AAGCACCACG  GGGTCACAAA  CGTATACAAG  ATTATGAGCA  ACTTTTAAAA  CATGTGGTTT
491  CTTATGCTGG  TGGGCTTCCA  CTAGCACTGT  CGAC
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SEQ. ID NO: 70
FLG1-EL69
[Strand]

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1 ATCGTAACCG TTGCTACGAG AAGCGCTGTC CTCCTTCATC TTTTGTGATA TGTGATATTC TCATNNATTH
71 TGCCACATAT AATTTTGTGG TTATTTTAAA TTAATTTTAA TTCCACATGT CATTTTATGA GTTTTCTAT
141 TTTATTGAGT TTCCACATAAT ATTTAAATGT AATAACAATA AATGCATATT TATTTTCTT TAAATAAACG
211 CATATAATAT ATAGATTAAA ATCATATAAT ACATAGGTTA AACTCATATA ATACATATGT TCATCCCCAG
281 TTTATTATATA TGCTCATACC TTAATTTATT TATTATTAT TATTAGAGT AGATGATCIT TGTGATATTA
351 AAAATTTAAT TTGTTCAAAA TTAATAATTA TTAATAATCC CACAATTIGA ATAAAAATTA AAAAAATGNN
421 CCCACCATTA GTCCATCACT TTTTCAGCTC ATCAATATCG TGAGTATTCT CCTTCGTTTC CACCTTAATC
491 AATATTTCCA GCGAATGACA GACTCCTACG GCGTTTCGTA ATTTGCGTTC CGACACTGTT CATTTGAAGGA
561 GATAATAAAT CAAATGGAGC TGCTCCAATG TTCAATGCTG ATGAAAGGTG AATTTGATGT GAAGANAATG
631 TCAGCGATCT ATCTCCATCC GGAACCCACC ACATTATCAG TGTACCACCA AACCTCAAA AACGGVGGAA
701 GTAGRRACAC WRKAAAGTCA TGAAGAATAG ATTATTTTGT TCCTCATGGG CTGACTGAGG AGCGGGTTTA
771 GTTCATCATT TTTCTTTGAN CAAAGAATTA TCGGTCCATC GAATTTTAC ATCGACAAAG AAGTTTCACT
841 TCGCAATGTT TTGTTAAACA ATTTTAAATC TTTTATCTT TTCGTGAAA CTCTCAATT GCAACTTGCA
911 ACTTGCAACT TTTGGCCCA CAAATTTGTG GTGGGCGTTA ATTTAATCCA CATATTCACT GTAAACAATA
981 ATTCAAATCG ATCTCTGTTT ATCCAATTC TCAACATCTC TTGATAATTG AAATCATCA CGCTTCATCC
1051 ATTTTCATCCA CATCTATACT ATATCTCTCT CTCTTATCAT ATTTAAAGAT GGCTGAAATC GTTCTTCTG
1121 CCTTCTTGAC AGTGGTGT TTGAAAGCTGG CATYTGAAGC CTGTAAGAG ATTTGTTCCCT CCAAAGAAAT
1191 TGAATCTGAG CTTAAGAAAT TGAAGGAGAC ATTGACCAA ATCCAAGATC TGCTTAAAGA TGCTTCCAG
1261 AAGGAAGTAA CTAATGAAGC CGTTAAAGA TGGCTGAATG ATCTCCAACA TTTGGCTTAT GACATAGAG
1331 ACCTACTTGA TGATTTTGCA ACTGAAGCTG TTCAACGTGA GTTGAACGAG GAGGGTGGAG CCTCTCCAG
1401 TATGGTAAGA AAACATAATC CAAGTTGTTG CACAAGTTTC TCACAAAGTA ATAGGATGCA TGCCAAGTTA
1471 GATGATATTT CCACCAGGTT ACAAGAATCT GTAGAGGCAA AAAATAATCT TGGTTTAAAT GTGATAACAT
1541 ATGAAAGGCC AAAAATGAA AGGTATGAGG CGTCTTGGT AGATGAAAGC GGTACTGTCT GACGTGAAGA
1611 TGATAAGAAA AAATTGCTGG AGAAGCTGTT GGGGGATAAA GATGAATCAG GGAGTCAAAA CTTGAGCATC
1681 GTGCCCATTA TTGCTATGGG TGGAGTTGGT AAAACAATCT TAGCTAGACT TTTGTATGAT GAAAGAGAG
1751 TGAAGGATCA CTTCGAATCT AGGGCTTGGG TTTGTGTTTC TGATGAGTTT AGTGTTCCTA ATATAAGCAG
1821 AGTTATTTAT CAATCTGTGA CTGGGAAAAA GAAGGAGTTT GAAGACTTAA ATCTGCTTCA AGAAGCTCTT
1891 AAAGAGAAAC AACTAATCC GCTATTTCTA ATAGTTTGG ATGATGTGTG GTCTGAAAGC TATGCTGATT
1961 GGGAGAAATT AGTGGGCCCA TTCTTGGGG GGCTCTCTGG AAGTAGAATA ATCATGACAA CTCGGAAGGA
2031 GCAATTTGCT AGAAAGCTGG GCTTTTCTCA TCAAGACCTT CTGAGGGTTC TATCACAAGA TGATGCTTTG
2101 TCTTTGTTTG CTCAACACCC ATTTTGGTGA CCAAACCTTG ATTCACATCC AACACTAAG CCACATGGAG
2171 AACTCTTTTG GAAGAAATGT GATGGCTTAC CTCTAGCTTT AAGAACACTT GGAAGGTTAT TAAGGACAAA
2241 AACAGAGGAG GAACAATGGA AGGAGCTGTT GGATAGTGAG ATATGGAGGT TAGGAAAGAG CGATGAGATT
2311 GTTCCGGCTT TTAGACTAAG CTACAATGAT CTPTCTGCCW CTTTGAAGCT RTTTRTTTGA TAYTGCTCTT
2381 TGTTTCCCAA GGACTATGAG TTTGACAAGG AGGAGTTGAT TCTATTGTGG ATGGCAGAAG GGTTTTGGCA
2451 CCAACCAACT AYAAACAAGT CAAAGCAAGC KTTGGGTCTT GAATATTTTR AAGAGTTRTT GTCAAGRTCR
2521 TTTTTCACAC ATGCTCCTAA TRRCAATCS TTGTTTGTA TGATGACCT AATGAATGAT TTGGCTACAT
2591 TTGTTGCTCG AGAATTTTTT TCAAGGTTAG ACATAGAGAT GAAGAAGGAA TTTAGGATGS AATCTTTGGA
2661 RAAGCACCCT CATATGTCAT TTGTATGTGA GRATTACATA GGTTCACAAA RGTTCGAGCT ATTTAGAGGA
2731 GCTAAAAATT TGAGAACATT TTTAGCATTG TCTGTTGGGG TGGTAGAAGA TTGGAAGATG TTTTACTTAT
2801 CAAACAAGGT CTGTAATGAC WTACTTCARG ATTTACCATT GTTAAGGGTC CTRAKTTTGA TTRRTCTTAY
2871 AATAASYRAG GTACCARAAK TCGTSGGTAG TATGAASCAC TTGCGGTATC TTAATCTTATC WGRAACTTWA
2941 ATCACACATT TACCGGAAWA TRCTTGCAAT CTTTATAATT TACARACCTT GATGTNTCT GGCTGTGAMT
3011 ATTTAGTTAA KTGCCCAAR ACCTTCTCAA ASCTTAAAAA TTTGCASCAT TTTGACATGA GGGTACTCC
3081 KAAKTTRAAK AACATGCCCT TARGGATTGG TGAATTTGAA ARTCTACAAA CTCTCTTMYG TAACATTGGC
3151 ATAGCAATAA CCGAGCTTAA GAACTTGCAM AAYCTCCATG GGAARITTTG TAITGGCGGG CTGGGAAAAA
3221 TGGAAAAATG HGTGGGATGC ACGTTAAGCG AACTTGTCTC A:AAAAAGGT TWAATGARTT ANAACTGGR
3291 WTKGGGGTGT ATRAAATTTAA TGTTTTCCGA AATGGGAACA CTGAAAAAAA NAAGGTCCTC AATGAATTGA
3361 ATGCCCTACA ATGGTATYCY AAMWAARRRY YYWTFARWAT TWMKAWRRK KGKTTYATRR TKPTMYRAAW
3431 WAGRGTKTYS KARGTAGGTT TCATCCAATC ACCCAAGTGG GAAAAATAGAT GATATTTTCA GGGCTTACTG
3501 ATGAGATGTT GAGAGGTATG ATAGGCTNVC TTGGGGCGGT AGAAGAAATA AGCATCCATT CTCTTAATGA
3571 AATAAGATAAT YTGTTGGAAAT CAGAAGCAGA GGCAAGTAAG GTTCTTATGA ATTTAAAGAA GTTGGATTTA
3641 GGTGAATGTT AAAATTTGGT GAGTTTAGGG GAGAAAAAGG AGGATAATCA TAATATTAAT AGTGGGAGCA
3711 GCCTAACATC TTTTAGGAGG TTGAATGTAT GGAGATGTAA CAGCTTGGAG CATTCGAGGT GTCCAGATAG
3781 CATGGAGAAAT TTGTATATGC ACATGTGTGA TTCAATNACA TCCGCTCTCT TCCCAACAGG AGGAGGACAG
3851 AAGATCAAGT CACTTACCAT CACTGATTGC AAGAAGCTTT CGGAAGAGGA GTTGGGAGGA CGAGAGAGGA
3921 CAAGAGTGGT TATAAATCA AAAATGCAGA TGCTTGAATC AGTAGATATA CTAATTTGCC CAARTCTGAA
3991 ATCTATCATG GAATGAGTT GCTTCATTCA CCTGAACAGA TTATATATAT CAAACTGTCC GAGTRTGGAG
4061 TCATTTCCTG ACCATGAGTT GCCAAATCTC ACCTCCTTAA CAGATCGAAG GAGAGGACAG CGATTTTCCT
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RLG1-E169
[Strand]

4131 ACGAACGGTT ACGATTGAC TGGCGTCGT TTT

SEQ ID NO: 70

Further Characterization of RG2 Family Members:

Further sequencing of cloned RG2 polynucleotide sequences, as discussed above, identified additional RG2 species, listed below. Additionally, further sequencing of the 5' sections of RG2 sequences listed above resulted in modified and/or new sequence information, also listed below. The AC15 sequences found in the 3' sections of RG2 family have not changed.

Listed below are: four full length species, RG2A, RG2B, RG2C and RG2S; two near complete, but with a gap in the largest intron, RG2D and RG2J; three nearly complete RG2 gene sequences, RG2K, RG2N, and RG2O. The deduced translation products (polypeptides) encoded by these RG2 species are listed below. The polynucleotide sequences do not contain any gaps (as with some of the polynucleotide sequences), because all of the gaps in the sequences are in introns, *i.e.*, there are no gaps in exon, or coding, sequences.

They include: an RG2A polynucleotide sequence (SEQ ID NO:87) and its deduced polypeptide sequence (SEQ ID NO:88); an RG2B polynucleotide sequence (SEQ ID NO:89) and its deduced polypeptide sequence (SEQ ID NO:90); an RG2C polynucleotide sequence (SEQ ID NO:91) and its deduced polypeptide sequence (SEQ ID NO:92); an RG2D polynucleotide sequence (SEQ ID NO:93) and (SEQ ID NO:94), and its deduced polypeptide sequence (SEQ ID NO:95); an RG2E polynucleotide sequence (SEQ ID NO:96) and its deduced polypeptide sequence (SEQ ID NO:97); an RG2F polynucleotide sequence (SEQ ID NO:98) and its deduced polypeptide sequence (SEQ ID NO:99); an RG2G polynucleotide sequence (SEQ ID NO:100) and its deduced polypeptide sequence (SEQ ID NO:101); an RG2H polynucleotide sequence (SEQ ID NO:102) and its deduced polypeptide sequence (SEQ ID NO:103); an RG2I polynucleotide sequence (SEQ ID NO:104) and its deduced polypeptide sequence (SEQ ID NO:105); an RG2J polynucleotide sequence (SEQ ID NO:106) and (SEQ ID NO:107), and its deduced polypeptide sequence (SEQ ID NO:108); an RG2K polynucleotide sequence (SEQ ID NO:109) and (SEQ ID NO:110), and its deduced polypeptide sequence (SEQ ID NO:111); an RG2L polynucleotide sequence (SEQ ID NO:112) and its deduced polypeptide sequence (SEQ ID NO:113); an RG2M polynucleotide sequence (SEQ ID NO:114) and its deduced polypeptide sequence (SEQ ID NO:115); an RG2N polynucleotide sequence (SEQ ID NO:116) and its deduced polypeptide sequence (SEQ ID NO:117); an RG2O

polynucleotide sequence (SEQ ID NO:118) and its deduced polypeptide sequence (SEQ ID NO:119); an RG2P polynucleotide sequence (SEQ ID NO:120) and its deduced polypeptide sequence (SEQ ID NO:121); an RG2Q polynucleotide sequence (SEQ ID NO:122) and its deduced polypeptide sequence (SEQ ID NO:123); RG2S polynucleotide
5 sequence (SEQ ID NO:124) and its deduced polypeptide sequence (SEQ ID NO:125); an RG2T polynucleotide sequence (SEQ ID NO:126) and its deduced polypeptide sequence (SEQ ID NO:127); an RG2U polynucleotide sequence (SEQ ID NO:128) and its deduced polypeptide sequence (SEQ ID NO:129); and RG2V polynucleotide sequence (SEQ ID NO:130) and its deduced polypeptide sequence (SEQ ID NO:131); and, an RG2W
10 polynucleotide sequence (SEQ ID NO:132) and its deduced polypeptide sequence (SEQ ID NO:133).

Characterization of New RG Family Groups and RG Species:

Further BAC insert characterization and sequencing, as discussed above,
15 identified new RG polynucleotide sequences. The new sequences were characterized as belonging to new RG families; designated RG5 and RG7. These RG polynucleotides sequences, and their predicted translation products (the polypeptides which are encoded by these sequences) are summarized and listed below.

Identified and listed below is an RG5 family member, designated as the RG5
20 polynucleotide sequence set forth in SEQ ID NO:134, and its deduced polypeptide sequence (SEQ ID NO:135). This sequence contains an NBS region sequence.

Also identified and listed below is an RG7 family member, designated as the
RG7 polynucleotide sequence set forth in SEQ ID NO:136. No deduced polypeptide
sequence is given for the new RG7 family member as this sequence appears to be a
25 pseudogene.

RG2A polynucleotide sequence (SEQ ID NO:87)

AAAGTTCATATCCAAGCTTGCCCTCCAACCTAGCTCCTTCAATGGCACC
TCCTTCTCTTCAAAAGCACACAAGAACACTTTCAAGCTCAACCACACTCA
30 CACAAGCTCTAGAACGAGGGTTAGGGCACATTTAGGGTTTTGCTCTCTGG
AAATGGTGTCTAAAAGTGAGGCCATAATGTTTCCTTATATAAGGCTCACTC
CCACAATTAGGCTTTCAATCTGAACGTANTACGCCAGTGTAACACTATGG
TACGCCCAACGTAACGCTAGTCTCCGCGTCAANAATACTCATGAGTA

CGCGCAACGTA CTTTCCCTTACGCCCAGCGTACTCAAAAGCCAAACATTC
TTTTCAAGGACTAATTTTGACAACCTTGAGGAAAGAAAAGGATCAAAGANA
TATACTTGAATTCCGGGATGTTACAATGAAGTTGANACCTTGGCTAAAAA
ATTAAATTGGTTGTGGAAGCCGTTGGCTGAGCAAGCAACAAGGGTAAAT
5 TCGTAATCTACAAATGGTGTTATTTTCTATTTCTTCTTATTATTTTACTT
GATTTACGGGTAGTTTTTTTTTCTTACAAAAAATATTAAAGTTGATAAAG
TATAGCCACTAAAATTGACTTTTTTCCAAAACATAATGTCAAATGGTGCGT
ATATGTATCATGTTGTATTANATAATGAATATGATGATNCTGTTCTATTT
AANCCGAAAAAATTATCTAATGATTTTATATTGGAACAAAGTTGTGAT
10 TTTTNGCATAATATAATCAAATCCNCTTTTGTNTGGGAGGTGGATAAATG
TGGTAAATTTANAACAAGTGTTTTNACNTTGAAGGGTNTGGAAAGGTTGA
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ATNATGTTTAAGGATNATTGTATTAAATTGTTGATATATAGTAAGCATAA
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15 CATTTTTGATATANATTAGGAATGAAAATGAGCAACCCTAACATACTTAT
CTTTGGTAGTTTGGTTATTATATTTTTATTANAATATAGAANCATCCCTT
TATTTTAAACCCATATTGTGGACGGACTTGAATAAATGGGAAAAATGTAC
CTTGCTATTTAGCACAAAAAAATTATAAAAAATGTACATTGCTATTTAGCA
CAAACAAAAAATACTTATCCTTTTTGCATTAGGTCACAAAGAAATA
20 TAAATGGGAAATGTGTTGCTATTTAATGCACTAAAAGAACTATTTTGC
CTTTATTAAACCGGGTAAACCAATAGAAAAATGGAAGTACATTGTCATTT
AGC.ATGAAAAAAATAACTTTCCATTTTTTGCATCCGGTCACAATAATAG
AAAAATGAAAGTACGTTGCTATTTAGCGAACTAACTTCCTTTTTTCTTT
TTGGCATCGTATCATAAAATATAGACTAAAATACGTTAGTTTTACATTTT
25 TAATACATTGAAATGTCTAATCCACATGTTATTCTATAAAAAGGGAAATG
TAATTTACTTATTCTTTGATTCTTTGGCTTCTTTTTAGTACCCAAAACAT
CCCTCTATCCATCTATTCCAATAAATAATGAAAATATATTCCTTCCA
TTGTAGGGATGTTATAAATTTTGTAAATTGTTTTTATGCAAAAAAGTGTTT
TTTGTTAACTAGATTAACGAGATTCATTTTTCAGCATTTTAGGAGAAGTT
30 CATCCATCTTTTGGATATGAAGTGCAAGCCAAGTTCTTTAACATGGAATA
TGAGGTCCCTATATGCTCAAAAAATAGCAAATGAGAAATTTTTTAAATTG
GATCCCCATAAAAGAAAATTTGTAAATGGTTGTTTAAATATTGGTCAATG
TGTCCACCGGATGAGCATAATACTAGTTTATAAGGGGTAAAGGTGGGTTT
GGTGGGCCCATTTATCTTTATTATTTCTAAAAGTCAGAATTAAGTAAAAA
35 AAATTATAAGATAAATAACCATAAGGATAAAAAATCATTTTATTTGGACCA
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GTCATCCTGGCGCATATATGTGTCACTGTCTTCAAAGGTCTTCAGACCTC
40 ATTTTAACCAAAAAAAGACCACCGGTTTTTTTTTTTTTTTTNTTC
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5 GGTAAAGTGTATGTGTGTACGCGCGCACCTGAAAGGTTTGAANGTAACTT
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GTGAAGGAATCAGCTGGAGGTTGGGGAATCGAGCTTCCACTATTAAGGTA
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TGTTGAATGAAAAAAGCATGCTCAAAAAACCAGTGTAAGGCACGGTATAT
10 GACATATTTATAGTTACTGATAACAAATTATGATAATTTTGGGTTTACGT
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TATGTATTTGGGGAATCACATTAGCAACGGGATTGTACTAGTAATTCGAA
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15 GTG.AACATAGACCTTTTCCATTTACTAATGCCTAATTATTAGTTTCTAAT
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ATTTACCTTTAAATATTAACAAAAATGACCATAAATAAAAAAATTAG
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20 AGTTTCATACAAGAAAATTTCAAGAAGAAAGCAAAGGTCCAGGTATTCTC
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25 GAGATAGAGATATGTTAAAACCTGGCTAGAAAATTGTTTTAATTTGAAATT
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35 AAACAACAGGTAAACCAGATCTTTGTTGCTTNNATAATTCTTAAACNACA
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40 ATTCCTTCATGATGATGTCAATCTTCTAATACCCCATTCATTGTTTGGTTG
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RG2A deduced polypeptide sequence (SEQ ID NO:88)

MDVVNAILKPVVETLMVPVKKHIGYLISCRQYMREMGIKMRGLNATRLGVEEHVN
RNISNQLEVPAQVRGWFEVVGKINAKVENFSPDVGSCFNLKVRHGVGKRASKIIEDI
DSVMREHSIIWNDHSIPLGRIDSTKASTSIPSTDHHDEFQSREQTFTEALNALDPNHK
5 SHMIALWGMGGVGKTTMMHRLKKVVKEKKMFNFIEAVVGEKTDPIAIQSAVADY
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RG2B polynucleotide sequence (SEQ ID NO:89)

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40

RG2B deduced polypeptide sequence (SEQ ID NO:90)

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15 LNGLFLKTEVLF LSVHGMNDLEDVEVKSTHPTQSSSFCNLKVLISKCVELRYLFKL
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RISECDGIKEVVSNRDEDEEMTTFTSTHTTTTFLPSLDSLTSFLENLKICGGGGAK
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25 GQMOKL

RG2C polynucleotide sequence (SEQ ID NO:91)

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5 CGGTTATAGGGGAAATATCGGACCCCATGCTATTCAGCAAGTTGTAGCA
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RG2C deduced polypeptide sequence (SEQ ID NO:92)

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RG2D polynucleotide sequence (SEQ ID NO:93) and (SEQ ID NO:94)

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Sequence gap

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RG2D deduced polypeptide sequence (SEQ ID NO:95)

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20 DFKQDSD.CEVNIK

RG2E polynucleotide sequence (SEQ ID NO:96)

TGGGAAGACACAATGATGCAAAGGTTGAAGAAGGTTGCTAAAGAAAATAGAAT
GTTCAATTATATGGTTGAGGCAGTTATAGGGGAAAAGACAGACCCACTTGCTAT
25 TCAACAAGCTGTAGCGGATTACCTTTGTATAGAGTTAAAAGAAAGCACTAAACC
AGCAAGAGCTGATAAGCTTCGTGAATGGTTTAAGGCCAACTCTGGAGAAGGTA
AGAATAAGTTCCTTGTAATATTTGATGATGTTTGGCAGTCCGTTGATCTGGAAG
ACATTGGTTTAAGTCATTTTCCAAATCAAGGTGTCGACTTCAAGGTCTTGTTGA
CTTCACGAGACGAACATGTTTGCACAGTAATGGGGGTTGAAGCTAATTCAATTC
30 TTAATGTGGGACTTCTAGTAGAAGCAGAAGCACAAAGTTTGTTCAGCAATTTG
TAGAAACTTTTGAGCCCGAGCTCCATAAGATAGGAGAAGATATCGTAAGGAAG
TGTTGTGGTTTACCTATTGCCATTA AAACCATGGCATGTACTCTAAGAAATAAA
AGAAAGGATGCATGGAAGGATGCACTTTTGCATTTAGAGTACCATGACATTAGC
AGTGTTGCGCCCAAAGTCTTTGAAACGAGCTACCATAATCTCCACAACAAGGAG
35 ACTAAATCTGTGTTTTTGATGTGTGGTTTTTTTCTGAAGACTTCAATATTCCAA
TCGAGGAGTTGATGAGGTATGGATGGGGCTTAAAGATATTTGATAGAGTTTATA
CTATTAGACAAGCAAGAATCAGGCTCAACACCTGCATTGAGCGACTGGTGCAG
ACAAATTTGTTAATAGAAAGTGATGATGGTGTGCACGTCAAGATGCATGATCTG
GTCCGTGCTTTCGTTTTGGTTATGTTTTCTGAAGTTGAACATGCTTCAATTATCA
40 ACCATGGTAATATGCTTGGATGGCCTGAAAATTATATGACCAACTCTTGCAAAA
CAATTTTCATTAAACATGCAAGAGTATGTCTGAATTTCCGGGAGATCTCAAGTTTC
CAAACCTAACGATTTTGAAACTCATGCATGGAGATAAGTTGCTAAGATATCCTC

AAGACTTTTATGAAGGAATGGAAAAGCTCTGGGTTATATCATATGATGAAATGA
AGTATCCATTGCTTCCCTCGTTACCTCAATGCTCCATCAACCTTCGAGTGCTTCA
CCTCCATCGATGCTCATTAATGATGTTTGATTGCTCTTGTATTGGAAATATGTTG
AATCTGGAAGTGCTTAGCTTTGTAAATCTGGCATTGAATGGTTACCTTCCACA
5 ATAGGAAATTTAAAGAAGCTAAGGTTACTTGATCTGAGAGATTGTTATGGTCTT
CGTATAGAAAAAGGTGTCTTGAAAAATTTGGTGAAAATTGGAGGAATTTATATT
GGTAGAGCAGATATTTTATAGAT

RG2E deduced polypeptide sequence (SEQ ID NO:97)

10 WEDTMMQRLKKVAKENRMFNYMVEAVIGEKT DPLAIQQA VADYLCIELKESTKP
ARADKLREWFKANS GEGKNKFLVIFDDVWQSV DLEDIGLSHF PNQGVDFKVLLTS
RDEHVCTVMGVEANSILNVGLLVEAEAQSLFQQFVETFEPELHKIGEDIVRKCCGL
PIAKTMACTLRNKRKDAWKDALLHLEYHDISSVAPKVFETSYHNLHNKETKSVFL
MCGFFPEDFNIPIEELMRYGWGLKIFDRVYTIRQARIRLNTCIERLVQTNLLIESDDG
15 VHVKMHDLVRAFVLVMFSEVEHASIINHGNMLGWPENYMTNSCKTISLTCKSMSE
FPGDLKFPNL TILKLMHGDKLLRYPQDFYEGMEKLWVISYDEMKYPLLPSLPQCSI
NLRVLHLHRCSLMMFDCSCIGNMLNLEVL SFVKSGIEWLPSTIGNLKKLRLLDLRD
CYGLRIEKGVLKNLVKIGGIYIGRADIL.

RG2F polynucleotide sequence (SEQ ID NO:98)

CTGTGGAAGACACAATGATGCAAAGGCTGAAAAAGGTTGTGCATGAAAAGAAA
ATGTTTAACTTTATTGTTGAAGCAGTTATAGGGGAAAAGACAGACCCCGTTGCC
ATTCAGGATGCTATAGCAGATTACCTAGGTGTAGAGCTCAATGAAAAATCTAAG
CAAGCAAGAGCTGATAAGCTCCGTCAAGGATTCAAGGACAAATCAGATGGAGG
25 CAAAATAAGTTCTTTGTAATACTTGACGATGTTTGGCAGTCTGTTGATCTGGA
AGATATTGGTTTAAAGTCCTTTTCCAAATCAAGGCGTCGACTTCAAGGTCTTGTT
GACATCACGAGACAGACATGTTTGCACAGTGATGGGGGTTGAAGCCAAATTAA
TTCTAAACGTGGGACTTCTAATTGAAGCTGAAGCACAAAGTTTGTTCACCAAT
TTGTTGTCACTTCTGAGCCCGAGCTCCATAAGATAGGAGAAGATATTGTAAAGA
30 AGTGTTCGGTCTGCCAATTGCCATCAAAACCATGGCATGTACTCTACGACATA
AAAGAAAGGATGCATGGAAGGATGCACTTTCACGTTTAGAGCACCATGACATT
CAAAGTGTTGTGCCTAAAGTATTTGAAACGAGCTACAACAATCTCAAAGACAA
GGAGACTAAATCCGTATTTTTGATGTGTGGTTTGTTCCTGAAGACTTGGATAT
ACCTATCGAGGAGTTGATGAGGTATGGATGGGGCTTAAGATTATTTGATAGAGT
35 TAATACTATTACACAAGCAAGAAACAGGCTCAACACCTGCATTGAGCGACTGG
TGCACACAAATTTGTTAATTGAAAGTGTTGATGGTGTGCATGTCAAGATGCATG
ATCTGGTTCGTGCTTTTGT TTTTGGGAATGTTTTCTGAAGTGGAGCATGCTTCAAT
TGTCAACCATGGTAATATGCCCCGAGTGGACTGAAAATGATATGACTGACTCTTG
CAAACAAATTTCAATTAACATGCAAGAGTATGTTGGAGTTTCTGGAGACCTCAA
40 GTTTCCAAACCTAAAGATTTTGAAACTTATGCATGGAGGTAAGTCACTAAGGTA
TCCTCAAGACTTTTATCAAGGAATGGAAAAGCTGGAGGTTATATCATACGATGA
AATGAAGTATCCATTGCTTCCCTCGTTGCCTCAATGTTCCACCATCCTTCGAGTG

CTTCATCTCCATGAATGTTCAATTAAGGATGTTTGATTGCTCTTCAATCGGTAATC.
TTTTCAACATGGAAGTGCTCAGCTTTGCTAATTCTAGCATTGAATTGTTACCTTC
CGTAATTGGAATTTGAAGAAGTTGCGGCTGCTAGATTTGACAACTGTTATGG
TGTTTCGTATAGAAAAGGATGTCTTGAAAAATTTGGTGAAACTTGAAGAGCTTTA
5 TATTAGGAATGGTCTACCAGTTTACAGAGGAT

RG2F deduced polypeptide sequence (SEQ ID NO:99)

VEDTMMQRLKKVVHEKKMFNFIVEAVIGEKTDPVAIQDAIADYLGVELNEKSKQA
RADKLRRQGFKDKSDGGKKNKFFVILDDVWQSVDDLEDIGLSPFPNQGVDFKVLLTSRD
10 RHVCTVMGVEAKLILNVGLLIEAEAQSLFHQFVVTSEPELHKIGEDIVKKCFGLPIAI
KTMACTLRHKRKDAWKDALSRLEHDIQSVVPKVFETSYNNLKDKETKSVFLMCG
LFPEDLDIPIEELMRYGWGLRLFDRVNTITQARNRLNTCIERLVHTNLLIESVDGVH
VKMHDLVRAFLVGMFSEVEHASIVNHGNMPEWTENDMTDSCKQISLTCKSMLEFP
GDLKFPNLKILKLMHGGKSLRYPQDFYQGMKLEVISYDEMKYPLLPQLPQCSTILR
15 VLHLHECSLRMFDCCSIGNLNFMEVLSFANSSIPELLPSVIGNLKKLRLDLTNCYGV
RIEKDVLKNLVKLEELYIRNGLPVYRG

RG2G polynucleotide sequence (SEQ ID NO:100)

GAAGACACGATGATGAAGAACTGAAGGAGGTCGTGGGACAAAAGAAATCATT
20 AATATTATTATTCAAGTGGTCATAGGAGAGAAGACAAACCCTATTGCAATTGAG
CAAGCTGTAGCAGATTACCTCTCTATAGAGCTGAAAGAAAACACTAAAGAAGC
AAGAGCTGATAAGCTTCGTAAACGGTTTGAAGCCGATGGAGGAAAGAATAAGT
TCCTTGTAATACTTGACGATGTATGGCAGTTTGTTCGATCTTGAAGATATTGGTTT
AAGTCCTCTGCCAAATAAAGGTGTCAACTTCAAGGTCTTGTTGACGTCAAGAGA
25 TTCACATGTTTGCACTCTGATGGGAGCTGAAGCAAATTCAATTCTTAATATAAA
AGTTTTTAAAGATGTAGAAGGACAAAGTTTGTTCGCCAGTTTGCTAAAAATGC
GGGTGATGATGACCTGGATCCTGCTTCAATGGGATAGCAGATAGTATTGCAAG
TAGATGTCAAGGTTTGCCCATTCATCAAAACCATTCCTTAAGTCTTAAAGG
TAGAAGCAAGTCTGCATGGGACGTTGCACTTTCTCGTCTGGAGAATCATAAGAT
30 TGGTAGTGAAGAAGTTGTGCGTGAAGTTTTTAAATTAGCTACGACAATCTCCA
AGATGAGGTTACTAAATCTATTTTTTTACTTTGTGCTTTATTCCTGAAGATTTT
GATATTCCTACTGAGGAGTTGGTGAGGTATGGGTGGGGCTTGAAATTATTTATA
GAAGCAAAAACATAAGAGAAGCAAGAAACAGGCTCAACACCTGCACTGAGCG
GCTTAGGGAGACAAATTTGTTATTTGGAAGTGATGACATTGGATGTGTCAAGAT
35 GCACGATGTGGTGCGTGATTTTGTGTTTGCATATATTCTCAGAAGTCCAACACGC
TTCAATTGTCAACCATGGTAACGTGTCAGAGTGGCTAGAGGAAAATCATAGCAT
CTACTCTTGTAAGAATTTCAATTAACATGCAAGGGTATGTCTCAGTTTCCCAA
AGACCTCAAATTTCCAAACCTTTCAATTTTGAACTTATGCATGGAGATAAGTC
ACTGAGCTTTTCTGAAAACCTTTTATGGAAAAGATGGAAAAGGTTCAAGTAATATC
40 ATATGATAAATTGATGTATCCATTGCTTCCCTCATCACTTGAATGCTCCACCAA
CGTTCGAGTGCTTCATCTTCATTACTGTTCAATTAAGGATGTTTGATTGCTCTTCA
ATTGGTAATCTTCTCAACATGGAAGTGCTCAGCTTTGCTAATTCTAACATTGAA

TGGTTACCATCTACAATTGGAAATTTGAAGAAGCTAAGGCTACTAGATTTGACA
AATTGTAAAGGTCTTCGTATAGATAATGGTGTCTTAAAAAATTTGGTCAAACCTT
GAAGAGCTTTATATGGGTGTTAATCGTCCGTATGGACAGGCCGTTAGCTTGACA
GATGAAAA

5

RG2G deduced polypeptide sequence (SEQ ID NO:101)

RHDDEELKEVVGQKKSFNIIQVVIGEKTNPQIAIQAVADYLSIELKENTKEARADKL
RKRFEADGGKNKFLVILDDVWQFVDLEDIGLSPLPNKGVNFKVLLTSRDSHVCTL
MGAEANSILNKKVLDVEGQSLFRQFAKNAGDDDLDPAFNGIADSIASRCQGLPIAI
10 KTIALSLKGRSKSAWDVALSRLENHKIGSEEVVREVFVKISYDNLQDEVTKSIFLLCAL
FPEDFDIPTTEELVRYGWGLKLFIEAKTIREARNRLNTCTERLRETNLLFGSDDIGCVK
MHDVVRDFVLHIFSEVQHASIVNHGNVSEWLEENHSIYCKRISLTCKGMSQFPKDL
KFPNLSILKLMHGDKSLSPENFYGKMEKVQVISYDKLMYPLLPSSLECSTNVRVLH
LHYCSLRMFDCSSIGNLLNMEVLSFANSNIEWLPSTIGNLKKLRLDLTNCKGLRID
15 NGVLKNLVKLEELYMGVNRPYQAVSLTDE

RG2H polynucleotide sequence (SEQ ID NO:102)

TGAAGGAGGTTGTGGAACGAAAGAAAATGTTCAGTATTATTGTTCAAGTG
GTCATAGGAGAGAAGACAAACCTATTGCTATTCAGCAAGCTGTAGCAGA
20 TTACCTCTCTATAGAGCTGAAAGAAAACACTAAAGAAGCAAGAGCTGATA
AGCTTCGTAAATGGTTCGAGGCCGATGGAGGAAAGAATAAGTTCCTTGTA
ATACTTGACGATGTATGGCAGTTTGTGCGATCTTGAAGATATTGGTTTAA
TCCTCTGCCAAATAAAGGTGTCAACTTCAAGGTCTTGTTGACGTCAAGAG
ATTCACATGTTTGCCTCTGATGGGAGCCGAAGCCAATTCAATTCTCAAT
25 ATAAAAGTTTTTAACAGCTGTAGAAGGACAAAGTTTGTTCGCCAGTTTGC
TAAAAATGCGGGTGATGATGACCTGGATCCTGCTTCAATAGGATAGCAG
ATAGTATTGCAAGTAGATGTCAAGGTTTGCCCATTGCCATCAAAACCATT
GCCTTAAGTCTTAAAGGTAGAAGCAAGCCTGCGTGGGACCATGCGCTTTC
TCGTTTGGAGAACCATAAGATTGGTAGTGAAGAAGTTGTGCGTGAAGTTT
30 TTAAAATTAGCTATGACAATCTCCAAGATGAGATTACTAAATCTATTTTT
TTACTTTGTGCTTTATTTCTGAAGATTTTGATATTCCTACTGAGGAGTT
GATGAGGTATGGATGGGGCTTGAAATTATTTATAGAAGCAAAAACCTATAA
GAGAAGCAAGAAACAGGCTCAACACCTGCACTGAGCGGCTTAGGGAGACA
AATTTGTTATTTGGAAGCGATGACATTGGATGCGTCAAGATGCACGATGT
35 GGTGCGTGATTTTGTGTTTGCATATATTCTCAGAAGTCCAGCACGCTTCAA
TTGTCAACCATGGTAACGTGTGAGAGTGGCTAGAGGAAAATCATAGCATC
TACTCTTGTAAGAATTTTCAATTAACATGCAAGGGTATGTCTGAGTTTCC
CAAAGACCTCAAATTTCAAACCTTTCAATTTTGAACTTATGCATGGAG
ATAAGTCGCTGAGCTTTCCTGAAAACCTTTTATGGAAAGATGGAAAAGGTT
40 CAGGTAATATCATATGATAAATTGATGTATCCATTGCTTCCCTCATCACT
TGAATGCTCCACTAACGTTTCGAGTGCTTCATCTCCATTATTGTTCAATTA
GGATGTTTGATTGCTCTTCAATTGGTAATCTTCTCAACATGGAAGTGCTC

AGCTTTGCTAATTCTAACATTGAATGGTTACCATCTACAATTGGAAATTT
GAAGAAGCTAAGGCTACTAGATTTGACAAATTGTAAAGGTCTTCGTATAG
ATAATGGTGTCTTAAAAAATTTGGTCAAACCTGAAGAGCTTTATATGGGT
GTTAATCATCCGTATGGAC

5

RG2H deduced polypeptide sequence (SEQ ID NO:103)

KEVVERKKMFSIIVQVVIGEKTNPQIAQAVADYLSIELKENTKEARADKLRKWFEA
DGGKNKFLVILDDVWQFVDLEDIGLSPLPNKGVNFKVLLTSRDSHVCTLMGAEAN
SILNIKVLTAVEGQSLFRQFAKNAGDDDLDPAFNRIADSIASRCQGLPIAKTIALSLK
10 GRSKPAWDHALSRLENHKIGSEEVVREVFKISYDNLQDEITKSIFLLCALFPEDFDIP
TEELMRYGWGLKLFIEAKTIREARNRLNTCTERLRETNLLFGSDDIGCVKMHDVVR
DFVLHIFSEVQHASIVNHGNVSEWLEENHSIYSCKRISLTCKGMSEFPKDLKFPNLSI
LKL.MHGDKSLSPFENFYGKMEKVQVISYDKLMYPLLPSSLECSTNVRVLHLHYCSL
RMFDCSSIGNLLNMEVLSFANSNIEWLPSTIGNLKKLRLLDLTNCKGLRIDNGVLKN
15 LVKLEELYMGVNHPYG

RG2I polynucleotide sequence (SEQ ID NO:104)

AAGAAGAGCTGAAGGAGGTTGTGGAACAAAAGAAAACGTTCAATATTATT
GTTCAAGTGGTCATAGGAGAGAAGACAAACCCTATTGCTATTCAGCAAGC
20 TGTAGCAGATTCCTCTCTATAGAGCTGAAAGAAAACACTAAAGAAGCAA
GAGCTGATAAGCTTCGTAAATGGTTCGAGGCTGATGGAGGAAAGAATAAG
TTCCTCGTNATACTTGACGATGTATGGCNGTTTGTGATCTTGAAGATAT
TGGTTTAAGTCCTCATCCAAATAAAGGTGTCANCTTCAAGGTCTTGTTGA
CGTCAAGAGATTACATGTTTGCACCTCTGATGGGAGCTGAAGCCAATTCA
25 ATTCTCAATATAAAAGTTTTTAAAGATGTAGAAGGAAAAAGTTTGTCCG
CCAGTTTGCTAAAAATGCGGGTGATGATGACCTGGATCCTGCTTTCATTG
GGATAGCAGATAGTATTGCAAGTAGATGTCAAGGTTTGCCCATTTGCCATC
AAAACCATTTGCCTTAAGTCTTAAAGGTAGAAGCAAGTCTGCATGGGACGT
TGC.ACTTTCTCGTCTGGAGAATCATAAGATTGGTAGTGAAGAAGTTGTGC
30 GTGAAGTTTTTAAATTAGCTATGACAATCTCCAAGATGAGGTTACTAAA
TCT.ATTTTTTTACTTTGTGCTTTATTTCTGAAGATTTTGATATTCCTAC
TGAGGAGTTGGTGAGGTATGGGTGGGGCTTGAAATTATTTATAGAAGCAA
AACTATAAGAGAAGCAAGAAACAGGCTCAACACCTGCACTGAGCGGCTT
AGGGAGACAAATTTGTTATTTGGAAGTGATGACATTGGATGCGTCAAGAT
35 GCACGATGTGGTGCGTGATTTTGTGTTTGCATATATTCTCAGAAGTCCAGC
ACGCTTCAATTGTCAACCATGGTAATGTGTCAGAGTGGCTAGAGGAAAAT
CATAGCATCTACTCTTGTAAGAAGATTTTCAATTAACATGCAAGGGTATGTC
TGAGTTTCCCAAAGACCTCAAATTTCCAAACCTTTCAATTTTGAACTTA
TGC.ATGGAGATAAGTCGCTGAGCTTTCCTGAAAACCTTTATGGAAAGATG
40 GAAAAGGTTCAAGTAATATCATATGATAAATTGATGTATCCATTGCTTCC
CTC.ATCACTTGAATGCTCCACCAACCTTCGAGTGCTTCATCTCCATGAAT
GTTCAATTAAGGATGTTTGATTGCTCTTCAATTGGTAATCTTCTCAACATG

GAAGTGCTCAGCTTTGCTAATTCTGGCATTGAATGGTTACCATCTACAAT
TGGAAATTTGAAGAAGCTAAGGCTACTGGATCTGACAGATTGTGGAGGTC
TTCATATAGATAATGGCGTCTTAAAAAATTTGGTCAAACCTGAAGAGCTT
TATATGGGTGCTAATCGTCTGTTTGGAAAGTGCCAT

5

RG2I deduced polypeptide sequence (SEQ ID NO:105)

EELKEVVEQKKTFNIIQVQVIGEKTNPIAIQQA VADSLSELKENTKEARADKLRKWF
EADGGKNKFLVILDDVW?FVDLEDIGLSPHPNKG V?FKVLLTSRDSHVCTLMGAEA
NSILNIKVLKDVEGKSLFRQFAKNAGDDDLDPAFIGIADSIASRCQGLPIAKTIALSL
10 KGRSKSAWDVALSRLENHKIGSEEVVREVFKISYDNLQDEVTKSIFLLCALFPEDFDI
PTEELVRYGWGLKLFIEAKTIREARNRLNTCTERLRETNLLFGSDDIGCVKMHDVV
RDFVLHIFSEVQHASIVNHGNVSEWLEENHSIYSCKRISLTCKGMSEFPKDLKFPNLS
ILKLMHGDKSLSFPENFYGKMEKVQVISYDKLMYPLLPSLE CSTNLRVLHLHECSL
RMFDCSSIGNLLNMEVLSFANS GIEWLPSTIGNLKKLRLLDLTDCGGLHIDNGVLKN
15 LVKLEELYMGANRLFGKCH

RG2J polynucleotide sequence (SEQ ID NO:106) and (SEQ ID NO:107)

ATGTCCGACCCAACAGGGATTGTTGGTGCCATTATTAACCCAATTGCTCA
AACGGCCTTGGTTCCCCTTACAGACCATGTAGGCTACATGATTTCTCTGCA
20 GAA AATATGTGAGGGACATGCAAATGAAAATGACAGAGTTAAATACCTCA
AGAATCAGTGCAGAGGAACACATTAGCCGGAACACAAGAAATCATCTTCA
GATTCATCTCAAATTAAGGATTGGTTGGACCAAGTAGAAGGGATCAGAG
CGAATGTTGCAAACCTTTCCAATTGATGTCATCAGTTGTTGTAGTCTCAGG
ATCAGGCACAAGCTTGGACAGAAAGCCTTCAAGATAACTGAGCAGATCGA
25 AAGTCTAACGAGACAAAATTCGCTGATTATCTGGACTGATGAACCTGTTC
CCCTGGGAAGAGTTGGTTCCATGATTGCATCCACCTCTGCAGCATCAAGT
GATCATCATGATGTCTTCCCTTCAAGAGAGCAAATTTTTAGGAAAGCACT
AGAAGCACTTGAACCCGTCCAAAAATCCACATAATAGCCTTATGGGGGA
TGGGCGGAGTGGGGAAGACCACGATGATGAAGAAGCTGAAAGAGGTCGTG
30 GAACAAAAGAAAACGTGCAATATTATTGTTCAAGTGGTCATAGGAGAGAA
GACAAACCCTATTGCTATCCAGCAAGCTGTAGCAGATTACCTCTCTATAG
AGCTGAAAGAAAACACTAAAGAAGCAAGAGCTGATAAGCTTCGTAAACGG
TTCGAAGCCGATGGAGGAAAGAATAAGTTCCTTGTAATACTTGACGATGT
ATGGCAGTTTTTTCGATCTTGAAGATATTGGTTTAAGTCCTCTGCCAAATA
35 AAGGTGTCAACTCAAGGTCTTGTTGACGTCAAGAGATTCACATGTTTGC
ACTCTGATGGGAGCTGAAGCCAATTCTATTCTCAATATAAAAGTTTTAAA
AGATGTAGAAGGAAAAAGTTTTGTTCCGCCAGTTTGCTAAAAATGCGGGTG
ATGATGACCTGGATCCTGCTTTCATTGGGATAGCAGATAGTATTGCAAGT
AGATGTCAAGGTTTGCCCATGCAAAACCATTCCTTAAGTCTTAA
40 AGGTAGAAGCAAGTCTGCATGGGACGTCGCACTTTCTCGTCTGGAGAATC
ATAAGATTGGTAGTGAAGAAGTTGTGCGTGAAGTTTTTAAAATTAGCTAT
GACAATCTCCAAGATGAGGTTACTAAATCTATTTTTTTACTCTGTGCTTT

ATTTCTGAAGATTTTGATATTCCTATTGAGGAGTTGGTGAGGTATGGGT
GGGGCTTGAAATTATTTATAGAAGCAAAAACATAAGAGAAGCAAGAAAC
AGGCTCAACAACCTGCACTGAGCGGCTTAGGGAGACAAATTTGTTATTTGG
AAGTCATGACTTTGGGTGCGTCAAGATGCACGATGTGGTGCGTGATTTTG
5 TTTTGCATATGTTTTCAGAAGTCAAGCATGCTTCAATTGTCAACCATGGT
AACATGTCAGAGTGGCCAGAGAAAAATGATACCAGCAACTCTTGTA AAAAG
AATTTCAATTAACATGCAAGGGTATGTCTAAGTTTCCTAAAGACATCAACT
ATCCAAACCTTTTGATTTTGAACTTATGCATGGAGATAAGTCGCTGTGC
TTTCTGAAAACCTTTATGGAAAGATGGAAAAGGTTTCAGGTAATATCATA
10 TGATAAATTGATGTATCCATTGCTTCCCTCATCACTTGAATGCTCCACTA
ACGTTTCGAGTGCTTCATCTCCATTATTGTTTCAATTAAGGATGTTTGATTGC
TCTTCAATTGGTAATCTTCTCAACATGGAAGTGCTCAGCTTTGCTAATTC
TAACATTGAATGGTTACCATCTACAATTGGAAATTTGAAGAAGCTAAGGC
TACTAGATTTGACAAATTGTAAAGGCTTTCGTATAGATAATGGTGTCTTA
15 AAAAATTTGGTCAAACCTTGAAGAGCTTTATATGGGTGTTAATCGTCCGTA
TGGACAGGCCGTTAGCTTGACAGATGAAAACCTGCAATGAAATGGTAGAAG
GTTCCAAAAAACCTTCTTGCACTAGAATATGAGTTGTTTAAATACAATGCT
CAAGTGAAGAATATATCCTTCGAGAATCTTAAACGATTCAAGATCTCAGT
GGGATGTTCTTTACATGGATCTTTCAGTAAAAGCAGGCACTCATACGAAA
20 ACACGTTGAAGTTGGCCATTGACAAAGGCGAACTATTGGAATCCCGAATG
AACGGGTTGTTTGAGAAAACGGAGGTTCTTTGTTTAAAGTGTGGGGGATAT
GTATCATCTTTCAGATGTTAAGGTGAAGTCCTCTTCGTTCTACAATTTAA
GAGTCCTTGTCGTTTCAGAGTGTGCAGAGTTGAAACACCTCTTCACACTT
GGTGTGCAAATACTTTGTCAAAGCTTGAGCATCTTAAAGTCTACAAATG
25 CGATAATATGGAAGAACTCATACATACCGGGGGTAGTGAAGGAGATACAA
TTACATTCCCCAAGCTGAAGCTTTTATATTTGCATGGGCTGCCAAACCTA
TTGGGTTTGTGTCTTAATGTCAACGCAATTGAGCTACCAAACTTGTGCA
AATGAAGCTTTACAGCATTCCGGGTTTCACAAGCATTATCCGCGGAACA
AGTTGGAAGCATCTAGTTTGTGAAAGAAGAGGTACATATACATATAGTT
30 TATGTTAATACATTTTAAACAATCTTTTCAACTAAAAGTTTCAGAATATA
TCTGTATTTTGATTGTATGATGTGTTAGTGTGTTGGATGTGGCTATTAAAG
GATAATTATTTGGCAGGTTGTGATTCCTAAGTTGGATATACTTGAAATTC
ATGACATGGAGAATTTAAAGGAAATATGGCCTAGTGAGCTTAGTAGAGGT
GAGAAAGTTAAGTTGAGAAAGATTAAAGTGAGAAATTGTGATAAACTTGT
35 GAATCTATTTCCACACAATCCCATGTCTCTGCTGCATCATCTTGAAGAGC
TTATAGTCGAGAAATGTGGTTCCATTGAAGAGTTGTTCAACATCGACTTG
GATTGTGCCAGTGTAATTGGAGAAGAAGACAACAACAGCAGCTTAAGAAA
CATCAATGTGGAGAATTCAATGAAGCTAAGAGAGGTGTGGAGGATAAAAAG
GTGCAGATAACTCTCGTCCCCTCTTTCGTGGCTTTCAAGTTGTTGAAAAG
40 ATAATCATTACGAGATGTAAGAGGTTTACAAATGTATTCACACCTATCAC
CACAAATTTTGATCTGGGGGCACTTTTGGAGATTTTCAGTTGATTGTAGAG
GAAATGATGAATCAGACCAAAGTAACCAAGAGCAAGAGCAGGTATGGATT
TCAATTTTACTCTTTTACTTAATTAATGATTAAGCCCCTGCTTTTTTAATA

AAAAGGGGACAAACCATTTCTTGACTTAATGTTGCAATACAAGTCATGTA
TAAGAGTGATTAACCTTTTTTTTATTTATAAAATAACTACAAAACATGTTT
TTTCATTATAGATCATGTATAAATGTGACTAATTTTTTTTCATCGCCTAAC
TTTTGTTGATAAATCATTAGAAATGTCACTAATTACTTTTTAGTATTTAT
5 AAAATAACTACAAAACATGTTTTTTTCATTATAGATCATGTATATATCAAC
TAAAAATATTATTCCCTTACACAAAAAAGGTTCAAGAAAGCCTGTA
TTTCGAAATAACTAAAAAGAAATATTTGATATTCATAAGAGAAATTTT
TTTCTAAACATGATCGCAAATGATTAAAACTTAAATTAAAACTAAAAAGA
TTTTTATATATGTTATNCAAAATTAATAATTTGAAATTAAGTTTATAATTC
10 TNGTNTCACAAAGGGATATATATAGTAAATATTATTTTTTTTGCAGTCAT
GCATAGTTGTATTTTTTAAATGATTTATTAACGTGGTAGGAGTGGAACCA
CTCAATCTAGTAGACCCACTATCACATGTCACATCAGCTTTACATCTATT
TTTCTTTCTCCTTTTTTCATCTTTTTTAAACTCATAACACNTAAAANTANC
ATATTTTCCAACACACTNAACTCATTGTCACATTATTATTTTTTAATTTAA
15 TTAATTNGAAATTAATAATTAANTAAANCNTAACATTTTTTAATTAATAA
AATATTAATCCAAATAAAAAANTNCACGATAAATTAATAANGTTTANTTTG
GAAAAAANCC (SEQ ID NO:106)
Sequence gap
ATAACCCTTTCAAGGGTCAACTCAAGTCCAAGTTAAAGTCAAGGTCAAAA
20 CCTTGGTTAAAGTCAACTTTGGTCAAAGTCAACATCTACTTGACTCACCT
CACCGAGTTGGTCCACCAACTTGTGCGAGTCCCTTAATCCACAACTTCAA
GAACCTCGATCCTACTCGTCGAGTCTTTCAAGAACTCTTCGAGTTTCCAT
TACACAGAATCGGGACCTTTTGCTCATGACTCGCCGAGTTCATCCTTGAA
CTTGTCGAGTCTAGCTTCATACGAGTTCGAGTGTTTAGTCCTTGACTCGT
25 CGAGTTCTTCCTTGAACCTCGTCGAGTCCATCTTCGTATAGTTGGGACATT
GCCCTGAACTCACCGAGTTCATCATTGAACCTCATCGAGTCCCTTCGATCTT
CAAGTCCATAATCCTGTCCATCTTGTTGAGTCCTCTTCTAGACTCAACCA
GATTCCTCAGAAACAGAAAAGGTTAGGGAACCATTACCTGACTCGCCGAG
TCCCAAGAACGAATCCCCGAGTCCCCCAATGTCCATGACCATAACAATCGA
30 TTTTCGTTGGGCTCATTGCATCCAAAGCATAGATCTAACCTCCTAGGGTC
CATATTACACGTAAAGCTACGAACCTTGACGTCCATGCATGGGGGATTTGG
CTCAAATGGCATTAAAATGGGGTTTATCTGATGCATGGGACTCCCATGGC
CATAAAGTTAACACCTTTATGCCATGGGAATCCTCAATGGTTCCATATCT
GAAGTTAACTCTACAATATGTTCTAAACCCGAAGGTGGCTTAGAAATG
35 CCCCAAAATGGCAAGATTCAAGCCTTAAAGGAGATCTAACAAATGATAAG
TCAAGGTTCAAGCTTTTTACCTTGAATAAGCTGGAAATGAAGCAAAATCT
CTGGATCCACTTGCTTCTTCAAGAACCCCCAAGCTTCCACTTCTTCCTTC
AAGTTTCAACAACCTTTAAACACTCAAAAATGGCTCAAGAACACTCAAAA
AGCTTTAGGGTTTCGAGTTAGGGCTTTTTGGAAGCGAGAGGGACGATGGG
40 GGCTGAAATGAGGCTAGAAAAAGTGTTTAAATAGGGGGCAAACCCTAAAT
ATTAGGGTTTCATCCAGGCAGCCCTACTCGTCGAGTCGGGCTCCCGACTC
GTCGAGTAGGTCACTTAAAACCCGCGTCCATAATCCAGTCTACTCGACGA
GTTGGGCCTCCAACCTCGTCGATTCCGAGTGCAAAACGTTCAATTACTTAA

ATTTAAATATGTACCAGGAACCGGGTGTACAGTTGAGACTTTATACCTC
CATAAGATAGATCTAGGTGCACATAGCCTGGATCCACAAGCTCCATGTCA
ACAAGCGACTCTTCAAGAAGTTCATTCTTCCTCCTTAAGCACCAAAAAAC
ACACAAAATCACCATGAAGCTCAAGAAATACTCAAATAGAGGATAGGGTT
5 TCGTTCGTAGGGTTAGAGAGGATGGAGGCTAGAGGAAATGAGGGATAGAG
GCGAGTTAAGGTCTTTAAATAGGGTCCAAGACCCTAAATTAGGGTTTTAA
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T (SEQ ID NO:107)

25

RG2J deduced polypeptide sequence (SEQ ID NO:108)

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15 SIHKIKQQDFKKAQDSI.CEVNTR

RG2K polynucleotide sequence (SEQ ID NO:109) and (SEQ ID NO:110)

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(SEQ ID NO:110)

10 **RG2K deduced polypeptide sequence (SEQ ID NO:111)**

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20 NHGNGGMLGWPENDMSASSCKRISLICKGMSDFPRDVKFPNLLILKLMHADKS
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GNLLNLEVLFSFANGIEWLPSTIGNLKELRVLDLTNCDGLRIDNGVLKKLVKLEELY
MRVGGRYQKAISFTDENCNEMAERSKNLSALEFEFFKNNAPKNMSFENLERFKIS
VGCFYKGDGFKIFHSFENTLRLVTNRTEVLESRLNELFEKTDVLYLSVGDMDNDLED
25 VEVKLAHLPKSSSFHNLRLVLISECIELRYLFTLDVANTLSKLEHLQVYECDNMEEII
HTEGRGEVTITFPKLKFLSLCGLPNLLGLCGNVHIINLPQLTELKLNIGIPGFTSIYPEK
DVETSSLLNKEVVIPNLEKLDISYMKDLKEIWPCELGMSQEVDVSTLRVIKVSSCDN
LVNLFPCNPMPLIHHLEELQVIFCGSIEVLFNIELDSIGQIGEGINNSSLRRIQLQNLGK
LSEVWRIKGADNSSLLISGFQGVESIIVNKCKMFRNVFTPTTTNFDLGALMEIRIQDC
30 GEKRRNNELVESSQEQQ

RG2L polynucleotide sequence (SEQ ID NO:112)

GGAAGACACAATGATGCAAAGACTGAAGAAGGTTGCCAAAGAAAATAGAA
TGTTCA GTTACATGGTTCGAGGCAGTTATAGGGGAAAAGACAGACCCAATT
35 GCTATTCAACAAGCTGTAGCCGATTACCTTCGTATACAGTTCAAAGAAAG
CACTAAACCAGCAAGAGCTGATAAGCTTCGTGAATGGTTCAAGGCCCACT
CTGNAGACGGTAAGAATAAGTTCCTCGTAATATTTGATGACGTCTGGCAG
TCCGTTGATCTGGAAGATATTGGNTTAAGTCCTTTTCCAAATCAAGGTGT
CGACTTCAAGGTCTTGTTGACTTCACGAGACGAACACGTTTGACAATGA
40 TGGGGGTTGAAGCTAATTCAGTTATTAATGTGGGACTTCTAACTGAAGTA
GAAGCACAAAGTCTGTTCCAGCAATTTGTAGAACTTTTGAGCCCCGAGCT
CTGTAAGATAGGAGAAGTTATCGTAAGAAAGTGTGCGGTCTACCTATTG

CCATCAAAACCATGGCGTGTACTCTAAGAAATAAAAGAAAGGATGCATGG
AAGGATGCACTTTCACGTATAGAGCACTATGACATTCGTAGTGTTCGCC
TAAAGTCTTTGAAACAAGCTATCACAATCTCCAAGACAGGGAGACTAAAT
CCGTGTTTTTGATGTGTGGTTTGTTCCTGAAGACTTCAATATTCCTACC
5 GAGGAGTTGATGAGGTATGGATGGGGCTTAAAGCTATTTGACAGAGTTTA
TACAATTAGAGAAGCAAGAACCAGGCTCAACACCTGCATTGAGCGACTTG
TGCAGACAAATTTGTTAATTGAAAGTGATGATGTTGGGTGTGTCAAGATG
CATGATCTGGTGCCTGCTTTTGTTCCTGAAGTTCGAGCA
TGCTTCAATTGTCAACCATGGTAATATGCATGGGTGGACTAAAAATGATA
10 TGAACGACTCTTGCAAAACAGTTTCTTTAACATGCGAGAGTGTGTCTGAG
TTTCCAGGAGACCTCAAGTTTCCAAACCTAAAGCTTTTGAACTTATGCA
TGGAGATAAGATGCTAAGGTTTTCTCAAGACTTTTATGAAGGAATGGAAA
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TCACCTCAATGCTCCACCAACCTTCGAGTGCTTCATCTTCATCGGTGTTT
15 ATTACGGATGCTTGATTGCTCTTGATCGGAAATTTGACGAATCTGGAAG
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AATTTGAAGAAGCTTAGGCAACTTGATCTGAGAGGTCGTTATGGTCTTTG
TATAGAACAGGGTGTCTTGAAAAATTTGGTCGAACTTGAAGAACTTTATA
TTGGAAATGCATCTGCGTTTAGAGATTATAACTGCAATGAGATGGCAG
20

RG2L deduced polypeptide sequence (SEQ ID NO:113)

EDTMMQRLKKVAKENRMFSYMVEAVIGEKTDPPIAQQAVADYLRIQFKESTKPAR
ADKLREWFKAHS?DGKNKFLVIFDDVWQSVLDLEDIGLSPFPNQGVDFKVLLTSRDE
HVCTMMGVEANSVINVGLLTEVEAQSLFQQFVETFEPELCKIGEIVVRKCCGLPIAI
25 KTMACTLRNKRKDAWKDALSRIEHYDIRSVAPKVFETSYHNLQDRETKSVFLMCG
LFPEDFNIPTEELMRYGWGLKLFDRVYTIREARLNTCIERLVQTNLLIESDDVGC
VKMHDLVRAFVLGMYSEVEHASIVNHGNMHGWTKNMNDSCKTVSLTCEVSEF
PGDLKFPNLKLLKLMHGDKMLRFSQDFYEGMEKLQVISYHKMKYPLLPSPQCST
NLRVLHLHRCSLRMLDCSCIGNLTNLEVLFSFANSIERIPSAIGNLKKLRQLDLRGR
30 YGLCIEQGV LKNLVELEELYIGNASAFRDYNCNEMA

RG2M polynucleotide sequence (SEQ ID NO:114)

GGGGAAGACACAATAGATGCAAAGGCTGAAGAAGTTGCCAAAGAAAAGAG
AATGTTCAAGTTATATCATTGAGGCGGTTATAGGGGAAAAGACAGACCCCA
35 TTTCCATTCAAGGAAGCTATATCATATTACCTTGGTGTAGAGCTCAATGCA
AATACTAAGTCAGTAAGAGCTGATATGCTTCGTCAAGGGTTCAAGGCCAA
ATCTGATGTAGGTAAGGATAAATTCTTAATAATACTCGACGATGTATGGC
AGTCTGTTGATTTGGAAGATATTGGATTAAAGTCCATTTCCAAATCAAGGT
GTTAACTTCAAGGTCCTGTAAACATCACGAGACCGACATATTTGCACTGT
40 GATGGGGGTTGAAGGTCATTGATTTTAAATGTGGGACTTCTCACAGAAG
CAGAATCAAAAAGATTGTTCTGGCAGTTTGTAGAAGGTTCTGATCCTGAG
CTCCATAAGATAGGAGAAGATATTGTAAGTAAGTGTGTGGTCTACCCAT

TGCCATTAAAACCATGGCATGTACACTTAGAGATAAAAGTACGGATGCAT
GGAAGGATGCACTGTCTCGTTTAGAGCATCATGACATTGAAAATGTTGCC
TCTAAAGTTTTTAGAGCGAGCTATGACCATCTCCAAGACGAGGAGACTAA
ATCCACTTTTTTCTATGTGGATTGTTTCCAGAAGATTCCAATATTCCTA
5 TGGAGGAGTTGGTGAGGTATGGGTGGGGATTGAAATTATTTAAAAAAGTG
TATACCATAAGAGAAGCAAGAACTAGGCTCAACACTTGCAATTGAGCGGCT
CATCTATACCAATTTGTTGATAAAAGTTGATGATGTTTCAGTGCATCAAGA
TGCATGATCTCATCCGTTCTTTTGTGTTTGGATATGTTTTCTAAAGTTGAG
CATGCTTCGATTGTCAACCATGGTAATACGCTAGAGTGGCCTGCAGATNA
10 TNTGCACGACTCTTGTAAGGGCTTTCATTAAACATGCAAGGGTANATGTG
AGTTTTGTGGAGACCTNAANTTTCCAACCCTAATGATTTTAAACTTATG
CATGGAGATAAATCGCTAAGGTTT

RG2M deduced polypeptide sequence (SEQ ID NO:115)

15 GEDTIDAKAEVAKERMFYSYIEAVIGEKTDPISIQEASYYLGVELNANTKSVRAD
MLRQGFKA KSDVGKDKFLILDDVWQSVLEDIGLSPFPNQGVNFKVLLTSRDRHI
CTVMGVEGHSIFNVGLL TEAESKRLFWQFVEGSDPELHKIGEDIVSKCCGLPIAKT
MACTLRDKSTDAWKDALSRLEHHDIENVASKVFRASYDHLQDEETKSTFFLCGLFP
EDSNIPMEELVRYGWGLKLFKKVYTIREARLNTCIERLIYTNLLIKVDDVQCIKM
20 HDLIRSFVLD MFSKVEHASIVNHGNTLEWPAD??HDSCKGLSLTCKG?CEFCGDL?F
PTLMILKLMHGDKSLRF

RG2N polynucleotide sequence (SEQ ID NO:116)

AGGTAAATCCATAACCCTAAATGTTGGTACGCTCATATATCAAATTGCG
25 TGTTTTGTTGAATGAAAAAGCATGCTCAAAAAACCAGTGTAAGGCACGG
TATATGACATATTTATAGTTACTGATAACAAATTATGATAATTTGGGTT
TACRTAAGTTAGGATTCGTACTTCAACCAAATGTAATAGTTTTGTGAGT
CTATCTATGTATTTGGGGAATCACATTAGCAACGGGATTGTACTAGTAAT
TCGAAAAAGTCTTTTAAATAATTTTCTGTTTATAATTTATGAATAGTTT
30 TAGCGACATCTAATATTAATAGAAATGTATCTGATATTGAATTAATGTCC
TTAATGTGAACATAGACCTTTTCCATTTACTAATGCCTAATTATTAGTTT
CTAATCAATAAATTTTAATTTCTGTTTTATGCTTCTAAGACAATAAAAAAT
CCATGATTTACCTTTAAATATTAACAAAAATGACCATAAATAAATAAAAA
ATTAGGATACCAAACCCCCCGCCATGCCCAATGTCTAAATATTCTTGAT
35 GCTTTTGCTTTTCCCTCTTTTCCCTGTTAGTCTATTATTCTGGAGAGTTT
GAGAGAGTTTCATACAAGAAAATTTCAAGAAGAAAGCAAAGGTCCAGGTA
TTCTCTTTTCTTAATTATGTATTAACCTTACAAGCATTTTTTACACGATCC
ATGGTTTTTTGTGTATGTTTTTCAAATTGAACTAGATTGGGACTTTTGC
CCTTGATGATTCATAAGATATTGCATGGAGTTGAGATTGTGTAAGAAAAG
40 TGGTGAATAGAAAGAGCAAGTGAATCCAGATATAGTATTGGTAATATATG
ATGATGAGATAGAGATATGTTAAACTGGCTAGAAAATTGTTTTAATTTG
AAATTTAGGTKGTTGAATTTGAAAGATACCAAGCTAATAACTAATTAGTT

ATGCTAAWTAGTTATAAAGAACAACAAACTCTTAGTTTTTTTTTTCATGA
TTTTCAACCTCTTTGTACCAAATAAATTATAGCAAAATTGAATATCATT
CTCTGCAATCAATCTTAACTTTTGTTATTATCATCATGTCTAAAATTGCC
ACAAGTTTATTTTCAAAGTCATATTGGATTATGAAAGGACTATTTTTACC
5 AATTACATCTTTACTTTATGGGCCAAAGCTAATACAATCCGACTAAACTA
AAGGAATATGGGATGCATATAGTTTGCTTCCCGATTATAGATTTCTATCT
AATTTGTCTATTGTACTAATTTAGGTGCCACCACAAGTAAATTTGTTAAA
TGGATATCGTTAATGCCATTCTTAAACCAGTTGTCGAGACTCTCATGGTA
CCCGTTAAGAAACACATAGGGTACCTCATTTCCCTGCAGGCAATATATGAG
10 GGAAATGGGTATCAAAATGAGGGGATTGAATGCTACTAGACTTGGTGTCG
AAGAGCATGTGAACCGGAACATAAGCAACCAGCTTGAGGTTCCAGCCCAA
GGCAGGGGTTGGTATGAAGAAGTAGGAAAGATCAATGCAAAAGTGGA AAA
TTTTCTAGCGATGTTGGCAGTTGTTTCAATCTTAAGGTTAGACACGGGG
TCGGAAGAGAGCCTCCAAGATAATTGAGGACATCGACAGTGTCTAGAGA
15 GAACACTCTATCATCATCTGGAATGATCATTCCATTCTTCTAGGAAGAAT
TGATTCCACGAAAGCATCCACCTCAATACCATCAACCGATCATCATGATG
AGTTCCAGTCAAGAGAGCAAACCTTTCACAGAAGCACTAAACGCACTCGAT
CCTAACCACAAATCCCACATGATAGCCTTATGGGGAATGGGCGGAGTGGG
GAAGACGACAATGATGCATCGGCTGAAAAAGGTTGTGAAAGAAAAGAAAA
20 TGTTTAATTTTATTGTTGAGGCGGTTGTAGGGGAAAAAACAGACCCCAT
GCTATTCAATCAGCTGTGGCAGATTACCTAGGTATAGAGCTCAATGAAAA
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GATCTGAATGATATTGGTTTAAAGTCCTTTACCAAATCAAGGTGTGCGACT
25 CAAGGTGTTGTTGACATCACGAGACAAAGATGTTTGCAGTGAAGATGGGAG
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CAAAGTTTATTCCACCAATTTGTAGAAATTTTCGGATGATGTTGATCGTGA
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TTGTCATCAAAACCATGGCGTGTACTCTTAGAGGAAAAAGCAAGGATGCA
30 TGGAAGAATGCACTTCTTCGTTTAGTGAACTACAACATTGAAAATATAGT
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AATCCACCTTTTTGCTTTGTGGAATGTTTCCCGAAGACTTTAATATTCCT
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GTATACTATAGGAGAAGCAAGAATCAGGCTCAACACATGCATTGAGCGGC
35 TCATTACATAAAATTTGTTGATTGAAGTTGATGATGTTAGGTGCATCAAG
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40 GCATGAAGATATATCATTGAGGTTTCCCAAAAACCTTTTATGAAGAAATGG
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TCATCACCGCAATGCTCCGTCAACCTTTGCGTGTTTCATCTCCATAAATG
CTCGTTAGTGATGTTTGACTGCTCTTGTATTGGAAATCTGTGCAATCTAG

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5 GATAACTGCAATGAGATGGCAGAACGTTCAAAAGACCTTTCTGCATTAGA
ACTTGAGTTCTTTGAAAACAATGCTCAGCCAAAGAATATGTCATTTGAGA
AGCTACAACGATTCCAGATCTCAGTGGGGTGCTATTTATATGGAGCTTCC
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10 TGTATGTTTAAGTGTGGGAGATATGAATGATCTTGAAGATRTTGAGGTT
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15 ATCCCCCAAGCTGAAGTTTTTATCTTTGTGTGGGCTACCAAAGCTATCGG
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20 TTTGATTGTATGGGGTATTATTGTTGGATGGGACTATTAATAAATGATTA
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25 AAGAATTGTGGTTCCATTGAATCGTTATTCAACATCCATTTGGATTGTGC
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ATCGTTATTCAACATTGACTTGGATTGTGCTGGTGCAATTGGGCAAGAAG
30 ACAACAGAAGCAGCTTAAGAAACATCAAAGTGGAGAATTTAGGGAAGCTA
AGAGAGGTGTGGAGGATAAAAGGTGGAGATAACTCTCGTCCCCCTTGTTCA
TGGCTTTCAATCTGTTGAAAGCATAAGGGTTACAAATGTAAGAGGTTTA
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35 AGAGAGTAGCCATGAGCAAGAGCAGGTAAGGATTTCAATTTCACTTTCKT
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40 TATAAATGTCACTAGTTACTTTTCAGTAAAATAACAAATTTAATAAATTA
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5 GTAGATTTCAAAGACACAAACACATCTTCATTTTATTTATTTATTTATTA
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10 TAATTAGGGACCAAAAACATAAATTCCTCCCAACCATAGGGACCATTCGT
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15 TACCAAAAAATTAATTACCTTAGCAAGTTATTTTCCCATTTAGGTTGTAT
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20 AGATCCAACCTATTTTTAATCTGTTGGCATTTCATCATTTGCAACTGTT
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25 GTATTCCCATCCTGTCTCATGCACTCTTTTCATAACCTCCATAAACTTAA
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GTCCAACAAGTAGAGAATTGGTAACAACCTCACCATAACCAACAACAACCT
ATTATACTTCCCAACCTCCAGGAATTGATTCTATGGAATATGGACAACAT
GAGTCATGTGTGGAAGTGCGGCAACTGGAATAAATTCTTCACTCTTCCAA
30 AAGAACAATCAGAATCCCCATTCCACAACCTCAGTAACATACATATTTAT
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:::GCACACAACCACCACTTTTTCCCTCATCTTGATTCTCTCACTCTAAA
35 GCAACTGAAGAATCTGAAGTGTATTGGTGGAGGTGGTGCCAAGGATGAGG
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40 ATGGGATTTGGAGGCATATAAAGTTACATTCATTTGAACAAGTATTACCA
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TTACTTTTCTAATTAGAAAAGGTCCACATGTCTAATTAGGTTTTCCATTC
TATGTGAATCCTCTATTCTGTCTGTAATCAAGCATCTTAGATTATTTATC

CATTTTCATAATTGTGTTTATATTGACAGTTTTTTTCTTTTATAGTTGT
AATTGCAACCTGTCATATWTTMWWKKCWWATKYWMWWARTAATACATTT
TATACCCWCTATACTAAGATA

5 **RG2N deduced polypeptide sequence (SEQ ID NO:117)**

LGKTTMMHRLKKVVKEKKMFNFIVEAVVGEKTDPIAIQSAVADYLGIELNEKTKPA
RTEKLRKWFVDNSAGKKILVILDDVWQFVDLNDIGLSPLPNQGVDFKVLLTSRDKD
VCTEMGAEVNSTFNVKMLIETEAQSLFHQFVEISDDVDRELHNIGVNIVRKCGGLPI
VIKTMACTLRGKSKDAWKNALLRLVNYNIENIVNGVFKMSYDNLQDEETKSTFLL
10 CGMFPEDFNPTTEELVRYGWGLKLFKKVYTIGEARIRLNTCIERLIHTNLLIEVDDVR
CIKMHDLVRAFLVDMYSKVEHASIVNHGNTLEWHVDNMHNSCKRLSLTCKGMSK
FPTDLKFPNLSILKLMHEDISLRFKPNFYEEMEKLEVISYDKMKYPLLSPSPQCSVNL
CVFHLHKCSLVMFDCSCIGNLSNLEVLFSFADSAIDLLPSTIGILKKLRLDLTNCYGL
CIANGVFKKLVKLEELYMTVVNNGGVRKAISL

15

RG2O polynucleotide sequence (SEQ ID NO:118)

TTGTAAAACGACGGCCAGTCGAATCGTAACCGTTCGTACGAGAATCGCTG
TCCTCTCCTTCATTTGAATCATGATATTTGAATATCGATACTTTTGACTG
TAGCTTTTGGGTCGATTTTTTAGCAAGATACATAACTGGCCAAACCCATT
20 GGCTATTTTAGCCCAAATATGAAATGGACTGGATTGTTTTTTCCTTTC
TAACACGCACACATCTGGCGATCAGTATCACTCCATTATGAAGACCTAGT
CAAATTCATTAACGTTTCAGTCGTTCTTCAAAGTTTCAAAGTTCCAACCT
CCAACCTCCCTCTTTTTTTTTCTTTCCTCGATTCTGATTTGAATCCGAT
TCTGCGACGAAGGAGAGCTTGGTCAGAGGGCTGTGATTCTTGAGTCTTGA
25 CCTCCGAATCTAGCTGGATTATTTTCGACACACCAGACCACGTATCAGGT
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30 TCCATTTTTGTGAAGGGATCTGGGTTAGTTTTTAAAGGATTGCTACAAC
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ATCCACCTTTATATATGTTAATTGGGAGTCTTCCATGGTGCAATGAATCT
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35 GCAATGGATGTTGTTAATGCCATTCTTAAACCAGTTGCCGAGACACTTAT
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TGAGGGATATGAGTAACAAAATGAGGGAGTTGAACGCTGCAAGACATGCT
GAAGAAGACCACTTGGACAGGAACATAAGAAGTCTGTTGAGATTTCAAA
TCAAGTTAGGAGTTGGTTAGAAGAAGTAGAAAAGATCGATGCAAAAAGTAA
40 AAGCCCTTCCTAGTGATGTCACCGCTTGTTGCAGTCTCAAGATCAAACAT
GAAGTCGGAAGGGAAGCCTTGAAGCTAATTGTGGAGATTGAAAGTGCCAC
AAGACAACACTCTTTGATCACCTGGACTGATCATCCCATTCCTCTGGGAA

AAGTTGATTCCATGAAGGCATCGATGTCCACAGCATCAACCGATTACAAT
GACTTTCAGTCAAGAGAAAAAACTTTTACTCAAGCATTGAAAGCACTTGA
ACCAAACAACGCTTCCACATGATAGCGTTATGTGGGATGGGTGGAGTGG
GGAAGACCACAATGATGCAAAGACTAAAAAAGTTGCTAAACAAAATAGA
5 ATGTTTCAGTTATATGGTTGAGGCAGTTATAGGGGAAAAGACGGACCCAAT
TGCTATTCAACAAGCTGTAGCGGATTACCTTCGTATAGAGTTAAAAGAAA
GCACTAAACCAGCAAGAGCTGATAAGCTTCGTGAATGGTTCAAGGCCAAC
TCTGGAGAAGGTAAGAATAAATTCCTTGTAATACTTGATGACGTCTGGCA
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10 TCGACTTCAAGGTCTTATTGACTTCACGAGACGAACATGTTTGCACAGTA
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15 GAAGGATGCACTTTCGCGTATAGAGCACTATGACCTTCGCAATGTTGCGC
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TCAGTGTTTTTGATGTGTGGTTTGTTCGGAAGACTTCAATATTCCTAC
TGAGGAGTTGATGAGGTATGGATGGGGATTAAAGATATTTGATAGAGTCT
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20 GTGCAGACAAATTTGTTAATTGAAAGTGATGATGTTGGGTGTGTCAAGAT
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AAACATTCCAGGAGACTTCAAGTTTCCAAACCTAACGATTTTGAAACTTA
25 TGCATGGAGATAAGTCGCTAAGATTTCCACAAGACTTTTATGAAGGAATG
GAAAGCTCCAGGTTATATCATACGATAAAATGAAGTATCCAATGCTTCC
CTTGTCTCCTCAATGCTCCACCAACCTTCGAGTGCTTCATCTCCATGAAT
GTTCAATTAAGATGTTTGATTGCTCTTGTATTGGAAATATGGCGAATGTG
GAAGTGTTGAGCTTTGCTAATTCTGGCATTGAAATGTTACCTTCCACTAT
30 CGGAAATTTAAAGAAGCTAAGGTTACTTGATTAAACAGATTGTCATGGTC
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TATATGGGATTTTCTGATCGACCTGATCAAACCTCGTGGTAATATTAGCAT
GACAGATGTCAGCTACAATGAATTAGCAGAACGTTCAAAAGGCCTTTCTG
CATTAGAGTTCCAGTTCTTTGAAAACAATGCCCAACCAAATAATATGTCG
35 TTTGGGAAACTTAAACGATTCAAGATCTCAATGGGATGCACTTTATATGG
AGGATCAGATTACTTTAAGAAAACGTATGCTGTCCAAAACACATTGAAGT
TGGTTACTAACAAAGGTGAACCTATTGGACTCTAGAATGAACGAGTTGTTT
GTTGAAACAGAAATGCTTTGTTTAAAGTGTTGATGATATGAATGATCTTGG
TGATGTTTGTGTGAAGTCCTCACGTTCTCCTCAACCTTCTGTGTTCAAAA
40 TTCTAAGAGTCTTTGTCGTTTCCAAGTGTTGAGTTGAGATACCTTTTC
ACAATTGGTGTAGCCAAGGATTTGTCAAATCTTGAGCATCTTGAAGTTGA
TTCATGTAATAATATGGAACAACCTCATATGTATTGAGAATGCTGGAAAAG
AGACAATTACATTCCTAAAGCTGAAGATTTTATCTTTGAGTGGGCTACCA

AAGCTTTCGGGTTTGTGCCAAAATGTCAACAACTTGAGCTACCACAACT
CATAGAGTTGAAACTTAAGGGCATTCCAGGGTTCACATGCATTTATCCGC
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5 TATATCTATATGTCTATAATTTGATTATATGATGTATTAGTGTGTTGGATG
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10 ATCTTGAAGAGCTTGAAGTCAAGAAATGTGGTTCCATTGAATCGTTATTC
AACATCGACTTGGATTGTGTTGATGCCATAGGAGAAGAAGACAACATGAG
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15 CACACCTACCACCACCAATTTTAATATGGGGGCACTTTTGGAGATATCAA
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25 TGAATTACAAGAAGTCACTGATACTATTTCTAATGTTGTATTACATCG
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30 ATGGAAGTGCAACAACCTGGAATAAATTTTACAACAATCAGAATCCCCAT
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TGGATGAAGAA

35

RG2O deduced polypeptide sequence (SEQ ID NO:119)

MDV V N A I L K P V A E T L M E P V K K H L G Y I I S S T K H V R D M S N K M R E L N A A R H A E E D H L D
R N I R T R L E I S N Q V R S W L E E V E K I D A K V K A L P S D V T A C C S L K I K H E V G R E A L K L I V E I E
S A T R Q H S L I T W T D H P I L G K V D S M K A S M S T A S T D Y N D F Q S R E K T F T Q A L K A L E P N N
40 A S H M I A L C G M G G V G K T T M M Q R L K K V A K Q N R M F S Y M V E A V I G E K T D P I A I Q Q A V A
D Y L R I E L K E S T K P A R A D K L R E W F K A N S G E G K N K F L V I L D D V W Q S V D L E D I G L S P F P
N Q G V D F K V L L T S R D E H V C T V M G V G S N S I L N V G L L I E A E A Q S L F Q Q F V E T S E P E L H K I

GEDIVRKCCGLPIAIKTMACTLRNKRKDAWKDALSRIEHYDLRNVAPKVFETSYHN
LHDKETKS VFLMCGLFPEDFNIPTEELMRYGWGLKIFDRVYTFIEARNRINTCIERL
VQTNLLIESDDVGCVKMHDLVRAFVLGMYSEVEHASVVNHGNIPGWTENDPTDSC
KAISLTCEMSGNIPGDFKFPNLTKLMHGDKSLRFPQDFYEGMEKLQVISYDKMK
5 YPMLPLSPQCSTNLRVLHLHECSLKMFDCCSIGNMANVEVLSFANSIEMIPLSTIGN
LKKLRLDLTDCHGLHITHGVFNNLVKLEELYMGFSDRPDQTRGNISMTDVSYNE
LAERSKGLSALEFQFFENNAQPNNMSFGKLKRFKISMGCTLYGGSDYFKKTYAVQ
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10 LSLSGLPKLSGLCQNVNKLLELPQLIELKLKGIPGFTCIYPQNKLETSSLLKEEVVIPKL
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EVKKCGSIESLFNIDLDCVDAIGEEDNMRS LRNIKVKNSWKLREVWC IKGENN SCPL
VSGFQAVESISIESCKRFRNVFTPTTTNFNMGALLEISIDDCGEYME NEKSEKSSQEQ
EQTDILSEEVLQEVTD TISNVVFTSCLIHSFYNNLRKLNLEKYGGVEV VFEIESSTS
15 RELVTYHKQQQQQQPIFPNLEELYLYMDNM SHVWKCNNWNKFLQQSESPFHN
LTIHMSDCKSIKYLFSPLMAELLSNLKRINIDECDGI

RG2P polynucleotide sequence (SEQ ID NO:120)

CCCATTGCTATT CAGGAAGCAGTAGCAGATTACCTCNGTATAGAGCTCAA
20 AGAAAAAACTAAATCNGCAAGAGCTGATATGCTTCGTAAAATGTTAGTTG
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25 GATGAAGAAGCACAAAGTTTGTTCATGGAGTTTGTACAAATTCGAGTGA
TGTTGATCCCAAGCTTCATAAGATAGGAGAAGATATTGTAAGAAAGTGTT
GTGGTTTGCCTATTGCCATCAAAACCATGGCCCTTACTCTTAGAAATAAA
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30 ACCAGGAGACTAAATATATCTTTTGTCTTGTGGATTGTTTCCCGAAGAC
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35 TTCTAAAGTGCAGGATGCTTCAATTGTCAACCATGGTAGCATGTCAGGGT
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40 CCGTTTCTTCCCTCGTCTCCTCAATATTGCTCCACCAACCTTCGAGTTCT
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TTGAAGAGGTTTATATGAGAGTTGCTGTTTCAAGCAAAAAGCCGAAAT
AGAAAAGCCATTAGCTTCACAGATGATAACTGCAATGAGATGGCAGAGCG
5 TTC

RG2P deduced polypeptide sequence (SEQ ID NO:121)

PIAIQEAVADYL?IELKEKTKSARADMLRKMLVAKSDGGKNKFLVILDDVWQFVDL
EDIGLSPLPNQGVNFKVLLTSRDVDVCTMMGVEANSILNMKILLDEEAQSLFMEFV
10 QISSDVDPKLHKIGEDIVRKCCGLPIAKTMALTLRNKSKDAWSDALSRLHHDLHN
FVNEVFGISYDYLQDQETKYIFLLCGLFPEDYNIPPEELMRYGWGLNLFKKVYTIRE
ARARLNTCIERLIHTNLLMEGDVVGCVKMHDALALAFVMDMFSKVQDASIVNHGS
MSGWPENDVSGSCQRISLTCKGMSGFPIDLNFPNLTILKLMHGDKFLKFPDFYEQ
MEKLQVVSFHEMKYPFLPSSPQYCSNLRVLHLHQCSLMFDCSCIGNLNFNLEVLSE
15 ANSGIEWLPSRIGNLKKLRLLDLTDCFGLRIDKGVKLNLVKLEEVYMRVAVRSKKA
GNRKAISFTDDNCNEMAERS

RG2Q polynucleotide sequence (SEQ ID NO:122)

TGGGGAAGACACAGTGATAGAAAARAAAAAGAATGTTGTGGAAAAGAGGA
20 AAATGTTTGATTATGCTGTTGTGGCGGTTATAGGGGAAAAGACGGACCCT
ATTGCTCTTCAGAAACTGTTGCGGATTACTTGCATATTGAGCTAAATGA
AAGCACTAACTAGCAAGAGCAGATAAACTTTGCAAATGGTTCAAGGACA
ACTCGGATGGAGGTAAGAAAAAGTTCCTCGTAATACTCGACGATGTTTGG
CAATCTGTTGATTTGGAAGATATTGGTTTAAGTACTCCTTTTCCAAATCA
25 AGGTGTCAACTTCAAGGTTTTGTTGACATCACGAAAGAGAGAAATTTGCA
CAATGATGGGAGTTGAAGCTGATTTAATTCTCAATGTCAAAGTCTTAGAA
GAAGAAGAAGCACAAAAGTTGTTCCCTCCAGTTTGTAGAAATTGGTGACCA
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30 AAGGATTCATGGAAGGACGCACTCTCTCGTTTAGAGGACCATGACACTGA
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35 TTG.AGCGACTCTTGGATTCAAATTTGTTGATTGAAAGTAACGATATTCGG
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40 GAACTTATGCANGGAGATAAGTCTCTAAGGTTTCCTCAAGACTTTTATC
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TGAATTTGGGAAGTCCTCAGCTTTTTTAATTCTAACATTGAATGGTTACCT
TCCACAATCAGAAATTTAAAAAAGCTAAGGCTACTAGATTTGAGATATTG
TGATCGTCTTCGTATAGAACAAGGTGTCTTGAAAAATTTGGTCAAACCTG
5 AAGAACTTTATACTGGATATACATCAGCGTTTACAGA

RG2Q deduced polypeptide sequence (SEQ ID NO:123)

GEDTVIEKKKNVVEKRKMFDYAVVAVIGEKTDPIALQKTVADYLHIELNESTKLAR
ADKLCKWFKDNSDGGKKKFLVILDDVWQSVLEDIGLSTPFPNQGVNFKVLLTSR
10 KREICTMMGV EADLILNVKVL EEEEEAQKLF LQFVEIGDQYHELHQIGVHIVKKCYG
LPIAIKTMALTLRNKRKDSWKDALSRLEDHDTENVANAVFEMNYRNLQDEETKAI
FLLCGLFPEDFDIPTEELVRYGWGLNLFKKVYTIRKARTRSHTCIERLLDSNLLIESN
DIRCVKIHDLVRAFLDMYCKVEHASIVNHGNMRTEYNMADSKTISLTYKSMMSG
FEFPGDLKFPNLTVLKLM?GDKSLRFPQDFYQSMEKLRVISYDKMKYPLLPSSPQCS
15 TNIRVLRRLHECSLRMFDCSCIGKLLNLEVL SFFNSNIEWLPSTIRNLKKLRLDLLRYC
DRLRIEQGV LKNLVKLEELYTG YTSAFTE

RG2S polynucleotide sequence (SEQ ID NO:124)

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20 CTTATTGATTCTTTGTGTTTCATTGAGTTGATTTTCATTATTACTACCTT
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25 AGGGTCATGCAGACGAAAATGACAGAGTTGAATACCTCAAGAATCAGTGT
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30 GACAGCTCTCCCTGATCAGTTGGACTGATGATCCAGTTCCTCTAGGAAGA
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35 CATAATATATCTGTAATTTGATTGTATGATGTGTTATTGTTTATATGTGG
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10 RCACACACACACACATAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGAG
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15 ACCAMACATGTGGACAMWTATGTATATAAWTAATTCACAATAGTCTTTAG
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5 CCAGGTACCATTGATCTTTTTAGAACCCAGTTGTCTGAAACACCCTGAT
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20 TTATTTGAAGGTGAAAGATCCAACCTATTTTTTAGCTGTTGGCATTTTCCA
TCATTTGCAACTGTTTCTTGAAAAAAAATACCTAAAATCAAAATAACCA
TTTTCAAATCCAAAATTATAAGAGAGAATTGTTAATGGACGTGGAATCGT
AAATCATTAACACAGTTCAGTACACAAGTTGCTAATTACATTTCTTGCTG
TGCAGATTGAAATTCTATCAGAGAAAGAGACATTACAAGAAGTCACTGAT
25 ACTAATATTTCTAATGATGTTGTATTATTCCTATCCTGTCTCATGCACTC
TTTTCATAACCTCCATAAACTTAAATTGGAGAGAGTTAAAGGAGTGGAGG
TGGTGTGTTGAGATAGAGAGTGAGAGTCCAACAAGTAGAGAATTGGTAACA
ACTCACCATAACCAACAACATCCTATTATACTTCCCAACCTCCAGGAATT
GGATCTAAGTTTTATGGACAACATGAGTCATGTGTGGAAGTGCAGCAACT
30 GGAATAAATTCTTCACTCTTCCAAAACAACAATCAGAATCCCCATTCCAC
AACCTCACAACCATAACATGTTTCACTGTCAGAAGCATTAAGTACTTGTT
TTCGCCTCTCATGGCAGAACTTCTTTCCAACCTAAAGGATATCTGGATAA
GTGGGTGTAATGGTATTAAAGAAGTTGTTTCAAAGAGAGATGATGAGGAT
GAAGAAATGACTACATTTACATCTACCCACACAACCACCATCTTGTTCCC
35 TCATCTTGATTCTCTCACTCTAAGACTACTGGAGAATCTGAAGTGTATTG
GTGGAGGTGGTGCCAAGGATGAGGGGAGCAATGAAATATCTTTCAATAAT
ACCCTGCAACTACTGCTGTTCTTGATCAATTTGAGGTATGCTTTGTACA
TATTCAATTATTTATTTAATTTTCCTTTTTTCTTTGCAATATTCTATAAAT
AATACATTTTATACCCACTATACTAAGATAATAATTACCTAGAGGGATGG
40 ATGCTATGACACAGCTGCTACACTTCAGAACTCTAGTAAGGGCAGTTAT
GGAAGTTCAATAAAATGATAATGGCATCTTTTGATGGGTAATATAGGCAA
TTTAAAGTTTTATTTCTGTTAAAGCAGTATTTAGCAAGTACTGGCCAGTAG
GAGAGGAGAATATCACCTTTTGTGAAAATCTGGTCATTGTACCCAAGAAT

TTAGTTAAATGTAACATTTTAGATATCAGGGGACATCAGGTGACAGATAT
TGTAGAATAGAACAAATATATAATATTACCCAAAACATTTTTTCTAAGGT
TATTCTGTAAATATGTGCTTTCTTGATTTCATTGAATTTGCATTCCCTAT
ATTTTAGGTGGTAAAGTGATTGTCTCTTCAATAAATCCCGAAATTAATTA
5 AAAAAAAAAAAACAAAAGTAAATTTTTGATATGGAGAGCACTGGTATCA
TTTAGTATATAAAAAAACTAGATTTTGAATTAAGTTTCTTATATAAAAGC
TGTGTATATAGTTTAATTAGTTTTACATCATTTTTTCCATGTGGTGTGCA
GTTGTCTGAAGCAGGTGGTGTCTTGGAGTTTATGCCAATACGCTAGAG
AGATAGAGATATCTAAGTGTAATGTATTGTCAAGTGTGATTCCATGTTAT
10 GCAGCAGGACAAATGCAAAAGCTTCAAGTGCTGAGAGTAACGGGTTGTGA
TGGCATGAAGGAGGTATTTGAACTCAATTAGGGACGAGCAGCAACAAAA
ACAGAAAGGGTGGTGGTGAAGGAAATGGTGGGAATCCAAGAGTAAAT
AACAAATGTTATTATGCTTCCCAATCTAAAGACATTGAAAATCTACATGTG
CGGGGGTTTGGAACATATATTCACATTCTCTGCACTTGAAAGCCTGACAC
15 AGCTCCAAGAGTTAAAGATAGTGGGTTGCTACGGAATGAAAGTGATTGTG
AAGAAGGAAGAAGATGAATATGGAGAGCAGCAAACAACAACAACAAC
AACGAAGGGGGCATCTTCTTCTTCTTCTTCTTCTTCTAAGAAGGTTG
TGGTCTTTCCCCGTCTAAAGTCCATTGAACTATTCAATCTACCAGAGCTG
GTAGGATTCTTCTTGGGGATGAATGAGTTCGGTTGCCTTCATTGGAAGA
20 AGTTACCATCAAGTATTGCTCAAAAATGATGGTGTTCGAGCTGGTGGGT
CCACAGCTCCCCAACTCAAGTATATACACACAAGATTAGGCAACATACT
CTTGATCAAGAATCTGGCCTTAACCTTCATCAGGTATATATATATTCCTT
TAATTGGCATGATCTAATTAAGAAAGATATCATTCCTGCCAAGTAAATTT
ACTTCAAACACATTACACTGGTTTCAGTCTAAGTTTATGTTGTTCTAGG
25 AAGGCCAAAATGGGAAAGCAAGATAGGGAAAAATAGTGTATTCAGTGGA
AAGGGTATTTTAGGTATTTTCTGTCAAAAGTTGTTATTGCAGGCTTTTTA
GTACCTGGAATCGTGTGTGGGAGGAGCGTTATTATTCTGATTTGCTTGTT
TCTTTATCATTTTTTTCTTAGCCTCTCGAACAGCTAGAAACCCTTTTAATC
TTTTGATTTTAAATGACAAAATTTTCCCTGTACTCTATTTGATTGTTG
30 TTCTTCATGGTTCTAAGTGAGTTATTGGCTCATCTGTTACTTCTTTTGAT
TGTTATTTTCATATCATGTTGTCCTTTGAATCAAGCTTTTCCATTTTCAA
CCAGGGCAAAGGTCAAAGTAACCTACTTTATGAGATCAAAAACAGCAA
CCCATCGGATAACTTTTAGTTGGAGTTAATAGTTACAATTACCATTGTGA
TTAATAATTATAATATCTTGTATTAATTCATTAAAAATTGGTACAGCACAT
35 ATATGACATTTTAAAGGTTTGTGTTTGTGACATATATATGCCTCTGGC
GTTTTCTTTATTGGACATGCAGACCTCATTCCAAAGTTTATACGGTGACA
CCTCGGGCCCTGCTACTTCAGAAGGGACAACCTGGTCTTTTCATAACTTG
ATCGAATTAGATATGGAATTAATTATGATGTTAAAAAGATTATTCCATC
CAGTGAGTTGCTGCAACTGCAAAAGCTGGAAAAGATTATGTGAGTAGTT
40 GTTATTGGGTAGAGGAGGTATTTGAACTGCATTGGAAGCAGCAGGGAGA
AATGGAAATAGTGGAATTGGTTTTGATGAATCGTCACAACTACTACTAC
TACTACTCTTTTCAATCTTCGAAACCTCAGAGAAATGAAGTTGCATTTTC
TACGTGGTCTGAGGTATATATGGAAGAGCAATCAGTGGACAGCATTTGAG

TTTCCAAACCTAACAAGAGTTCATATAAGTAGGTGTAGAAGGTTAGAACA
TGTATTTACTAGTTCCATGGTTGGTAGTCTATTGCAACTCCAAGAGCTAG
ATATTAGTTGGTGCAACCATATGGAGGAGGTGATTGTTAAGGATGCAGAT
GTTTCTGTTGAAGAAGACAAAGAGAGAGAATCTGATGGCAAGACGAATAA
5 GGAGATACTTGTGTTACCTCGTCTAAAATCCTTGAAATTAAAATGCCTTC
CATGTCTTAAGGGGTTTAGCTTGGGGAAGGAGGATTTTTCATTCCCATT
TTGGATACTTTAGAAATCTACAAATGCCAGCAATAACGACCTTCACCAA
GGGAAATTCTGCTACTCCACAGCTAAAAGAAATAGAAACAAGATTTGGCT
CGTTTTATGCAGGGGAAGACATCAACTCCTCTATTATAAAAAGATCAAAC
10 AACAGGTAAATCAGATCTTTGTTGCTTTAATAATTCTTAAACTACATTTG
AAAAGCTTCATGCAAGTTTTTTTTGTTATATTGTCAAAAACCGCAACCTA
CATTTTCAGCTTTATATTTATGTACTTTATGCAGGAGTTCAAACAAAAC
CTGATTAATGTGAAGTGAATATTAAAGGTAAATTATATTTTCATGTTCT
AGTTGCCTATTAATTAATGGCCTTTTAGTTTCRTGATTTTGGATGTAGTY
15 WTCATGATGATGTGAATCTTCTAATACCCCATTCATTGTTTGGTTGAATG
TTGACTCTATGTCAGGATGAATATTCAAGGGAAGAATTGTTTCATCATATG
AAGGACATTAAAGAACATGGATGCTATGAAGATGTTGGAARAC

RG2S deduced polypeptide sequence (SEQ ID NO:125)

20 MSDPTGIAGAIINPIAQRALVPVTDHVGYMISCRKYVRVMQTKMTELNTSRISVEEH
ISRNTRNHLQIPSQIKDWLDQVEGIRANVENFPIDVITCCSLRIRHKLQKAFKITEQI
ESLTRQLSLISWTDDPVPLGRVGSMASTASSSDDFPSREKTFTQALKALEPNQQF
HMYALCGMGGVGKTRMMQRLKKAEEKLKNFYIVRAVIGEKTDPFAIQEAIADYL
GIQLNEKTKPARADKLREWFKKNSDGGKTKFLIVLDDVWQLVDLEDIGLSPFPNQG
25 VDFKVLTSRDSQVCTMMGVEANSIINVGLLTEAEAQSLFQQFVETSEPELQKIGED
IVRKCCGLPIAIKTMACTLRNKRKDAWKDALSRIEHYDIHNVAPKVFETSYHNLQE
EETKSTFLMCGLFPEDFDIPTTEELMRYGWGLKLFDRVYTIREARLNTCIERLVQT
NLLIESDDVGCVKMHDLVRAFLGMFSEVEHASIVNHGNMPEWTENDITDSCKRIS
LTCKSMSKFPKDFKFPNLMILKLMHGDKSLRFPQDFYEGMEKLHVISYDKMKYPLL
30 PLAPRCSTNIRVLHLTKCSLKMFDCCIGNLSNLEVLSFANSRIEWLPSTVRNLKKLR
LLDLRFCDGLRIEQGVLSLVKLEEFYIGNASGFIDDNCNEMAERSDNLSALEFAFF
NNKAEVKNMSFENLERFKISVGRSFDGNINMSSHSYENMLQLVTNKGVDLDSKLN
GLFLKTKVLFVSVHGMNDLEDVEVKSTHPTQSSSFCNLKVLISKVELRYLFLKLN
ANTLSRLEHLEVCECENMEELIHTGICGEETITFPKLKFLSLSQLPKLSSLCHNVNIIG
35 LPHLVDLILKGIPGFTVIYPQNKLRSSLLKEEVVIPKLETQIDDMENLEEIWPCELS
GGEKVKLREIKVSSCDKLVNLFPRNPMSLLHHLEELKVKNCGSIESLFNIDLDCVGA
IGEEDNKSLLRSINMENLGKLREVWRIKGADNSHLINGFQAVESIKIECKRFSNIFT
PITANFYLVALLEIQIEGCGGNHESEEQIEILSEKETLQEVTDNINSDVVLFPSCLMH
SFHNLHKLKLERVKGEVVFEIESESPTSRELVTTHHNQQHPIILPNLQELDLSFMD
40 NMSHVWKCSNWNKFFTLPKQQSES PFHNLTIHMFSCRSIKYLFSPMAELLSNLK
DIWISGCNGIKEVSKRDEDEEMTTFTSTHTTTILFPHLDSLTLRLLENLKCIGGGG
AKDEGSNEISFNNTTATTAVLDQFELSEAGGVSWSLCQYAREIEISKCNVLSSVIPCY

AAGQMQLQVLRVTGCDGMKEVFETQLGTSSNKNRKGGGDEGNNGGIPRVNNNVI
MLPNLKTLLKIYMC GGLEHIFTFSALESLTQLQELKIVGCYGMKVIVKKEDEYGEQ
QTTTTTTTKGASSSSSSSSKKVVVFPRLKSIELFNLPELVGFFLGMNEFRLPSLEEV
IKYCSKMMVFAAGGSTAPQLKYIHTRLGKHTLDQESGLNFHQTSFQSLYGDTSGPA
5 TSEGTTWSFHNLIELDMELNYDVKKIIPSELLQLQKLEKIHVSSCYWVEEVFETAL
EAAGRNGNSGIGFDESSQTTTTTTLFNLRLNREMKLHFLRGLRYWKSQWTAFF
PNLTRVHISRCRRLEHVFTSSMVGSLLQLQELDISWCNHMEEVIVKDADVSVEEDK
ERESDGKTNKEILVLPRLKSLKLKCLPCLKGFSLGKEDFSFPLDLEIYKCPAITTFT
KGN SATPQLKEIETRFSGSYAGEDINSSIIKRSNNRSSNKT LINVK.ILK

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RG2T polynucleotide sequence (SEQ ID NO:126)

GGAAGACGACAATGGTGCAACGGTTGAAGAAGGTTGTGAAAGATAAGAAG
ATGTTCCATTATATTGTCGAGGTGGTTGTAGGGGCAAACACTGACCCCAT
TGCTATCCAGGATACTGTTGCAGATTACCTCAGCATAGAACTGAAAGGAA
15 ATACGAGAGATGCAAGGGCTTATAAGCTTCGTGAATGCTTTAAGGCCCTC
TCTGGTGGAGGTAAGATGAAGTTCCTAGTAATTCTTGACGATGTATGGAG
CCCTGTTGATCTGGATGATATCGGTTTAAGTTCCTTGCCAAATCAAGGTG
TTGACTTCAAGGTCTTGCTGACATCACGCAACAGTGATATCTGCATGATG
ATGGGAGCTAGTTTAATTTTCAACCTCAATATGTTAACAGACGAGGAAGC
20 ACATAATTTTTTCCGTCGATACGCAGAAATTTCTTATGATGCTGATCCCG
AGCTTATTAAGATAGGAGAAGCTATTGTAGAGAAATGTGGTGGTTTACCC
ATTGCCATCAAACTATGGCCGTTACTCTTAGAAATAAACGCAAAGATGC
ATGGAAAGATGCACTTTCTCGTTTAGAGCACCGTGACACTCATAATGTTG
TGGCTGATGTTCTTAAATTGAGCTACAGCAATATCCAAGACGAGGAGACT
25 CGGTCGATTTTTTTGCTATGTGGTTTGTTCCTGAAGACTTTGATATTCC
TACCGAAGACTTAGTGAGGTATGGATGGGGATTGAAAATATTTACCAGAG
TGTATACTATGAGACATGCAAGAAAAAGGTTGGACACGTGCATTGAGCGG
CTTATGCATGCCAACATGTTGATAAAAAGTGATAATGTTGGATTTGTCAA
GATGCATGATCTGGTTCGTGCTTTTGTTTTGGGCATGTTATCTGAAGTCG
30 AGCATGCATCAATTGTCAACCATGGGGATATGCCAGGGTGGTTTGAAACT
GCAAATGATAAGAACAGCTTGTGCAAAAAGAAATTCATTAACATGCAAAGG
TATGTCTGCGATTCTGAAGACCTCACGTTTCCAAACCTCTCGATCCTGA
AATTAATGGATGGAGACGAGTCACTGAGGTTTCCTGAAGGCTTTTATGGA
GAAATGGAAAACCTTCAGGTTATATCATATGATAACATGAAGCAGCCATT
35 TCTTCCACAATCACTTCAATGCTCCAATGTTTCGAGTGCTTCATCTCCATC
ACTGCTCATTAAATGTTTGATTGCTCTTCTATTGGAAATCTTTGAATCTC
GAGGTGCTCAGCATTGCTAATTCTGCCATTAAATTGTTACCCTCCACTAT
TGGAGATCTGAAGAAGCTAAGGCTCCTGGATTTGACAAATTGTGTTGGTC
TCTGTATAGCTAATGGCGTCTTTAGAAATTTGGTCAAACCTGAAGAGCTT
40 TATATGAGAGTTGATGATCGAGATTTCGTTTTTTGTGAAAGCTGATGACAG
CAAGACCATTACCT

RG2T deduced polypeptide sequence (SEQ ID NO:127)

KTMTMVQRLKKVVKDKKMFHYIVEVVVGANTDPIAIQDTVADYLSIELKGNTRDAR
AYKLRECFKALSGGGKMKFLVILDDVWSPVDLDDIGLSSLPNQGVDFKVLLTSRNS
DICMMMGASLIFNLNMLTDEEAHNFFRRYAEISYDADPELIKIGEAIVEKCGGLPIAI
5 KTMAVTLRNRKDAWKDALSRLEHRDTHNVVADVLKLSYSNIQDEETRSIFLLCG
LFPEDFDIPTEDLVRYGWGLKIFTRVYTMRHARKRLDTCIERLMHANMLIKSDNVG
FVKMHDLVRAFVLGMLSEVEHASIVNHGDMPGWFETANDKNSLCKRISLTCKGMS
AIPEDLTFPNLSILKLMDGDESLRFPGEFYGEMENLQVISYDNMKQPFLPQSLQCSN
VRVLHLHHCSLMFDCSSIGNLLNLEVLSIANSIAIKLLPSTIGDLKKLRLLDLTNCVGL
10 CIANGVFRNLVKLEELYMRVDDRDSFFVKADDSKTTT

RG2U polynucleotide sequence (SEQ ID NO:128)

GCCTTGTGTGGGATGGGTGGAGTGGGAAAGACCACTGTGATGAAGAAGCT
GAAGGAGGTTGTGGTAGGAAAGAAACTGTTTAATCATTATGTTGAGGCGG
15 TTATAGGGGAAAAGACAGACCCCATTTGCTATTCAACAAGCTGTTGCCGAG
TACCTTGGTATAAGTCTAACCGAAACCACTAAACCAGCAAGAACTGATAA
GCTCCGTACATGGTTTGCAAACAACCTCAAATGGAGGAAAGAAGAAGTTCC
TGGTAATACTAGACGATGTATGGCAACCAGTTGATTTGGAAGATATTGGT
TTAAGTCGTTTTCCAAATCAAGATGTTGACTTCAAGGTCTTGATTACATC
20 ACGGGACCAATCAGTTTGCAGTGGAGGTTAAAGCTGATTTAGTTC
TCAAGGTGAGTGTCTTGGAGGAAGCGGAAGCACACAGTTTGTTCCTCCAA
TTTTTAGAACCTTCTGATGATGTCGATCCTGAGCTCAATAAAATCGGAGA
AGAAATTGTAAAGAAGTGTTCGAGACTACCCATTGCTATCAAAACCATGG
CCTGAACCTCTTAGAAGTAAAAGTAAGGATACATGGAAGAATGCCCTTTCT
25 CGTTTACAACACCATGACATTAACACAATTGCGTCTACTGTTTTCCAAAC
TAGCTATGACAATCTCGAAGACGAGGTGACTAAAGCTACTTTTTTGCTTT
GTGGTTTATTTCCGGAGGACTTCAATATTCTTACCGAGGACCTATTGAGG
TATGGATGGGGATTGAAGTTATTCAAGGAAGTAGATACTATACGAGAAGC
AAGATCCAAGTTGAAAGCCTGCATTGAGCGGCTCATGCATACCAATTTGT
30 TGATCGAAGGTGATGATGTTAGGTACGTAAAGATGCATGATCTGGTGCGT
GCTTTTGTGTTTGGATATGTTTTCTAAAGCCGAGCATGCATCTATTGTCAA
CCATGGTAGTAGTAAGCCAAGGTGGCCTGAAACTGAAAGTGATGTGAGCT
CCTCTTGCAAAGAATTTCATTAACATGCAAGGGTNTG

RG2U deduced polypeptide sequence (SEQ ID NO:129)

ALCGMGGVGKTTVMKKLKEVVVGKKLFNHYVEAVIGEKTDPPIAIQQA VAEYLGIS
LTETTKPARTDKLRTWFA NNSNGGKKKFLVILDDVWQPVDLEDIGLSRFPNQDVD
FKVLITSRDQSVCTEMGVKADLV LKVSVLEEAEAHSLFLQFLEPSDDVDPELNKIGE
EIVKKCCRLPIAIKTMA.TLRSKSKDTWKNALSRLQHHDINTIASTVFQTSYDNLEDE
40 VTKATFLLCGLFPEDFNIPTEDLLRYGWGLKLFKEVD TIREARSKLKACIERLMHTN

LLIEGDDVRYVKMHDLVRAFVLDMFSKAEHASIVNHGSSKPRWPETESDVSSCKR-
ISLTCKG?

RG2V polynucleotide sequence (SEQ ID NO:130)

5 CTGTGGAAGACACGAATGATSAAGAAGCTGAAGGAGGTCGTGGAACAAAA
GAAAATGTTCAATATTATTGTTCAAGTGGTCATAGGAGAGAAGACAAACC
CTATTGCTATTCAGCAAGCTGTAGCAGATTACCTCTCTATTGAGCTGAAA
GAAAACACTAAAGAAGCAAGAGCTGATAAGCTTCGTNAATGGTTCGAGGA
CGATGGAGGAAAGAATAAGTTCCTTGTAATACTTGATGATGTATGGCAGT
10 TTGTCGATCTTGAAGATATTGGTTTAAGTCCTCTGCCAAATAAAGGTGTC
AACTTCAAGGTCTTGTGACGTTAAGAGATTCACATGTTTGCACTCTGAT
GGGAGCTGAAGCCAATTCAATTCTCAATATAAAAAGTTTTAAAAGATGTTN
AAGGACAAAGTTTTGTTCCGCCAGTTTGCTAAAAATGCAGGTGATGATGAC
CTGGATCCTGCTTTCAATGGGATAGCAGATAGTATTGCAAGTAGATGTCA
15 AGGTTTGCCCATGCCATCAAAACCATTGCCTTAAGTCTTAAAGGTAGAA
GCAAGCCTGCGTGGGACCATGCGCTTTCTCGTTTGGAGAACCATAAGATT
GGTAGTGAAGAAGTTGTGCGTGAAGTTTTTAAAATTAGCTATGACAATCT
CCAAGATGAGGTTACTAAATCTATTTTTWTACTTTGTGCTTTATTTCTG
AAGATTTTGATATTCCTATTGAGGAGTTGGTGAGGTATGGGTGGGGCTTG
20 AAATTATTTATAGAAGCAAAAACCTATAAGAGAAGCAAGAAACAGGCTCAA
CACCTGCACTGAGCGGCTTAGGGAGACAAATTTGTTATTTGGAAGTGATG
ACATTGGATGCGTCAAGATGCACGATGTGGTGCGTGATTTTGTTTGGTAT
ATATTCTCAGAAGTCCAGCACGCTTCAATTGTCAACCATGGTAATGTGTC
AGAGTGGCTAGAGGAAAATCATAGCATCTACTCTTGTAAGAATTTTCAT
25 TAACATGCAAGGGTATGTCTGAGTTTCCCAAAGACCTCAAATTTCCAAAC
CTTTCATTTTGAACTTATGCATGGAGATAAGTCGNTGAGCTTTCCTGA
AGACTTTTATGGAAAGATGGAAAAGGTTTCAGGTAATATCATATGATAAAT
TGATGTATCCATTGCTTCCCTCATCACTTGAATGCTCCACTAACGTTCTGA
GTGCTTCATCTCCATTATTGTTTCATTAAGGATGTTTGATTGCTCTTCAAT
30 TGGTAATCTTCTCAACATGGAAGTGCTCAGCTTTGCTAATTCTAACATTG
AATGGTTACCATCTACAATTGGAAATTTGAAGAAGCTAAGGCTACTAGAT
TTGACAAATTGTAAAGGTCTTCGTATAGATAATGGTGTCTTAAAAAATTT
GGTCAAACCTGAAGAGCTTTATATGGGTGTTAATGTCCGTATGGACCAGG
CCGT

35

RG2V deduced polypeptide sequence (SEQ ID NO:131)

LWKTRM?KKLKEVVEQKKMFNIIVQVVIGEKTNPPIAQAVADYLSIELKENTKEAR
ADKLR?WFEDDGGKNKFLVILDDVWQFVDLEDIGLSPLPNKGVNFKVLLTLRDSH
VCTLMGAEANSILNIKVLKDV?GQSLFRQFAKNAGDDDLDPAFNGIADSIASRCQGL
40 PIAIKTIALSLKGRSKPAWDHALSRLNHNKIGSEEVVREVFKISYDNLQDEVTKSIF?L
CALFPEDFDIPEELVRYGWGLKLFIEAKTIREARNRLNTCTERLRETNLLFGSDDIG

CVKMHDVVRDFVWYIFSEVQHASIVNHGNVSEWLEENHSIYSCKRISLTCKGMSEF
PKDLKFPNLSILKLMHGDKS?SFPEDFYGKMEKVQVISYDKLMYPLLPSSLECSTNV
RVLHLHYCSLRMFDCSSIGNLLNMEVLSFANSNIEWLPSTIGNLKKLRLLDLTNCKG
LRIDNGVLKNLVKLEELYMGVNVVRMDQAV

5

RG2W polynucleotide sequence (SEQ ID NO:132)

TTGGGAAAGAGACAATGATGAAGAATTGAAAGAGGTTGTGGTTGAAAAGA
AAATGTTTAATCATTATGTGGAGGCGTTATAGGGGAGAAGACGGACCCC
ATTGCTATTCAAGCAAGCCGTTGCAGAGTACCTTGGTATAATTCTAACAGA
10 AACCCTAAGGCAGCAAGAACCGATAAGCTACGTGCATGGCTTTCTGACA
ATTCAGATGGAGGAAGAAAGAAGTTCCTAGTAATACTAGACGATGTATGG
CATCCGGTTGATATGGAAGATATTGGTTTAAGTCGTTTCCCAAATCAAGG
TGTCGACTTCAAGGTCTTGATTACATCACGGGACCAAGCTGTTTGCCTG
AGATGGGAGTTAAAGCTGATTCAAGGTATCAAGGTGAGTGTCTAGAGGAA
15 GCTGAAGCACAAAGCTTATTCTGCCAACTTTGGGAACCTTCTGATGATGT
CGATCCTGAGCTCCATCAGATTGGAGAAGAAATTGTAAGGAAGTGTGTG
GTTTACCCATTGCAATAAAAACCATGGCCTGCACTCTTAGAAGTAAAAGC
AAGGATACATGGAAGAATGCACTTTCTCGTTTACAACACCATGACATTAA
CACAGTCGCGCCTACTGTTTTTCAAACCAGCTATGACAATCTCCAAGATG
20 AGGTGACTGGAGATACTTTTTTGCTATGTGGTTTGTTCGGAGGACTTC
GATATTCTACTGAAGACTTATTGAAGTATGGATGGGGCTTAAATTATT
CAAGGGAGTGGATTCTGTAAGAGAAGCAAGATACCAGTTGAACGCCTGCA
TTGAGCGGCTCGTGCATACCAATTTGTTGATTGAAAGTGATGTTGTTGGG
TGCGTCAAGTTGCACGATCTGGTGCCTGCTTTATTTTGGATATGTTTTG
25 TAAAGCGGAGCATGCTTCGATTGTCAACCATGGTAGTAGTAAGCCTGGGT
GGCCTGAAACTGAAAATGATGTGATCAGGACCTCCTGCAAAAGAATCTCA
TTAACATGCAAGGGTATGATTGAGTTTTCTAGTGACCTCAAGTTTCCAAA
TGTCTTGATTTTAAACTTATGCATGGAGATAAGTCGCTAAGGTTT

30 **RG2W deduced polypeptide sequence (SEQ ID NO:133)**

WERDNDEELKEVVVEKKMFNHYVEAVIGEKTDPIDQAVAEYLGILTETTKAAR
TDKLRAWLSDNSDGGRRKKFLVILDDVWHPVDMEDIGLSRFPNQGVDFKVLITSRD
QAVCTEMGVKADSVIKVSVLEEAEQSLFCQLWEPSSDDVDPELHQIGEEIVRKCCG
LPLAIKTMACTLRSSKSDTWKNALSRLQHHDINTVAPTQFQTSYDNLQDEVTGDTF
35 LLCGLFPEDFDIPTEDLLKYGWGLKLFKGVDSVREARYQLNACIERLVHTNLLIESD
VVGCVKLHDLVRAFILDMFCKAEHASIVNHGSSKPGWPETENDVIRTSCKRISLTCK
GMIEFSSDLKFPNVLILKLMHGDKSLRF

RG5 polynucleotide sequence (SEQ ID NO:134)

40 GGGGGGGTGGGGAAGNCGACTCTAGCCCAGAAGNTCTATAATGACCATAA
AATAAAAGGAAGCTTTAGTAAACAAGCATGGATCTGTGTTTCTCAACAAT

ATTCTGATATTTTCAGTTTTGAAAGAAGTCCTTCGGAACATCGGTGTTGAT
TATAAGCATGATGAAACTGTTGGAGAACTTAGCAGAAGGCTTGCAATAGC
TGTCGAAAATGCAAGTTTCTTTCTTGTGTTGGATGATATTTGGCAACATG
AGGTGTGGACTAATTTACTCAGAGCCCCATTAAACACTGCAGCTACAGGA
5 ATAATTCTAGTAACAACTCGTAATGATACAGTTGCACGAGCAATTGGGGT
GGAAGATATTCATCGAGTAGAATTGATGTCAGATGAAGTAGGATGGAAT
TGCTTTTGAAGAGTATGAACATTAGCAAAGAAAGTGAAGTAGAAAACCTA
CGAGTTTTAGGGGTTGACATTGTCGTTTGTGTGGTGGCCTCCCCCTAGC
CTT

10

RG5 deduced polypeptide sequence (SEQ ID NO:135)

GGVGKTTLAQK?YNDHKIKGSFSKQAWICVSQQYSDISVLKEVLRNIGVDYKHDET
VGELSRRLAIAVENASFLLVLDIWDQHEVWTNLLRAPLNTAATGILVTRNDTVA
RAIGVEDIHRVELMSDEVGWKLLKSMNISKESEVENLRVLGVLDIVRLCGGLPLAL

15

RG7 polynucleotide sequence (SEQ ID NO:136)

GGTGGGGTTGGGAAGACAACGGGCACAAGGAGGCGACTGCCAATACTTCC
GACTTTTATTCATAGAGATGACGAGTCTTATTTTCTACTACTATAGGGA
GGATATTTGGTTGCGCGAGACGATTCATTGCGCGAAGGGATTCTATCCTT
20 CTTTTTTTCCGCGAAGACTTCGTTCCGGAGGACGGGCTATATTCCCTTTA
ATATTAGTCTAGCCCAGTCTAGGCCAACCATATGGCGATGCGGTAGACCT
CCCAGAGATAGATACTTGATCTTAGAGGATTCACACGTTCAATGGTGGAA
ACTTAAGGAACCGGCTAAGAGTGACTAAACGGAAAAACCCTATTCATTCC
ATAGCCTCATCCGGTCGAGGCATTAAACAATCCATCCCAATCCTCTTTCC
25 TTTGGTCTACTCTAATGATGTGCCCGTTTCGTTGGTGGGAATATCTCTTTAT
ACCGACGATTTATATGGGGATTGCCACTAGCGTTG

30

The above examples are provided to illustrate the invention but not to limit
its scope. Other variants of the invention will be readily apparent to one of ordinary skill
in the art and are encompassed by the appended claims. All publications, patents, and
patent applications cited herein are hereby incorporated by reference.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid construct comprising an RG polynucleotide which encodes an RG polypeptide having at least 60% sequence identity to an RG polypeptide from an RG family selected from the group consisting of: an RG1 polypeptide, an RG2 polypeptide, an RG3 polypeptide, an RG4 polypeptide, an RG5 polypeptide, and an RG7 polypeptide.
2. The nucleic acid construct of claim 1, wherein the RG polynucleotide encodes an RG polypeptide comprising an leucine rich region (LRR).
3. The nucleic acid construct of claim 1, wherein the RG polynucleotide encodes an RG polypeptide comprising a nucleotide binding site (NBS).
4. The nucleic acid construct of claim 1, wherein the polynucleotide is a full length gene.
5. The nucleic acid construct of claim 1, wherein the further encodes a fusion protein.
6. The nucleic acid construct of claim 1, wherein the RG1 polypeptide is encoded by an RG1 polynucleotide sequence.
7. The nucleic acid construct of claim 6, wherein the RG1 polypeptide is encoded by a polynucleotide sequence selected from the group consisting of SEQ ID NO:1 (RG1A), SEQ ID NO:2 (RG1B), SEQ ID NO: 3 (RG1C), SEQ ID NO:4 (RG1D), SEQ ID NO:5 (RG1E), SEQ ID NO:6 (RG1F), SEQ ID NO:7 (RG1G), SEQ ID NO:8 (RG1H), SEQ ID NO:9 (RG1I), and SEQ ID NO:10 (RG1J).
8. The nucleic acid construct of claim 1, wherein the RG2 polypeptide is encoded by an RG2 polynucleotide sequence.
9. The nucleic acid construct of claim 8, wherein the RG2 polypeptide is encoded by a polynucleotide sequence selected from the group consisting of: SEQ ID NO:21 (RG2A);

SEQ ID NO:23 (RG2B); SEQ ID NO:25 (RG2C); SEQ ID NO:27 (RG2D); SEQ ID NO:29 (RG2E); SEQ ID NO:31 (RG2F); SEQ ID NO:33 (RG2G); SEQ ID NO:35 (RG2H); SEQ ID NO:37 (RG2I); SEQ ID NO:39 (RG2J); SEQ ID NO:41 (RG2K); SEQ ID NO:43 (RG2L); SEQ ID NO:45 (RG2M); SEQ ID NO:87 (RG2A); SEQ ID NO:89 (RG2B); SEQ ID NO:91 (RG2C); SEQ ID NO:93 (RG2D) and SEQ ID NO:94 (RG2D); SEQ ID NO:96 (RG2E); SEQ ID NO:98 (RG2F); SEQ ID NO:100 (RG2G); SEQ ID NO:102 (RG2H); SEQ ID NO:104 (RG2I); SEQ ID NO:106 (RG2J) and SEQ ID NO:107 (RG2J); SEQ ID NO:109 (RG2K) and (SEQ ID NO:110 (RG2K); SEQ ID NO:112 (RG2L); SEQ ID NO:114 (RG2M); SEQ ID NO:116 (RG2N); SEQ ID NO:118 (RG2O); SEQ ID NO:120 (RG2P); SEQ ID NO:122 (RG2Q); SEQ ID NO:124 (RG2S); SEQ ID NO:126 (RG2T); SEQ ID NO:128 (RG2U); SEQ ID NO:130 (RG2V); and, SEQ ID NO:132 (RG2W).

10. The nucleic acid construct of claim 1, wherein the RG3 polypeptide is encoded by an RG3 polynucleotide sequence.

11. The nucleic acid construct of claim 10, wherein the RG3 polypeptide is encoded by a polynucleotide sequence as set forth in SEQ ID NO:68.

12. The nucleic acid construct of claim 1, wherein the RG4 polypeptide is encoded by an RG4 polynucleotide sequence.

13. The nucleic acid construct of claim 12, wherein the RG4 polypeptide is encoded by a polynucleotide sequence as set forth in SEQ ID NO:69.

14. The nucleic acid construct of claim 1, wherein the RG5 polypeptide is encoded by an RG5 polynucleotide sequence.

15. The nucleic acid construct of claim 14, wherein the RG5 polypeptide is encoded by a polynucleotide sequence as set forth in SEQ ID NO:134.

16. The nucleic acid construct of claim 1, wherein the RG7 polypeptide is encoded by an RG7 polynucleotide sequence.
17. The nucleic acid construct of claim 16, wherein the RG7 polypeptide is encoded by a polynucleotide sequence as set forth in SEQ ID NO:136.
18. The nucleic acid construct of claim 1, further comprising a promoter operably linked to the RG polynucleotide.
19. The nucleic acid construct of claim 18, wherein the promoter is a plant promoter.
20. The nucleic acid construct of claim 19, wherein the plant promoter is a disease resistance promoter.
21. The nucleic acid construct of claim 19, wherein the plant promoter is a lettuce promoter.
22. The nucleic acid construct of claim 18, wherein the promoter is a constitutive promoter.
23. The nucleic acid construct of claim 18, wherein the promoter is an inducible promoter.
24. The nucleic acid construct of claim 18, wherein the promoter is a tissue-specific promoter.
25. A nucleic acid construct comprising a promoter sequence from an RG gene linked to a heterologous polynucleotide.
26. A transgenic plant comprising a recombinant expression cassette comprising a promoter operably linked to an RG polynucleotide.

27. The transgenic plant of claim 26, wherein the plant promoter is a plant promoter.
28. The transgenic plant of claim 26, wherein the plant promoter is a viral promoter.
- 5 29. The transgenic plant of claim 26, wherein the plant promoter is a heterologous promoter.
30. The transgenic plant of claim 26, wherein the plant is lettuce.
- 10 31. The transgenic plant of claim 26, wherein the RG polynucleotide is selected from the group consisting of SEQ ID NO:1 (RG1A), SEQ ID NO:2 (RG1B), SEQ ID NO: 3 (RG1C), SEQ ID NO:4 (RG1D), SEQ ID NO:5 (RG1E), SEQ ID NO:6 (RG1F), SEQ ID NO:7 (RG1G), SEQ ID NO:8 (RG1H), SEQ ID NO:9 (RG1I), and SEQ ID NO:10 (RG1J).
- 15 32. The transgenic plant of claim 26, wherein the RG polynucleotide is selected from the group consisting of SEQ ID NO:21 (RG2A); SEQ ID NO:23 (RG2B); SEQ ID NO:25 (RG2C); SEQ ID NO:27 (RG2D); SEQ ID NO:29 (RG2E); SEQ ID NO:31 (RG2F); SEQ ID NO:33 (RG2G); SEQ ID NO:35 (RG2H); SEQ ID NO:37 (RG2I); SEQ ID NO:39 (RG2J); SEQ ID NO:41 (RG2K); SEQ ID NO:43 (RG2L); SEQ ID NO:45 (RG2M); SEQ ID NO:87 (RG2A); SEQ ID NO:89 (RG2B); SEQ ID NO:91 (RG2C); SEQ ID NO:93 (RG2D) and SEQ ID NO:94 (RG2D); SEQ ID NO:96 (RG2E); SEQ ID NO:98 (RG2F); SEQ ID NO:100 (RG2G); SEQ ID NO:102 (RG2H); SEQ ID NO:104 (RG2I); SEQ ID NO:106 (RG2J) and SEQ ID NO:107 (RG2J); SEQ ID NO:109 (RG2K) and (SEQ ID NO:110 (RG2K); SEQ ID NO:112 (RG2L); SEQ ID NO:114 (RG2M); SEQ ID NO:116 (RG2N); SEQ ID NO:118 (RG2O); SEQ ID NO:120 (RG2P); SEQ ID NO:122 (RG2Q); SEQ ID NO:124 (RG2S); SEQ ID NO:126 (RG2T); SEQ ID NO:128 (RG2U); SEQ ID NO:130 (RG2V); and, SEQ ID NO:132 (RG2W).
- 20 33. The transgenic plant of claim 26, wherein the RG polynucleotide is selected from the group consisting of SEQ ID NO:68 (RG3) and SEQ ID NO:69 (RG4).
- 25 34. The transgenic plant of claim 26, wherein the RG polynucleotide is selected from the group consisting of SEQ ID NO:70 (RG5) and SEQ ID NO:71 (RG6).
- 30 35. The transgenic plant of claim 26, wherein the RG polynucleotide is selected from the group consisting of SEQ ID NO:72 (RG7) and SEQ ID NO:73 (RG8).

34. The transgenic plant of claim 26, wherein the RG polynucleotide comprises a sequence as set forth in SEQ ID NO:134 (RG5).

35. The transgenic plant of claim 26, wherein the RG polynucleotide comprises a
5 sequence as set forth in SEQ ID NO:136 (RG7).

36. The transgenic plant of claim 26, wherein the RG polynucleotide encodes an RG1 polypeptide selected from the group consisting of SEQ ID NO:11 (RG1A), SEQ ID NO:12 (RG1B), SEQ ID NO: 13 (RG1C), SEQ ID NO:14 (RG1D), SEQ ID NO:15 (RG1E), SEQ
10 ID NO:16 (RG1F), SEQ ID NO:17 (RG1G), SEQ ID NO:18 (RG1H), SEQ ID NO:19 (RG1I), and SEQ ID NO:20 (RG1J).

37. The transgenic plant of claim 26, wherein the RG polynucleotide encodes an RG2 polypeptide selected from the group consisting of SEQ ID NO:22 and SEQ ID NO:41
15 (RG2A); SEQ ID NO:24 and SEQ ID NO:42 (RG2B); SEQ ID NO:43 (RG2C); SEQ ID NO:44 (RG2D); SEQ ID NO:45 (RG2E); SEQ ID NO:46 (RG2F); SEQ ID NO:47 (RG2G); SEQ ID NO:48 (RG2H); SEQ ID NO:49 (RG2I); SEQ ID NO:50 (RG2J); SEQ ID NO:51 (RG2K); SEQ ID NO:52 (RG2L); SEQ ID NO:53 (RG2M); SEQ ID NO:88 (RG2A); SEQ ID NO:90 (RG2B); SEQ ID NO:92 (RG2C); SEQ ID NO:95 (RG2D); SEQ
20 ID NO:97 (RG2E); SEQ ID NO:99 (RG2F); SEQ ID NO:101 (RG2G); SEQ ID NO:103 (RG2H); SEQ ID NO:105 (RG2I); SEQ ID NO:108 (RG2J); SEQ ID NO:111 (RG2K); SEQ ID NO:113 (RG2L); SEQ ID NO:115 (RG2M); SEQ ID NO:117 (RG2N); SEQ ID NO:119 (RG2O); SEQ ID NO:121 (RG2P); SEQ ID NO:123 (RG2Q); SEQ ID NO:125 (RG2S); SEQ ID NO:127 (RG2T); SEQ ID NO:129 (RG2U); SEQ ID NO:131 (RG2V); and, SEQ ID
25 NO:133 (RG2W).

38. The transgenic plant of claim 26, wherein the RG polynucleotide encodes an RG3 polypeptide with a sequence as set forth by SEQ ID NO:138.

30 39. The transgenic plant of claim 26, wherein the RG polynucleotide encodes an RG4 polypeptide with a sequence as set forth by SEQ ID NO:139.

40. The transgenic plant of claim 26, wherein the RG polynucleotide encodes an RG5 polypeptide with a sequence as set forth by SEQ ID NO:135.

5 41. A method of enhancing disease resistance in a plant, the method comprising introducing into the plant a recombinant expression cassette comprising a promoter functional in the plant and operably linked to an RG polynucleotide sequence.

42. The method of claim 41, wherein the plant is a lettuce plant.

10 43. The method of claim 41, wherein the RG polynucleotide encodes an RG polypeptide selected from the group consisting of SEQ ID NO:22 and SEQ ID NO:41 (RG2A); SEQ ID NO:24 and SEQ ID NO:42 (RG2B); SEQ ID NO:43 (RG2C); SEQ ID NO:44 (RG2D); SEQ ID NO:45 (RG2E); SEQ ID NO:46 (RG2F); SEQ ID NO:47 (RG2G); SEQ ID NO:48 (RG2H); SEQ ID NO:49 (RG2I); SEQ ID NO:50 (RG2J); SEQ ID NO:51
15 (RG2K); SEQ ID NO:52 (RG2L); SEQ ID NO:53 (RG2M); SEQ ID NO:88 (RG2A); SEQ ID NO:90 (RG2B); SEQ ID NO:92 (RG2C); SEQ ID NO:95 (RG2D); SEQ ID NO:97 (RG2E); SEQ ID NO:99 (RG2F); SEQ ID NO:101 (RG2G); SEQ ID NO:103 (RG2H); SEQ ID NO:105 (RG2I); SEQ ID NO:108 (RG2J); SEQ ID NO:111 (RG2K); SEQ ID NO:113 (RG2L); SEQ ID NO:115 (RG2M); SEQ ID NO:117 (RG2N); SEQ ID NO:119 (RG2O);
20 SEQ ID NO:121 (RG2P); SEQ ID NO:123 (RG2Q); SEQ ID NO:125 (RG2S); SEQ ID NO:127 (RG2T); SEQ ID NO:129 (RG2U); SEQ ID NO:131 (RG2V); and, SEQ ID NO:133 (RG2W).

25 44. The method of claim 41, wherein the RG polynucleotide encodes an RG polypeptide selected from the group consisting of SEQ ID NO:138 (RG3); SEQ ID NO:139 (RG4); and SEQ ID NO:135 (RG5).

45. The method of claim 41, wherein the promoter is a tissue-specific promoter or a plant disease resistance promoter.

46. The method of claim 41, wherein the promoter is a constitutive promoter or an inducible promoter.

5 47. A method of detecting RG resistance genes in a nucleic acid sample, the method comprising:

contacting the nucleic acid sample with an RG polynucleotide to form a hybridization complex; and,

wherein the formation of the hybridization complex is used to detect the RG resistance gene in the nucleic acid sample.

10

48. The method of claim 47, wherein the RG polynucleotide is an RG1 polynucleotide.

49. The method of claim 47, wherein the RG polynucleotide is an RG2 polynucleotide.

15 50. The method of claim 47, wherein the RG polynucleotide is an RG3 polynucleotide, an RG4 polynucleotide, an RG5 polynucleotide or an RG7 polynucleotide.

51. The method of claim 47, wherein the RG resistance gene is amplified prior to the step of contacting the nucleic acid sample with the RG polynucleotide.

20

52. The method of claim 51, where the RG resistance gene is amplified by the polymerase chain reaction.

53. The method of claim 47, wherein the RG polynucleotide is labeled.

25

54. An RG polypeptide having at least 60% sequence identity to a polypeptide selected from the group consisting of: an RG1 polypeptide, an RG2 polypeptide, an RG3 polypeptide, an RG4 polypeptide, an RG5 polypeptide, and an RG7 polypeptide.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/00615

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.

US CL : 435/6, 91.2, 418, 419; 530/350; 536/23.1, 23.6, 24.1; 800/205

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/6, 91.2, 418, 419; 530/350; 536/23.1, 23.6, 24.1; 800/205

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PARAN et al. Development of Reliable PCR-Based Markers Linked to Downy Mildew Resistance Genes in Lettuce. Theor. Appl. Genet. 1993. Vol. 85, No. 8, pages 985-993, see entire article.	1-6, 8, 10, 12, 14, 16, 18-30, 41-42, 45-54
Y	KESSELI et al. Analysis of a Detailed Genetic Linkage Map of Lactuca sativa (Lettuce) Constructed From RFLP and RAPD Markers. Genetics. April 1994. Vol. 136, No. 4, pages 1435-1446, see entire document.	1-6, 8, 10, 12, 14, 16, 18-30, 41-42, 45-54
Y	MICHELMORE, RW. Isolation of Disease Resistance Genes from Crop Plants. Current Opinion in Biotechnology. 1995. Vol. 6, No. 2, pages 145-152, see entire document.	1-6, 8, 10, 12, 14, 16, 18-30, 41-42, 45-54

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:		*T*	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A	document defining the general state of the art which is not considered to be of particular relevance	*X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
B	earlier document published on or after the international filing date	*Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*A*	document member of the same patent family
O	document referring to an oral disclosure, use, exhibition or other means		
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Date of the actual completion of the international search

14 MARCH 1998

Date of mailing of the international search report

13 APR 1998

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/00615

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PARAN et al. Recent Amplification of Triose Phosphate Isomerase Related Sequences in Lettuce. Genome. 1992. Vol. 35, No. 4, pages 627-635, see entire document.	1-6, 8, 10, 12, 14, 16, 18-30, 41-42, 45-54
Y	PARAN et al. Identification of Restriction Fragment Length Polymorphism and Random Amplified Polymorphic DNA markers linked to Downy Mildew Resistance Genes in Lettuce, Using Near-Isogenic Lines. Genome. 1991. Vol. 34, No. 6, pages 1021-1027, see entire document.	1-6, 8, 10, 12, 14, 16, 18-30, 41-42, 45-54

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US98/00615

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☒ Claims Nos.: 7, 9, 11, 13, 15, 17, 31-40, 43-44
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

these claims are drawn to numerous sequences identified by SEQ ID NOs. However, since no computer readable form was submitted, no meaningful search could be carried out.

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐
☐

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/00615

A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

A01H 1/00; C07H 21/04; C07K 14/00; C12N 5/04, 5/10; C12P 19/34; C12Q 1/68

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